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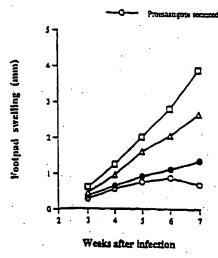
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(54) Title: LEISHMANIA ANTIGENS FOR USE IN THE THERAPY AND DIAGNOSIS OF LEISHMANIASIS

#### (57) Abstract

Compositions and methods for preventing, treating and detecting leishmaniasis and stimulating immune responses in patients are disclosed. The compounds provided include polypeptides that contain at least an immunogenic portion of one or more *Leishmania* antigens, or a variant thereof. Vaccines and pharmaceutical compositions comprising such polypeptides, or DNA molecules encoding such polypeptides, are also provided and may be used, for example, for the prevention and therapy of leishmaniasis, as well as for the detection of *Leishmania* infection.

Protection against infection with L. major in BALB/c mice immunized leishmanial antigens.



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# LEISHMANIA ANTIGENS FOR USE IN THE THERAPY AND DIAGNOSIS OF LEISHMANIASIS

## REFERENCE TO RELATED APPLICATIONS

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### TECHNICAL FIELD

The present invention relates generally to compositions and methods for preventing, treating and detecting leishmaniasis, and for stimulating immune responses in patients. The invention is more particularly related to polypeptides comprising an immunogenic portion of a *Leishmania* antigen or a variant thereof, and to vaccines and pharmaceutical compositions comprising one or more such polypeptides. The vaccines and pharmaceutical compositions may be used, for example, for the prevention and therapy of leishmaniasis, as well as for the detection of *Leishmania* infection.

## 15 BACKGROUND OF THE INVENTION

Leishmania organisms are intracellular protozoan parasites of macrophages that cause a wide range of clinical diseases in humans and domestic animals, primarily dogs. In some infections, the parasite may lie dormant for many years. In other cases, the host may develop one of a variety of forms of leishmaniasis. For example, the disease may be asymptomatic or may be manifested as subclinical visceral leishmaniasis, which is characterized by mild symptoms of malaise, diarrhea and intermittent hepatomegaly. Patients with subclinical or asymptomatic disease usually have low antibody titers, making the disease difficult to detect with standard techniques. Alternatively, leishmaniasis may be manifested as a cutaneous disease, which is a severe medical problem but is generally self-limiting, or as a highly destructive mucosal disease, which is not self-limiting. Finally, and most seriously, the disease may be manifested as an acute visceral infection involving the spleen, liver and lymph nodes, which, untreated, is generally a fatal disease. Symptoms of acute visceral leishmaniasis include hepatosplenomegaly, fever. leukopenia, anemia and hypergammaglobulinemia.

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Leishmaniasis is a serious problem in much of the world, including Brazil, China, East Africa, India and areas of the Middle East. The disease is also endemic in the Mediterranean region, including southern France, Italy, Greece, Spain, Portugal and North Africa. The number of cases of leishmaniasis has increased dramatically in the last 20 years, and millions of cases of this disease now exist worldwide. About 2 million new cases are diagnosed each year, 25% of which are visceral leishmaniasis. There are, however, no vaccines or effective treatments currently available.

Accurate diagnosis of leishmaniasis is frequently difficult to achieve. There are 20 species of Leishmania that infect humans, including L. donovani, L. chagasi, L. infantum, L. major, L. amazonensis, L. braziliensis, L. panamensis, L. mexicana, L. tropica, and L. guyanensis, and there are no distinctive signs or symptoms that unambiguously indicate the presence of Leishmania infection. Parasite detection methods have been used, but such methods are neither sensitive nor clinically practical. Current skin tests typically use whole or lysed parasites. Such tests are generally insensitive, irreproducible and prone to cross-reaction with a variety of other diseases. In addition, the preparations employed in such tests are often unstable. Thus, there is a need for improved methods for the detection of Leishmania infection.

Current experimental vaccines consisting of whole organisms have not proven effective in humans. Accordingly, there remains a need in the art for vaccines to prevent leishmaniasis in humans and dogs, and for improved therapeutic compositions for the treatment of leishmaniasis.

### SUMMARY OF THE INVENTION

Briefly stated, the present invention provides compositions and methods for preventing, treating and detecting leishmaniasis, as well as for stimulating immune responses in patients. In one aspect, polypeptides are provided which comprise at least an immunogenic portion of a *Leishmania* antigen, or a variant of such an antigen that differs only in conservative substitutions and/or modifications. In specific embodiments of the invention, the *Leishmania* antigen comprises an amino acid

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sequence selected from the group consisting of SEQ ID Nos: 2, 4, 20, 22, 24, 26, 36-38, 41, 50-53 and 82. DNA sequences encoding the above polypeptides, recombinant expression vectors comprising these DNA sequences and host cells transformed or transfected with such expression vectors are also provided.

In related aspects, the present invention provides pharmaceutical compositions which comprise one or more of the polypeptides described herein, or a DNA molecule encoding such polypeptides, and a physiologically acceptable carrier. Vaccines which comprise one or more such polypeptides or DNA molecules, together with a non-specific immune response enhancer are also provided. In specific embodiments of these aspects, the *Leishmania* antigen has an amino acid sequence selected from the group consisting of SEQ ID Nos: 2, 4, 20, 22, 24, 26, 36-38, 41, 50-53 and 82.

In still further related embodiments, the pharmaceutical compositions and vaccines comprise at least two different polypeptides, each polypeptide comprising an immunogenic portion of a *Leishmania* antigen having an amino acid sequence selected from the group consisting of sequences recited in SEQ ID Nos: 2, 4, 6, 8, 10, 20, 22, 24, 26, 36-38, 41, 50-53, 82, and variants thereof that differ only in conservative substitutions and/or modifications. In other embodiments, the inventive pharmaceutical compositions comprise one or more of the inventive polypeptides in combination with a known *Leishmania* antigen.

In yet other related embodiments, the pharmaceutical compositions and vaccines comprise soluble *Leishmania* antigens.

In another aspect, the present invention provides methods for inducing protective immunity against leishmaniasis in a patient, comprising administering to a patient a pharmaceutical composition or vaccine as described above.

In further aspects, methods and diagnostic kits are provided for detecting Leishmania infection in a patient. The methods comprise: (a) contacting dermal cells of a patient with a pharmaceutical composition as described above; and (b) detecting an immune response on the patient's skin, therefrom detecting Leishmania infection in the patient. The diagnostic kits comprise: (a) a pharmaceutical composition as described

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above; and (b) an apparatus sufficient to contact the pharmaceutical composition with the dermal cells of a patient.

In further aspects, the present invention provides methods for stimulating a cellular and/or humoral immune response in a patient, comprising administering to a patient a pharmaceutical composition or vaccine as described above.

In a related aspect, methods are provided for treating a patient afflicted with a disease responsive to IL-12 stimulation, comprising administering to a patient a pharmaceutical composition or vaccine as described above.

These and other aspects of the present invention will become apparent upon reference to the following detailed description and attached drawings. All references disclosed herein are hereby incorporated by reference in their entirety as if each was incorporated individually.

#### BRIEF DESCRIPTION OF THE DRAWINGS

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Figure 1 shows the stimulation of proliferation of T-cells obtained from L. donovani-immunized BALB/c mice (represented by stimulation index) by L. donovani-infected macrophages after incubation for 24, 48 and 72 hours.

Figure 2 illustrates representative HPLC profiles of peptides isolated from MHC class II molecules of P388D1 macrophages. Panel A shows peptides isolated from uninfected macrophages and panel B shows peptides isolated from L. donovani infected macrophages. The arrows in panel B indicate peptide peaks present only in the infected macrophage preparation.

Figure 3 illustrates the expression and purification of the *Leishmania* antigen Ldp23 as a recombinant fusion protein. Panel A shows a Coomassie blue-stained SDS-PAGE gel of lysed *E. coli* without (lane 1) and with (lane 2) IPTG induction of Ldp23 expression. Arrow indicates the recombinant fusion protein. Panel B shows the fusion protein following excision from a preparative SDS-PAGE gel, electroelution, dialysis against PBS and analytical SDS-PAGE.

Figure 4 presents a Northern blot analysis of total RNA prepared from L.

donovani, L. major, L. amazonensis and L. pifanoi with a <sup>32</sup>P labeled Ldp23 gene. 1, 2

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and 3 refer to RNA obtained from promastigotes at the logarithmic growth phase, promastigotes at the stationary growth phase and amastigote forms, respectively.

Figure 5 shows a Western blot analysis of *L. donovani* promastigote antigens incubated with pre-immune rabbit serum (lane A) or with anti-Ldp23 rabbit antiserum (lane B).

Figure 6 illustrates the surface expression of Ldp23 on live *L. donovani* promastigotes. The dotted line shows the indirect immunofluorescence performed using pre-immune mouse serum and the solid line shows the result obtained with mouse anti-GST-Ldp23 antiserum. Fluorescence intensity was analyzed by FACScan.

Figure 7 shows the stimulation of Leishmania-specific T-cell proliferation by Ldp23. The results are presented as relative cell number as a function of fluorescence intensity. T-cells (10<sup>5</sup>/well) were purified from lymph nodes of BALB/c mice immunized in the foot pad with L. donovani promastigotes in CFA and were cultured with various concentrations of the purified recombinant Ldp23 in the presence of 2 x 10<sup>5</sup> Mitomycin C-treated normal BALB/c spleen mononuclear cells. Proliferation of T-cells was measured at 27 hours of culture. Values are expressed as cpm and represent the mean of [3H]TdR incorporation of triplicate cultures.

Figure 8 illustrates Ldp23-induced cytokine production by lymph node cells of BALB/c mice. Cultures were incubated with varying amounts of Ldp23 or *Leishmania* lysate, presented as μg/mL, and were assayed by ELISA for the production of interferon-γ (panel A) or interleukin-4 (panel B), both of which are shown as ng/mL.

Figure 9 shows the PCR amplification of cytokine mRNAs isolated from mucosal leishmaniasis (Panel A) and cutaneous leishmaniasis (panel B) patient PBMC before and after stimulation with representative polypeptides of the present invention. Lanes O and - indicate the level of PCR products at the initiation of culture and after 72 hours of culture, respectively, in the absence of added polypeptide; lanes Lb, 83a and 83b indicate the level of PCR products following culturing of PBMC with L. braziliensis lysate, and the Leishmania antigens Lbhsp83a and Lbhsp83b, respectively.

Figure 10 presents a comparison of the levels of interferon- $\gamma$  (panel A) and TNF- $\alpha$  (panel B) in the supernatants of 72 hour PBMC cultures from *Leishmania*-

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infected and control individuals in response to stimulation with parasite lysate or the indicated polypeptides.

Figure 11 illustrates the levels of IL-10 p40 (in pg/mL) in the supernatant of PBMC cultures from *L. braziliensis*-infected individuals and uninfected controls 72 hours following stimulation with parasite promastigate lysate (Lb), Lbhsp83a or Lbhsp83b.

Figure 12 presents the reactivities of sera from *L. braziliensis* infected-patients with representative polypeptides of the present invention in a standard ELISA. Values are expressed as absorbance at 405 nm.

Figures 13A and 13B illustrate the level of secreted IL-4 and IFN-γ (in pg/mL) stimulated in mouse lymph node cultures by the addition of representative polypeptides of the present invention.

Figure 14 shows the level of IFN-γ (in pg/mL) secreted by Leishmania-infected and uninfected human PBMC stimulated by the Leishmania antigen M15, as compared to the levels stimulated by L. major lysate and L-Rack, an antigen that does not appear to be recognized by Leishmania-infected humans.

Figure 15 shows the level of IFN-γ (in pg/mL) secreted by infected and uninfected human PBMC stimulated by soluble *Leishmania* antigens (S antigens), as compared to the levels stimulated by *L. major* lysate and L-Rack.

Figure 16 illustrates the proliferation of murine lymph node cultures stimulated by the addition of representative polypeptides of the present invention. Values are expressed as cpm.

Figure 17 shows the proliferation of human PBMC, prepared from Leishmania-immune and uninfected individuals, stimulated by M15 as compared to the proliferation stimulated by L. major lysate and L-Rack. Values are expressed as cpm.

Figure 18 illustrates the proliferation of human PBMC, prepared from *Leishmania*-infected and uninfected individuals, stimulated by soluble *Leishmania* antigens as compared to the proliferation stimulated by culture medium, *L. major* lysate and L-Rack. Values are expressed as cpm.

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Figure 19 presents a comparison of a Lbhsp83 sequence (SEQ ID NO:6) with homologous sequences from *L. amazonensis* (Lahsp83) (SEQ ID NO:16), *T. cruzi* (Tchsp83) (SEQ ID NO:17) and humans (Huhsp89) (SEQ ID NO:18).

Figure 20 illustrates the reactivity of rabbit sera raised against soluble Leishmania antigens with Leishmania promastigote lysate (lane 1) and soluble Leishmania antigens (lane 2).

Figure 21 shows the cDNA and predicted amino acid sequence for the Leishmania antigen Lmspla.

Figure 22 shows a Southern blot of genomic DNA from *L. major* digested with a panel of restriction enzymes (lanes 1 to 7) and six other *Leishmania* species digested with PstI (lanes 8 to 13) probed with the full-length cDNA insert of Lmsp1a.

Figure 23 shows a Southern blot of genomic DNA from *L. major* digested with a panel of restriction enzymes, six other *Leishmania* species digested with PstI and the infectious pathogens *T. cruzi* and *T. brucei*, probed with the full-length cDNA insert of the *Leishmania* antigen MAPS-1A.

Figure 24 illustrates the proliferation of PBMC isolated from uninfected-individuals, patients with active mucosal leishmaniasis and patients post kala-azar infection, stimulated by MAPS-1A.

Figure 25 illustrates the proliferation of murine lymph node cultures stimulated by MAPS-1A.

Figure 26 illustrates the reactivity of MAPS-1A with sera from human leishmaniasis patients.

Figure 27 illustrates the reactivity of MAPS-1A with sera from mice immunized against and/or infected with leishmaniasis.

Figure 28 illustrates the effectiveness of immunization with either soluble Leishmania antigens or a mixture of Ldp23, LbeiF4A and M15 plus adjuvant in conferring protection against infection (as measured by footpad swelling) in a murine leishmaniasis model system, as compared to the administration of adjuvant alone.

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Figure 29 illustrates the effectiveness of immunization with MAPS-1A plus adjuvant in conferring protection against infection (as measured by footpad swelling) in a murine leishmaniasis model system, as compared to the administration of adjuvant alone.

Figures 30A and B illustrate the proliferation of murine lymph node cultures stimulated with either LcgSP8, LcgSP10 or LcgSP3.

## DETAILED DESCRIPTION OF THE INVENTION

As noted above, the present invention is generally directed to compositions and methods for preventing, treating and detecting leishmaniasis, as well as for stimulating immune responses in patients. The compositions of the subject invention include polypeptides that comprise at least an immunogenic portion of a Leishmania antigen, or a variant of such an antigen that differs only in conservative substitutions and/or modifications. In one preferred embodiment, compositions of the present invention include multiple polypeptides selected so as to provide enhanced protection against a variety of Leishmania species.

Polypeptides within the scope of the present invention include, but are not limited to, polypeptides comprising immunogenic portions of *Leishmania* antigens comprising the sequences recited in SEQ ID NO:2 (referred to herein as M15), SEQ ID NO:4 (referred to herein as Ldp23), SEQ ID NO:6 (referred to herein as Lbhsp83), SEQ ID NO:8 (referred to herein as Lt-210), SEQ ID NO:10 (referred to herein as LbeIF4A), SEQ ID NO: 20 (referred to herein as Lmsp1a), SEQ ID NO: 22 (referred to herein as Lmsp9a), SEQ ID NOs: 24 and 26 (referred to herein as MAPS-1A), and SEQ ID NO: 36-42, 49-53 and 55. As used herein, the term "polypeptide" encompasses amino acid chains of any length, including full length proteins (*i.e.*, antigens), wherein the amino acid residues are linked by covalent bonds. Thus, a polypeptide comprising an immunogenic portion of one of the above antigens may consist entirely of the immunogenic portion, or may contain additional sequences. The additional sequences may be derived from the native *Leishmania* antigen or may be heterologous, and such sequences may (but need not) be immunogenic. An antigen "having" a particular

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sequence is an antigen that contains, within its full length sequence, the recited sequence. The native antigen may, or may not, contain additional amino acid sequence.

An immunogenic portion of a *Leishmania* antigen is a portion that is capable of eliciting an immune response (*i.e.*, cellular and/or humoral) in a presently or previously *Leishmania*-infected patient (such as a human or a dog) and/or in cultures of lymph node cells or peripheral blood mononuclear cells (PBMC) isolated from presently or previously *Leishmania*-infected individuals. The cells in which a response is elicited may comprise a mixture of cell types or may contain isolated component cells (including, but not limited to, T-cells, NK cells, macrophages, monocytes and/or B cells). In particular, immunogenic portions are capable of inducing T-cell proliferation and/or a dominantly Th1-type cytokine response (*e.g.*, IL-2, IFN-γ, and/or TNF-α production by T-cells and/or NK cells; and/or IL-12 production by monocytes, macrophages and/or B cells). Immunogenic portions of the antigens described herein may generally be identified using techniques known to those of ordinary skill in the art, including the representative methods provided herein.

The compositions and methods of the present invention also encompass variants of the above polypeptides. A polypeptide "variant," as used herein, is a polypeptide that differs from the native antigen only in conservative substitutions and/or modifications, such that the ability of the polypeptide to include an immune response is retained. Polypeptide variants preferably exhibit at least about 70%, more preferably at least about 90% and most preferably at least about 95% identity to the identified polypeptides. Alternatively, such variants may be identified by modifying one of the above polypeptide sequences and evaluating the immunogenic properties of the modified polypeptide using, for example, the representative procedures described herein.

A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide chemistry would expect the secondary structure and hydropathic nature of the polypeptide to be substantially unchanged. In general, the following groups of amino acids represent conservative changes: (1) ala, pro, gly, glu, asp, gln,

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asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe; (4) lys, arg, his; and (5) phe, tyr, trp, his.

Variants may also (or alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the immunogenic properties, secondary structure and hydropathic nature of the polypeptide. For example, a polypeptide may be conjugated to a signal (or leader) sequence at the N-terminal end of the protein which co-translationally or post-translationally directs transfer of the protein. The polypeptide may also be conjugated to a linker or other sequence for ease of synthesis, purification or identification of the polypeptide (e.g., poly-His), or to enhance binding of the polypeptide to a solid support. For example, a polypeptide may be conjugated to an immunoglobulin Fc region.

A nucleotide "variant" is a sequence that differs from the recited nucleotide sequence in having one or more nucleotide deletions, substitutions or additions. Such modifications may be readily introduced using standard mutagenesis techniques, such as oligonucleotide-directed site-specific mutagenesis as taught, for example, by Adelman et al. (*DNA*, 2:183, 1983). Nucleotide variants may be naturally occurring allelic variants, or non-naturally occurring variants. Variant nucleotide sequences preferably exhibit at least about 70%, more preferably at least about 80% and most preferably at least about 90% identity to the recited sequence. Such variant nucleotide sequences will generally hybridize to the recited nucleotide sequence under stringent conditions. As used herein, "stringent conditions" refers to prewashing in a solution of 6X SSC, 0.2% SDS; hybridizing at 65 °C, 6X SSC, 0.2% SDS overnight; followed by two washes of 30 minutes each in 1X SSC, 0.1% SDS at 65 °C and two washes of 30 minutes each in 0.2X SSC, 0.1% SDS at 65 °C.

"Polypeptides" as described herein also include combination polypeptides. A "combination polypeptide" is a polypeptide comprising at least one of the above immunogenic portions and one or more additional immunogenic *Leishmania* sequences, which are joined via a peptide linkage into a single amino acid chain. The sequences may be joined directly (*i.e.*, with no intervening amino acids) or may be

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joined by way of a linker sequence (e.g., Gly-Cys-Gly) that does not significantly diminish the immunogenic properties of the component polypeptides.

In general, Leishmania antigens having immunogenic properties, and DNA sequences encoding such antigens, may be prepared using any of a variety of procedures from one or more Leishmania species including, but not limited to, L. donovani, L. chagasi, L. infantum, L. major, L. amazonensis, L. braziliensis, L. panamensis, L. mexicana, L. tropica, and L. guyanensis. Such species are available, for example, from the American Type Culture Collection (ATCC), Rockville, MD. For example, peptides isolated from MHC class II molecules of macrophages infected with a Leishmania species may be used to rescue the corresponding Leishmania donor antigens. MHC class II molecules are expressed mainly by cells of the immune system, including macrophages. These molecules present peptides, which are usually 13-17 amino acids long, derived from foreign antigens that are degraded in cellular vesicles. The bound peptide antigens are then recognized by CD4 T-cells. Accordingly, foreign peptides isolated from MHC class II molecules of, for example, Leishmania-infected murine macrophages may be used to identify immunogenic Leishmania proteins.

Briefly, peptides derived from Leishmania antigens may be isolated by comparing the reverse phase HPLC profile of peptides extracted from infected macrophages with the profile of peptides extracted from uninfected cells. Peptides giving rise to distinct HPLC peaks unique to infected macrophages may then be sequenced using, for example, Edman chemistry as described in Edman and Berg, Eur J. Biochem, 80:116-132 (1967). A DNA fragment corresponding to a portion of a Leishmania gene encoding the peptide may then be amplified from a Leishmania cDNA library using an oligonucleotide sense primer derived from the peptide sequence and an oligo dT antisense primer. The resulting DNA fragment may then be used as a probe to screen a Leishmania library for a full length cDNA or genomic clone that encodes the Leishmania antigen. Such screens may generally be performed using techniques well known to those of ordinary skill in the art, such as those described in Sambrook et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratories, Cold Spring Harbor, NY (1989).

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This approach may be used to identify a 23 kD Leishmania donovani antigen (referred to herein as Ldp23). The sequence of a DNA molecule encoding Ldp23 is provided in SEQ ID NO:3 and the amino acid sequence of Ldp23 is provided in SEQ ID NO:4. Using the methods described herein, Ldp23 has been shown to induce a Th1 immune response in T-cells prepared from Leishmania-infected mice.

Alternatively, a *Leishmania* cDNA or genomic expression library may be screened with serum from a *Leishmania*-infected individual, using techniques well known to those of ordinary skill in the art. DNA molecules encoding reactive antigens may then be used to express the recombinant antigen for purification. The immunogenic properties of the purified *Leishmania* antigens may then be evaluated using, for example the representative methods described herein.

For example, sera from Leishmania-infected mice may be used to screen a cDNA library prepared from Leishmania amastigotes. Reactive clones may then be expressed and recombinant proteins assayed for the ability to stimulate T-cells or NK cells derived from Leishmania-immune individuals (i.e., individuals having evidence of infection, as documented by positive serological reactivity with Leishmania-specific antibodies and/or a Leishmania-specific DTH response, without clinical symptoms of leishmaniasis). This procedure may be used to obtain a recombinant DNA molecule encoding the Leishmania antigen designated M15. The sequence of such a DNA molecule is provided in SEQ ID NO:1, and the amino acid sequence of the encoded protein is provided in SEQ ID NO:2.

A similar approach may be used to isolate a genomic DNA molecule encoding an immunogenic *Leishmania braziliensis* antigen, referred to herein as Lbhsp83. More specifically, a genomic clone encoding Lbhsp83 may be isolated by screening a *L. braziliensis* expression library with sera from a *Leishmania*-infected individual. The DNA encoding Lbhsp83 is homologous to the gene encoding the eukaryotic 83 kD heat shock protein. The sequence of a DNA molecule encoding nearly all of Lbhsp83 is presented in SEQ ID NO:5, and the encoded amino acid sequence is provided in SEQ ID NO:6. Using the methods described below, Lbhsp83 has been found to stimulate proliferation, and a mixed Th1 and Th2 cytokine profile, in

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PBMC isolated from *L. braziliensis*-infected patients. Accordingly, Lbhsp83 is an immunogenic *Leishmania* antigen. Regions of Lbhsp83 that are not conserved with the mammalian gene have been found to be particularly potent for T-cell stimulation and antibody binding. Such regions may be identified, for example, by visual inspection of the sequence comparison provided in Figure 19.

This approach may also be used to isolate a DNA molecule encoding a 210 kD immunogenic *L. tropica* antigen, referred to herein as Lt-210. The preparation and characterization of Lt-210, and immunogenic portions thereof (such as Lt-1 and immunogenic repeat and non-repeat sequences), is described in detail in U.S. Patent Application Serial No. 08/511,872, filed August 4, 1995. The sequence of a DNA molecule encoding Lt-1 is provided in SEQ ID NO:7 and the encoded amino acid sequence is presented in SEQ ID NO:8.

The above approach may further be used to isolate a DNA molecule encoding a *L. braziliensis* antigen referred to herein as LbeIF4A. Briefly, such a clone may be isolated by screening a *L. braziliensis* expression library with sera obtained from a patient afflicted with mucosal leishmaniasis, and analyzing the reactive antigens for the ability to stimulate proliferative responses and preferential Th1 cytokine production in PBMC isolated from *Leishmania*-infected patients, as described below. The preparation and characterization of LbeIF4A is described in detail in U.S. Patent Application Serial Nos. 08/454,036 and 08/488,386, which are continuations-in-part of U.S. Patent Application Serial No. 08/232,534, filed April 22, 1994. The sequence of a DNA molecule encoding LbeIF4A is provided in SEQ ID NO:9 and the encoded amino acid sequence is presented in SEQ ID NO:10. Homologs of LbeIF4A, such as that found in *L. major*, may also be isolated using this approach, and are within the scope of the present invention.

Compositions of the present invention may also, or alternatively, contain soluble Leishmania antigens. As used herein, "soluble Leishmania antigens" refers to a mixture of at least 8 different Leishmania antigens that may be isolated from the supernatant of Leishmania promastigotes of any species grown for 8-12 hours in protein-free medium. Briefly, the organisms are grown to late log phase in complex

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medium with serum until they reach a density of 2-3 x 10<sup>7</sup> viable organisms per mL of medium. The organisms are thoroughly washed to remove medium components and resuspended at 2-3 x 10<sup>7</sup> viable organisms per mL of defined serum-free medium consisting of equal parts RPMI 1640 and medium 199, both from Gibco BRL, Gaithersburg, MD. After 8-12 hours, the supernatant containing soluble Leishmania antigens is removed, concentrated 10 fold and dialyzed against phosphate-buffered saline for 24 hours. The presence of at least eight different antigens within the mixture of Leishmania antigens may be confirmed using SDS-PAGE (i.e., through the observation of at least 8 different bands). The immunogenic properties of the soluble Leishmania antigens may be confirmed by evaluating the ability of the preparation to elicit an immune response in cultures of lymph node cells and/or peripheral blood mononuclear cells (PBMC) isolated from presently or previously Leishmania-infected individuals. Such an evaluation may be performed as described below.

Individual antigens present within the mixture of soluble Leishmania antigens may be isolated by immunizing mice or rabbits with Leishmania culture supernatant, containing soluble antigens, and employing the resultant sera to screen a Leishmania cDNA expression library as described in detail below. This procedure may be used to isolate recombinant DNA molecules encoding the L. major antigens referred to herein as Lmsp1a, Lmsp9a and MAPS-1A. DNA sequences encoding Lmsp1a, Lmsp9a and MAPS-1A are provided in SEQ ID NO: 19, 21 and 23, respectively, with the corresponding predicted amino acid sequences being presented in SEQ ID NO: 20, 22 and 24, respectively. Similarly, sera from mice or rabbits immunized with L. major culture supernatant may be used to screen an L. major genomic DNA library. As detailed below, this procedure may be used to isolate DNA molecules encoding the L. major antigens referred to herein as LmgSP1, LmgSP3, LmgSP5, LmgSP8, LmgSP9, LmgSP13, LmgSP19, and DNA molecules encoding the L. chagasi antigens LcgSP1, LcgSP3, LcgSP4, LcgSP8, and LcgSP10. The DNA sequences encoding these antigens are provided in SEQ ID NO:29-35 and 44-48, respectively, with the corresponding amino acid sequences being provided in SEQ ID NO: 36-42 and 49-53. The L. major antigens referred to herein as 1G6-34, 1E6-44, 4A5-63, 1B11-39, 2A10-37, 4G2-83,

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4H6-41 and 8G3-100 may be isolated by means of CD4+ T cell expression cloning as described below. DNA sequences encoding these antigens are provided in SEQ ID NO: 72-79, respectively, with the corresponding predicted amino acid sequences being provided in SEQ ID NO: 80-87. The immunogenic properties of the isolated *Leishmania* antigens may be evaluated using, for example, the representative methods described herein.

Regardless of the method of preparation, the antigens described herein are immunogenic. In other words, the antigens (and immunogenic portions thereof) are capable of eliciting an immune response in cultures of lymph node cells and/or peripheral blood mononuclear cells (PBMC) isolated from presently or previously Leishmania-infected individuals. More specifically, the antigens, and immunogenic portions thereof, have the ability to induce T-cell proliferation and/or to elicit a... dominantly Th1-type cytokine response (e.g., IL-2, IFN-γ, and/or TNF-α production by T-cells and/or NK cells; and/or IL-12 production by monocytes, macrophages and/or B cells) in cells isolated from presently or previously Leishmania-infected individuals: A Leishmania-infected individual may be afflicted with a form of leishmaniasis (such as subclinical, cutaneous, mucosal or active visceral) or may be asymptomatic. Such individuals may be identified using methods known to those of ordinary skill in the art. Individuals with leishmaniasis may be identified based on clinical findings associated with at least one of the following: isolation of parasite from lesions, a positive skin test with Leishmania lysate or a positive serological test. Asymptomatic individuals are infected individuals who have no signs or symptoms of the disease. Such individuals can be identified based on a positive serological test and/or skin test with Leishmania lysate.

The term "PBMC," which refers to a preparation of nucleated cells consisting primarily of lymphocytes and monocytes that are present in peripheral blood, encompasses both mixtures of cells and preparations of one or more purified cell types. PBMC may be isolated by methods known to those in the art. For example, PBMC may be isolated by density centrifugation through, for example, Ficoll<sup>TM</sup> (Winthrop Laboratories, New York). Lymph node cultures may generally be prepared by

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immunizing BALB/c mice (e.g., in the rear foot pad) with Leishmania promastigotes emulsified in complete Freund's adjuvant. The draining lymph nodes may be excised following immunization and T-cells may be purified in an anti-mouse Ig column to remove the B cells, followed by a passage through a Sephadex G10 column to remove the macrophages. Similarly, lymph node cells may be isolated from a human following biopsy or surgical removal of a lymph node.

The ability of a polypeptide (e.g., a Leishmania antigen or a portion or other variant thereof) to induce a response in PBMC or lymph node cell cultures may be evaluated by contacting the cells with the polypeptide and measuring a suitable response. In general, the amount of polypeptide that is sufficient for the evaluation of about 2 x 10<sup>5</sup> cells ranges from about 10 ng to about 100 µg, and preferably is about 1-10 µg. The incubation of polypeptide with cells is typically performed at 37°C for about 1-3 days. Following incubation with polypeptide, the cells are assayed for an appropriate response. If the response is a proliferative response, any of a variety of techniques well known to those of ordinary skill in the art may be employed. For example, the cells may be exposed to a pulse of radioactive thymidine and the incorporation of label into cellular DNA measured. In general, a polypeptide that results in at least a three fold increase in proliferation above background (i.e., the proliferation observed for cells cultured without polypeptide) is considered to be able to induce proliferation.

Alternatively, the response to be measured may be the secretion of one or more cytokines (such as interferon- $\gamma$  (IFN- $\gamma$ ), interleukin-4 (IL-4), interleukin-12 (p70 and/or p40), interleukin-2 (IL-2) and/or tumor necrosis factor- $\alpha$  (TNF- $\alpha$ )) or the change in the level of mRNA encoding one or more specific cytokines. In particular, the secretion of interferon- $\gamma$ , interleukin-2, tumor necrosis factor- $\alpha$  and/or interleukin-12 is indicative of a Th1 response, which is responsible for the protective effect against *Leishmania*. Assays for any of the above cytokines may generally be performed using methods known to those of ordinary skill in the art, such as an enzyme-linked immunosorbent assay (ELISA). Suitable antibodies for use in such assays may be obtained from a variety of sources such as Chemicon, Temucula, CA and PharMingen,

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San Diego, CA, and may generally be used according to the manufacturer's instructions. The level of mRNA encoding one or more specific cytokines may be evaluated by, for example, amplification by polymerase chain reaction (PCR). In general, a polypeptide that is able to induce, in a preparation of about 1-3 x 10<sup>5</sup> cells, the production of 30 pg/mL of IL-12, IL-4, IFN-γ, TNF-α or IL-12 p40, or 10 pg/mL of IL-12 p70, is considered able to stimulate production of a cytokine.

Immunogenic portions of the antigens described herein may be prepared and identified using well known techniques, such as those summarized in Paul, Fundamental Immunology, 3rd ed., 243-247 (Raven Press, 1993) and references cited therein. Such techniques include screening polypeptides derived from the native antigen for immunogenic properties using, for example, the representative techniques described herein. An immunogenic portion of a polypeptide is a portion that, within such representative assays, generates an immune response (e.g., proliferation and/or cytokine production) that is substantially similar to that generated by the full length antigen. In other words, an immunogenic portion of an antigen may generate at least about 25%, and preferably at least about 50%, of the response generated by the full length antigen in the model assays described herein.

Portions and other variants of immunogenic *Leishmania* antigens may be generated by synthetic or recombinant means. Synthetic polypeptides having fewer than about 100 amino acids, and generally fewer than about 50 amino acids, may be generated using techniques well known to those of ordinary skill in the art. For example, such polypeptides may be synthesized using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where amino acids are sequentially added to a growing amino acid chain. *See* Merrifield, *J. Am. Chem. Soc.* 85:2149-2146, 1963. Equipment for automated synthesis of polypeptides is commercially available from suppliers such as Perkin Elmer/Applied BioSystemsDivision, Foster City, CA, and may be operated according to the manufacturer's instructions.

Recombinant polypeptides containing portions and/or variants of a native antigen may be readily prepared from a DNA sequence encoding the antigen.

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For example, supernatants from suitable host/vector systems which secrete recombinant protein into culture media may be first concentrated using a commercially available filter. Following concentration, the concentrate may be applied to a suitable purification matrix such as an affinity matrix or an ion exchange resin. Finally, one or more reverse phase HPLC steps can be employed to further purify a recombinant protein.

In general, any of a variety of expression vectors known to those of ordinary skill in the art may be employed to express recombinant polypeptides of this invention. Expression may be achieved in any appropriate host cell that has been transformed or transfected with an expression vector containing a DNA molecule that encodes a recombinant polypeptide. Suitable host cells include prokaryotes, yeast and higher eukaryotic cells. Preferably, the host cells employed are *E. coli*, yeast or a mammalian cell line such as COS or CHO. The DNA sequences expressed in this manner may encode naturally occurring antigens, portions of naturally occurring antigens, or other variants thereof. For example, variants of a native antigen may generally be prepared using standard mutagenesis techniques, such as oligonucleotide-directed site-specific mutagenesis, and sections of the DNA sequence may be removed to permit preparation of truncated polypeptides.

In another aspect, the present invention provides epitope repeat sequences, or antigenic epitopes, of a *Leishmania* antigen, together with polypeptides comprising at least two such contiguous antigenic epitopes. As used herein an "epitope" is a portion of an antigen that reacts with sera from *Leishmania*-infected individuals (i.e. an epitope is specifically bound by one or more antibodies present in such sera). As discussed above, epitopes of the antigens described in the present application may be generally identified using techniques well known to those of skill in the art.

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In one embodiment, antigenic epitopes of the present invention comprise an amino acid sequence provided in SEQ ID NO:43, 56, 57 or 58. As discussed in more detail below, antigenic epitopes provided herein may be employed in the diagnosis and treatment of *Leishmania* infection, either alone or in combination with other

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Leishmania antigens or antigenic epitopes. Antigenic epitopes and polypeptides comprising such epitopes may be prepared by synthetic means, as described generally above and in detail in Example 15.

In certain aspects of the present invention, described in detail below, the polypeptides, antigenic epitopes and/or soluble *Leishmania* antigens may be incorporated into pharmaceutical compositions or vaccines. For clarity, the term "polypeptide" will be used when describing specific embodiments of the inventive therapeutic compositions and diagnostic methods. However, it will be clear to one of skill in the art that the antigenic epitopes of the present invention may also be employed in such compositions and methods.

Pharmaceutical compositions comprise one or more polypeptides, each of which may contain one or more of the above sequences (or variants thereof), and a physiologically acceptable carrier. Vaccines comprise one or more of the above polypeptides and a non-specific immune response enhancer, such as an adjuvant (e.g., LbeIF4A, interleukin-12 or other cytokines) or a liposome (into which the polypeptide is incorporated). Vaccines may additionally contain a delivery vehicle, such as a biodegradable microsphere (disclosed, for example, in U.S. Patent Nos. 4,897,268 and 5,075,109). Pharmaceutical compositions and vaccines within the scope of the present invention may also contain other *Leishmania* antigens, either incorporated into a combination polypeptide or present within one or more separate polypeptides.

DNA encoding one or more of the polypeptides as described above, such that the polypeptide is generated *in situ*. In such pharmaceutical compositions and vaccines, the DNA may be present within any of a variety of delivery systems known to those of ordinary skill in the art, including nucleic acid expression systems, bacteria and viral expression systems. Appropriate nucleic acid expression systems contain the necessary DNA sequences for expression in the patient (such as a suitable promoter and terminating signal). Bacterial delivery systems involve the administration of a bacterium (such as *Bacillus-Calmette-Guerrin*) that expresses an immunogenic portion of the polypeptide on its cell surface. In a preferred embodiment, the DNA may be

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introduced using a viral expression system (e.g., vaccinia or other pox virus, retrovirus, or adenovirus), which may involve the use of a non-pathogenic (defective), replication competent virus. Techniques for incorporating DNA into such expression systems are well known to those of ordinary skill in the art. The DNA may also be "naked," as described, for example, in Ulmer et al., *Science 259*:1745-1749 (1993) and reviewed by Cohen, *Science 259*:1691-1692 (1993). The uptake of naked DNA may be increased by coating the DNA onto biodegradable beads, which are efficiently transported into the cells.

While any suitable carrier known to those of ordinary skill in the art may be employed in the pharmaceutical compositions of this invention, the type of carrier will vary depending on the mode of administration. For parenteral administration, such as subcutaneous injection, the carrier preferably comprises water, saline, alcohol, a fat, a wax or a buffer. For oral administration, any of the above carriers or a solid carrier, such as mannitol, lactose, starch, magnesium stearate, sodium saccharine, talcum, cellulose, glucose, sucrose, and magnesium carbonate, may be employed. Biodegradable microspheres (e.g., polylactic galactide) may also be employed as carriers for the pharmaceutical compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Patent Nos. 4,897,268 and 5,075,109.

Any of a variety of adjuvants may be employed in the vaccines of this invention to nonspecifically enhance the immune response. Most adjuvants contain a substance designed to protect the antigen from rapid catabolism, such as aluminum hydroxide or mineral oil, and a nonspecific stimulator of immune responses, such as lipid A, *Bordella pertussis* or *Mycobacterium tuberculosis*. Suitable adjuvants are commercially available as, for example, Freund's Incomplete Adjuvant and Complete Adjuvant (Difco Laboratories, Detroit, MI), Merck Adjuvant 65 (Merck and Company, Inc., Rahway, NJ), alum, biodegradable microspheres, monophosphoryl lipid A and quil A. Preferred adjuvants include LbeIF4A, IL-12 and other cytokines such as IFN-γ or granulocyte-macrophage colony stimulating factor (GM-CSF). By virtue of its ability to induce an exclusive Th1 immune response, the use of LbeIF4A, and variants thereof, as an adjuvant in the vaccines of the present invention is particularly preferred.

In one preferred embodiment, compositions of the present invention include multiple polypeptides selected so as to provide enhanced protection against a variety of Leishmania species. Such polypeptides may be selected based on the species of origin of the native antigen or based on a high degree of conservation of amino acid sequence among different species of Leishmania. A combination of individual polypeptides may be particularly effective as a prophylactic and/or therapeutic vaccine because (1) stimulation of proliferation and/or cytokine production by individual polypcptides may be additive, (2) stimulation of proliferation and/or cytokine production by individual polypeptides may be synergistic, (3) individual polypeptides may stimulate cytokine profiles in such a way as to be complementary to each other and/or (4) individual polypeptides may be complementary to one another when certain of them are expressed more abundantly on the individual species or strain of Leishmania responsible for infection. A preferred combination contains polypeptides that comprise immunogenic portions of M15, Ldp23, Lbhsp83, Lt-1 and LbeIF4A. Alternatively, or in addition, the combination may include one or more polypeptides comprising immunogenic portions of other Leishmania antigens disclosed herein, and/or soluble Leishmania antigens.

The above pharmaceutical compositions and vaccines may be used, for a example, to induce protective immunity against *Leishmania* in a patient, such as a human or a dog, to prevent leishmaniasis. Appropriate doses and methods of administration for this purposes are described in detail below.

The pharmaceutical compositions and vaccines described herein may also be used to stimulate an immune response, which may be cellular and/or humoral, in a patient. For Leishmania-infected patients, the immune responses that may be generated include a preferential Th1 immune response (i.e., a response characterized by the production of the cytokines interleukin-1, interleukin-2, interleukin-12 and/or interferon- $\gamma$ , as well as tumor necrosis factor- $\alpha$ ). For uninfected patients, the immune response may be the production of interleukin-12 and/or interleukin-2, or the stimulation of gamma delta T-cells. In either category of patient, the response stimulated may include IL-12 production. Such responses may also be elicited in

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biological samples of PBMC or components thereof derived from *Leishmania*-infected or uninfected individuals. As noted above, assays for any of the above cytokines may generally be performed using methods known to those of ordinary skill in the art, such as an enzyme-linked immunosorbent assay (ELISA).

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Suitable pharmaceutical compositions and vaccines for use in this aspect of the present invention are those that contain at least one polypeptide comprising an immunogenic portion of a *Leishmania* antigen disclosed herein (or a variant thereof). Preferably, the polypeptides employed in the pharmaceutical compositions and vaccines are complementary, as described above. Soluble *Leishmania* antigens, with or without additional polypeptides, may also be employed.

The pharmaceutical compositions and vaccines described herein may also be used to treat a patient afflicted with a disease responsive to IL-12 stimulation. The patient may be any warm-blooded animal, such as a human or a dog. Such diseases include infections (which may be, for example, bacterial, viral or protozoan) or diseases such as cancer. In one embodiment, the disease is leishmaniasis, and the patient may display clinical symptoms or may be asymptomatic. In general, the responsiveness of a particular disease to IL-12 stimulation may be determined by evaluating the effect of treatment with a pharmaceutical composition or vaccine of the present invention on clinical correlates of immunity. For example, if treatment results in a heightened Th1 response or the conversion of a Th2 to a Th1 profile, with accompanying clinical improvement in the treated patient, the disease is responsive to IL-12 stimulation. Polypeptide administration may be as described below, or may extend for a longer period of time, depending on the indication. Preferably, the polypeptides employed in the pharmaceutical compositions and vaccines are complementary, as described above. A particularly preferred combination contains polypeptides that comprise immunogenic portions of M15, Ldp23, Lbhsp83, Lt-1 and LbeIF4A, Lmsp1a, Lmsp9a, and MAPS-Soluble Leishmania antigens, with or without additional polypeptides, may also be employed.

Routes and frequency of administration, as well as dosage, for the above aspects of the present invention will vary from individual to individual and may parallel

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those currently being used in immunization against other infections, including protozoan, viral and bacterial infections. In general, the pharmaceutical compositions and vaccines may be administered by injection (e.g., intracutaneous, intramuscular, intravenous or subcutaneous), intranasally (e.g., by aspiration) or orally. Between 1 and 12 doses may be administered over a 1 year period. For therapeutic vaccination (i.e., treatment of an infected individual), 12 doses are preferably administered, at one month For prophylactic use, 3 doses are preferably administered, at 3 month intervals. In either case, booster vaccinations may be given periodically thereafter. Alternate protocols may be appropriate for individual patients. A suitable dose is an amount of polypeptide or DNA that, when administered as described above, is capable of raising an immune response in an immunized patient sufficient to protect the patient from leishmaniasis for at least 1-2 years. In general, the amount of polypeptide present in a dose (or produced in situ by the DNA in a dose) ranges from about 100 ng to about lmg per kg of host, typically from about 10 µg to about 100 µg. Suitable dose sizes will vary with the size of the patient, but will typically range from about 0.1 mL to about 5 mL.

In another aspect, this invention provides methods for using one or more of the polypeptides described above to diagnose *Leishmania* infection in a patient using a skin test. As used herein, a "skin test" is any assay performed directly on a patient in which a delayed-type hypersensitivity (DTH) reaction (such as induration and accompanying redness) is measured following intradermal injection of one or more polypeptides as described above. Such injection may be achieved using any suitable device sufficient to contact the polypeptide or polypeptides with dermal cells of the patient, such as a tuberculin syringe or 1 mL syringe. Preferably, the reaction is measured at least 48 hours after injection, more preferably 72 hours after injection.

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The DTH reaction is a cell-mediated immune response, which is greater in patients that have been exposed previously to a test antigen (i.e., an immunogenic portion of a polypeptide employed, or a variant thereof). The response may measured visually, using a ruler. In general, induration that is greater than about 0.5 cm in diameter, preferably greater than about 1.0 cm in diameter, is a positive response,

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indicative of Leishmania infection, which may or may not be manifested as an active disease.

The polypeptides of this invention are preferably formulated, for use in a skin test, as pharmaceutical compositions containing at least one polypeptide and a physiologically acceptable carrier, as described above. Such compositions typically contain one or more of the above polypeptides in an amount ranging from about 1 µg to 100 µg, preferably from about 10 µg to 50 µg in a volume of 0.1 mL. Preferably, the carrier employed in such pharmaceutical compositions is a saline solution with appropriate preservatives, such as phenol and/or Tween 80™.

The inventive polypeptides may also be employed in combination with one or more known Leishmania antigens in the diagnosis of leishmaniasis, using, for example, the skin test described above. Preferably, individual polypeptides are chosen in such a way as to be complementary to each other. Examples of known Leishmania antigens which may be usefully employed in conjunction with the inventive polypeptides include K39 (Burns et al., Proc. Natl. Acad. Sci. USA, 1993 90:775-779).

The following Examples are offered by way of illustration and not by way of limitation.

#### **EXAMPLES**

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#### EXAMPLE 1

#### PREPARATION OF M15

This Example illustrates the preparation of a Leishmania antigen M15, having the sequence provided in SEQ ID NO:2.

An L. major (Friedlan strain) amastigote cDNA expression library prepared in the \(\lambda ZAP\) II vector (Stratagene, La Jolla, CA) was screened according to manufacturer's instructions using sera obtained from L. major infected BALB/c mice (8 weeks post inoculation). Approximately 40,000 plaques were screened and four clones expressing reactive antigens were purified to homogeneity by two subsequent rounds of low density screening. Bluescript phagemid inserts were excised from positive clones

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for further analysis. An *EcoRI/Sst*II restriction fragment from the 5' end of one partial cDNA insert isolated during first round screening (pLma1-1) was subsequently used as a probe to rescreen for clones containing full length cDNA inserts. The probe was labeled to high specific activity ( 10° cpm/μg) with [ -<sup>32</sup>P]dCTP using the random primer method and was used to screen 10,000 plaques of the *L. major* expression library described above. Positive clones were compared by restriction enzyme digestion and the clone with the largest insert (pfl1-1) was chosen for subsequent analysis.

DNA sequence analyses were performed on an Applied Biosystems automated sequencer using Taq polymerase and dye coupled ddNTP terminators or dyelabeled sequencing primers. The complete sequence of the 2685 bp insert was determined using a combination of primer-directed sequencing and by sequencing a series of overlapping Exonuclease III deletion subclones generated using the Erase-a-base system (Promega, Madison, WI). The sequence of this insert is provided in SEQ ID NO:1, and the deduced amino acid sequence is provided in SEQ ID NO:2.

The complete insert of clone pfl-1 was excised by digestion with BamHI/KpnI and was subcloned in frame into BamHI/KpnI digested pQE31 (QUIAGEN) to generate the construct pM151A. E. coli containing this construct inducibly expressed high levels of the L. major antigen encoded by pfl1-1 (designated as M15) with the addition of a 6-histidine tag at the amino terminus. Large volume cultures (500 ml) of E. coli host cells containing the pM151A construct were induced to express recombinant protein by the addition of 2mM IPTG at mid-log phase of growth. Growth was continued for 4 to 5 hours and bacteria were then pelleted and washed once with cold PBS. Bacteria were resuspended in 20 ml of lysis buffer (50 mM Na<sub>2</sub>HPO<sub>4</sub>, pH 8.0, 300 mM NaCl, 10 mM  $\beta$ -mercaptoethanol) containing 20 mg of lysozyme and were lysed by a 1 hour incubation at 4°C followed by brief sonication. Insoluble material was removed by centrifugation at 10,000xg for 10 minutes and although the recombinant protein was found to be evenly distributed between the soluble and insoluble fractions the insoluble material was discarded at this point. Recombinant protein containing the amino terminal histidine tag was affinity purified using Ni-NTA resin (QIAGEN) according to the manufacturer's recommendations. Briefly, 8 ml of

Ni-NTA resin resuspended in lysis buffer was added to the soluble lysate fraction and binding was conducted with constant mixing for 1 hour at 4°C. The mixture was then loaded into a gravity flow column and the non-binding material was allowed to flow through. The Ni-NTA matrix was washed 3 times with 25 ml of wash buffer (50 mM Na<sub>2</sub>HPO<sub>4</sub>, pH 6.0, 300 mM NaCl, 10 mM β-mercaptoethanol) and bound material was eluted in 25 ml of elution buffer (50 mM Na<sub>2</sub>HPO<sub>4</sub>, pH 5.0, 300 mM NaCl, 10mM β-mercaptoethanol). The eluted material was then dialyzed against 3 changes of PBS, sterile filtered and stored at -20°C. The purified recombinant protein was shown by SDS-PAGE analysis to be free of any significant amount of E. coli protein. A small number of bands of lower molecular weight were assumed to be proteolytic products of the L. major antigen based on their reactivity by western blot analysis. A high titre polyclonal antisera against M15 was generated in rabbits by repeated subcutaneous injection of recombinant protein. Western blot analysis of lysates from L. major promastigotes and amastigotes using this antisera indicated that the protein is constitutively expressed throughout the parasite lifecycle.

## **EXAMPLE 2**

### PREPARATION OF LDP23

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This Example illustrates the preparation of a *Leishmania* antigen Ldp23, having the sequence provided in SEQ ID NO:4.

# A. Purification of MHC Class II-associated Peptides from P388D1 Macrophages Infected with L. donovani

To ascertain that *in vitro* infection of macrophages would load their MHC class II molecules with parasite peptides, initial experiments were carried out to test the ability of *L. donovani*-infected macrophage cell line P388D1 to present parasite antigens to *L. donovani* specific T-cells. This macrophage cell line was chosen because it has the same H-2 haplotype as the BALB/c mouse, which is a strain of mouse

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moderately susceptible to *L. donovani* infection and selected to conduct the *in vivo* experiments. Using a proportion of 3-5 parasites per cell and an initial incubation at room temperature for 4-6 hours follows by 37°C for 24-48 hours, close to 90% of the macrophages were infected. The level of MHC class II molecule expression, as determined by FACS analysis, indicated that infection did not cause an effect on the levels of MHC class II expression when compared to non-infected control cells.

To test the ability of the *L. donovani*-infected P388D1 cells to present parasite antigens, macrophages were infected as indicated above and incubated at 26°C for 6 hours, and then as 37°C for either 24, 48 or 72 hours. At each of these time points the non-adherent cells and free parasites were washed out and the adherent cells were mechanically dislodged, washed and fixed with paraformaldehyde. These cells were then used as antigen presenting cells (APCs) for purified lymph node T-cells from BALB/c mice immunized with *L. donovani* promastigotes. To generate these anti-*L. donovani* specific T-cells, BALB/c mice (H-2<sup>d</sup>) of both sexes (The Jackson Laboratory, Bar Harbor, ME) were immunized at 8 to 14 weeks of age in the rear foot pad with 5-10 x 10<sup>6</sup> *L. donovani* promastigotes emulsified in complete Freünd's adjuvant (CFA) (Difco Laboratories, Madison, MI) as described in Rodrigues et al., *Parasite Immunol.* 14:49 (1992). The draining lymph nodes were excised 8 days after the immunization and T-cells were purified in an anti-mouse Ig column to remove the B cells, as described in Bunn-Moreno and Campos-Neto, J. *Immunol.* 127:427 (1981), followed by a passage through a Sephadex G10 column to remove the macrophages.

Stimulation index was calculated by dividing the cpm obtained for the cells cultured in the presence of infected P388D1 macrophages by the cpm obtained for the cells cultured in the presence of non-infected macrophages, but subjected to the same conditions as the infected macrophages. The results shown Figure 1 indicate that *L. donovani*-infected P388D1 macrophage process parasite antigens and that optimal presentation occurs after 48 hours of infection. No stimulation of the T-cells by the non-infected macrophages was observed.

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To isolate the MHC class II associated *L. donovani* peptides, P388D1 macrophages were infected with *L. donovani* promastigotes for an initial incubation of 6

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hours at room temperature. The cultures were then transferred to 37°C for the remainder of the 48 hour incubation period. At a ratio of 3-5 parasites per macrophage nearly 90% of the macrophages were infected after 24 hours of incubation at 37°C.

The MHC class II molecules were then affinity-purified. Approximately 1.5 x 10<sup>10</sup> L. donovani-infected or an equal number of non-infected P388D1 macrophages were used for each purification. The cells were harvested, washed with PBS and incubated for 30 minutes in cold lysis buffer (PBS, 1% Nonidet P40, 25mM iodoacetamide, 0.04% sodium azide, 1mM aprotinin and 1mM PMSF). The insoluble material was removed by centrifugation at 40,000g for 1 hour and the supernatant was 10 recycled overnight at 4°C over a 5ml anti-MHC class II molecules (H-2<sup>d</sup>) Sepharose column (Protein G Sepharose column to which the monoclonal antibody MK-D6 has been bound). Culture supernatants of MK-D6 hybridoma cells (American Type Culture Collection, Rockville, MD) were employed as the source for anti-MHC class II (H-2<sup>d</sup>) monoclonal antibody. The column was washed with 50ml of lysis buffer and then with 50ml of PBS containing 0.5% octyl glucopyranoside detergent. Bound molecules were eluted from the column with 1M acetic acid in 0.2% NaCl. The MHC/peptide molecules were separated from the IgG (MK-D6 monoclonal antibody) using a Centricon 100 filter unit (Amicon Division, W.R. Grace & Co., Beverly, MA). The peptides were then dissociated from the class II molecules by the addition of acetic acid to 2.5M, followed by separation using a Centricon 10 filter unit. The resulting peptide preparation, present in the low molecular weight sample, was then dried using a speed vac concentrator (Savant Instrument Inc., Farmingdale, NY).

The peptides were redissolved in 200µl of 0.05% TFA and separated by reverse-phase high performance liquid chromatography (RP-HPLC) using a 2.1mm x 25cm Vydac C-18 column at a flow rate of 0.15ml/min employing a 1 to 30% acetonitrile gradient (60 min) followed by a 30 to 60% gradient (30 min) and then a 60 to 80% gradient (90-110 min). Non-infected P388D1 cells were similarly processed to serve as background control for endogenous MHC class II associated peptides. Figure 2 shows a representative experiment; four distinct peaks which are present only in the

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material isolated from infected macrophages (panel B), and not in the material isolated from uninfected macrophages (panel A) are indicated.

Out of three independent peptide extractions, twenty five distinct HPLC peptide peaks were isolated from L. donovani-infected macrophages and were subjected to protein sequence analysis using automated Edman degradation on an Applied Biosystems 477 gas-phase protein sequencer. Protein sequence and amino acid analysis were performed by the W.M. Keck Foundation, Biotechnology Resource Laboratory, Yale University, New Haven, CT. In practically all determinations, no assignment could be made for the first position. Also, in most cases the definition of the amino acid residues of the 10-15 positions was based on the quantitative dominance of one residue over others. Using this approach, the sequences obtained for several peptides showed the presence of 3-6 different residues in many of the 10-15 sequence cycles analyzed for each determination, reflecting a mixture of peptides. In addition, sequences could not be obtained for some peaks because the peptides were blocked. Notwithstanding, three peptides sequences were determined. Amino-acid sequences were searched for identity with proteins in the GenBank database using the GENPETP, PIR and SWISSPROT programs. The sequence data base analysis revealed that one of the peptides was highly homologous to glyceraldehyde-3-phosphate dehydrogenase of various species. Another peptide had homology with elongation factor of several species, including Leishmania.

The third sequence was not clearly related to any known proteins, and is shown below:

XQXPQ(L/K)VFDEXX (SEQ ID NO:11).

# B. Cloning and Sequencing of the Ldp23 Gene

In order to retrieve the *L. donovani* protein that was processed into a peptide associated with the MHC class II molecules of infected macrophages, the peptide sequence of uncertain origin was chosen to guide the strategy for cloning the corresponding parasite gene. A DNA fragment was initially amplified from *L. donovani* promastigote cDNA by PCR. The sense primer was a peptide derived oligonucleotide (5' > GGAATTCCCCInCAGCTInGTInTTCGAC < 3') (SEQ ID NO:12) containing an *EcoRI* restriction endonuclease site (underlined). The bases were

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selected following the preferential codon usage of *L. donovani*, as described in Langford et al., *Exp. Parasitol.* 74:360 (1992). Inosine was used for the residues of positions 4, 6 and 7 because of the low codon usage assurance for the corresponding amino acids. In addition, the carboxyl-terminal L-glutamic acid was not included for the design of the primer. The antisense primer was a poly-thymidine oligonucleotide (oligo dT, downstream primer) containing a *XhoI* restriction endonuclease site.

The gene fragment was amplified from a *L. donovani* promastigote cDNA preparation using the following reaction conditions: one cycle of 3 min at 94°C immediately followed by 35 cycles of 1 min at 94°C, 1 min at 45°C and 1 min at 72°C. The *L. donovani* cDNA was prepared from 5 x 10<sup>7</sup> washed promastigote forms harvested at the log growth phase (3 days culture). The cDNA was obtained using an Invitrogen cDNA cycle<sup>TM</sup> kit (Invitrogen Co., San Diego, CA). Oligonucleotide primers were synthesized by the DNA Synthesis Laboratory, Department of Pathology, Yale University School of Medicine.

The PCR products were analyzed by gel electrophoresis. Only one band of approximately 300 bp was obtained. This fragment was cloned and its sequence confirmed the sequence of the peptide-based primer including the glutamic acid codon, deliberately not included in the primer sequence.

The PCR amplified gene fragment was ligated into the pCR<sup>™</sup> vector using the TA cloning system (Invitrogen Co., San Diego, CA). Transformants were selected in LB medium containing 100µg/ml ampicillin and the plasmid DNA was isolated using the Wizard<sup>™</sup> Minipreps DNA purification kit (Promega Co., Madison, WI). Insert DNA was released with the restriction enzymes *Eco*RI and *Xho*I (New England Biolabs, Beverly, MA), purified from an agarose gel electrophoresis and labeled with <sup>32</sup>P using a random priming method (Megaprime Labeling Kit, Amersham Life Science, Buckinghamshire, England).

This DNA fragment was used as probe to screen a *L. donovani* promastigote cDNA library as described in Skeiky et al., *Infect. Immun.* 62:1643 (1994). An approximately 650 bp cDNA (Ldp23) was excised from the phagemid by *in vivo* excision using the Stratagene protocol. DNA sequencing was performed using the

Sequenase version 2 system (DNA sequencing kit) in the presence or absence of 7-deaza-GTP (United States Biochemical, Cleveland, OH). The sequence is provided as SEQ ID NO:3, and shows complete homology with the original 300 bp PCR fragment. A 525 bp open reading frame containing an ATG codon that follows the last 4 bases of the spliced leader sequence and 3 stop codons adjacent to the poly A tail was identified. This frame also codes the carboxyl terminal sequence (KVFDE) (SEQ ID NO:13) of the purified MHC class II associated peptide. The sequence analysis of the deduced protein sequence revealed one potential glycosylation site (Asn-Cys-Ser) at positions 68-70.

Sequence analysis was performed using the University of Wisconsin-Genetics Computer Group Programs and the GenBank and EMBL data bases of protein and DNA sequences. The search for homology of the Ldp23 gene with known sequences revealed no significant homology.

## C. <u>Bacterial Expression and Purification of Recombinant Protein</u>

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The recombinant L. donovani peptide donor protein was produced in E. coli transformed with the pGEX 2T expression vector in which the Ldp23 gene was subcloned in frame. PCR was used to subclone the cloned gene in frame into the expression vector pGEX 2T. Primers containing the appropriate restriction site enzymes, initiation and termination codons were: GGATCCATGGTCAAGTCCCACTACATCTGC <3' (SEQ ID NO:14) for the upstream primer and 5' > GAATTCAGACCGGATAGAAATAAGCCAATGAAA <3' (SEQ ID NO:15) for the downstream primer (restriction sites of BamHI and EcoRI are underlined respectively). PCR conditions were as indicated above for the amplification of the original peptide related DNA fragment. The template used was pBluescript plasmid containing the cloned gene from the cDNA library.

Overexpression of the recombinant fusion protein was accomplished by growing the transformed *E. coli* (DH5α) and inducing the *tac* promoter with 1mM isopropyl-β-thiogalactopyranoside (IPTG) (Stratagene, La Jolla, CA). Cells were collected, centrifuged, and analyzed for the presence of the fusion protein by SDS-PAGE. A glutathione-S-transferase fusion protein of 43-44 kD was produced,

indicating a leishmanial protein of approximately 18 kD, as glutathione-S-transferase (GST) has a MW of 26 kD. However, the fusion protein was very insoluble and therefore could not be purified by affinity chromatography using a glutathione column. The use of low concentrations of detergents like SDS, sarcosyl, deoxycolate, and octyl-glucopyranoside during the extraction steps was efficient to solubilize the protein but unfortunately prevented its binding to the glutathione column. Other maneuvers, such as the growth of the *E. coli* and incubation and induction of the *tac* promoter with IPTG at 33°C, did not improve the protein solubility. However, the purification was achieved by preparative SDS-PAGE. The band was visualized with 0.1M KCl, cut and electroeluted from the gel followed by extensive dialysis against PBS and concentration on Centricon 10 filters.

Approximately 500μg of purified protein was obtained. The purified protein is shown in Figure 3. In panel A, E. coli (DH5α) transformed with the expression vector pGEX 2T containing the Ldp23 gene was grown in LB medium and the tac promoter was induced with IPTG for 3 hours. The cells were pelleted, resuspended in loading buffer and submitted to SDS-PAGE (10%) under reducing condition. The gel was stained with Coomassie blue. Lane 1 shows the uninduced E. coli and land 2 shows the induced E. coli. The arrow indicates the recombinant protein. Panel B shows the protein prepared as in panel A and submitted to a preparative SDS-PAGE. The band corresponding to the overexpressed recombinant fusion protein was identified by KCl, cut out, electroeluted from the gel strip, dialyzed against PBS and submitted to analytical SDS-PAGE (12%). Numbers on the left side indicate the molecular weights of the markers. Attempts to further purify the leishmanial protein by cleaving it out from the fusion protein GST with thrombin were unsuccessful.

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## D. Expression of Ldp23

To ascertain that the Ldp23 peptide is expressed in *Leishmania* organisms, a Northern blot analysis was performed using RNA prepared from different promastigote growth phases (logarithmic and stationary) and from the amastigote form of these parasites.

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The RNA was prepared from 2 x 10<sup>7</sup> parasite cells using the Micro RNA isolation kit (Stratagene, La Jolla, CA) according to the company's recommended instructions. RNA was prepared from *L. donovani* promastigotes (logarithmic growth phase); from *L. major* promastigotes (logarithmic and stationary growth phases); from *L. amazonensis*, both promastigotes (logarithmic and stationary growth phases) and amastigotes purified from CBA/J infected mice; and from *L. pifanoi*, both promastigotes (logarithmic and stationary growth phases) and amastigotes (from axenic culture medium). *L. donovani* (1S strain), *L. amazonensis* (MHOM/BR/77/LTB0016), *L. major* (MHOM/IR/79/LRC-L251) and *L. pifanoi* (MHOM/VE/60/Ltrod) promastigotes were grown and maintained at 26°C in Schneider's medium containing 20% FCS and 50µg/ml gentamicin. The amastigote forms of *L. amazonensis* were obtained by differential centrifugation of a "pus-like" foot pad lesion of a CBA/J mouse infected for 6 months with this parasite. *L. pifanoi* amastigotes were obtained from axenic culture as previously reported by Pan et al., *J. Euk. Microbiol.* 40:213 (1993).

The hybridization was carried out at 45°C in the presence of 50% formamide, 5x Denhardt's solution, 0.1% SDS,  $100\mu g/ml$  single stranded salmon sperm DNA and 5x SSPE using  $0.45\mu m$  Nytran membrane filters (Schleicher & Schuell, Keene, NH). The probe was the  $^{32}P$  labeled Ldp23 gene.

Figure 4 shows that one single RNA band of 680 bp was observed for all growth phases and forms of all tested *Leishmania*. Within Figure 4, the numbers 1, 2 and 3 refer to RNA obtained from promastigotes at the logarithmic growth phase, promastigotes at the stationary growth phase and amastigote forms, respectively, and the numbers on the left side indicate the molecular weights of the markers in base pairs. This result is consistent with the corresponding gene size (525 bp) and with the molecular weight of the expressed protein and points to the ubiquitous distribution and expression of this gene within the genus *Leishmania*.

E. <u>Induction of Anti-L. donovani Antibody Response in Mice and Rabbits by</u>

<u>Purified Recombinant Protein</u>

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In order to evaluate the immunogenicity of the recombinant leishmanial protein, and to investigate its expression in the parasites, mice and rabbits were immunized with the GST-fusion protein in CFA. BALB/c mice were immunized in the rear foot pad with 5-10µg of protein emulsified in CFA. Protein concentration was determined using the Bio-Rad Protein Assay reagent (Bio-Rad Laboratories, Richmond, CA). The mice were boosted 7 days later with 5-10µg of protein emulsified in incomplete Freünd's adjuvant (IFA) inoculated into the peritoneal cavity. The mice were bled 7 days after the second immunization. New Zealand white rabbits (Millbrook Farm, Amherst, MA) were immunized according to the following protocol: one intramuscular (IM) injection of 25-30µg of purified recombinant protein emulsified in CFA into each thigh on day one; one IM injection of 25-30µg of purified protein emulsified in IFA into each shoulder on day 7; on day 15, 25-30µg of the purified protein in PBS was injected into the subcutaneous tissue. The rabbit was bled 7 days after the last immunization.

Sera were prepared and the anti-Leishmania antibody response was measured by Western blot analysis and by FACScan. In both cases L. donovani promastigotes were used as antigen. Approximately 2 x 10<sup>6</sup> L. donovani promastigotes were grown in Schneider's medium for 3 days (log phase), were washed with PBS, lysed with SDS-PAGE loading buffer and submitted to electrophoresis under reducing conditions using a 15% polyacrylamide gel. The proteins were transferred onto 0.45  $\mu$ Immobilon-P transfer membrane (Millipore Co., Bedford, MA) using a wet-type electroblotter (Mini Trans-Blot Electrophoretic Transfer Cell, Bio Rad Life Science Division, Richmond, CA) for 2 hours at 50 V. The membranes were blocked overnight at room temperature with PBS containing 3% normal goat serum (NGS), 0.2% Tween-20 and 0.05% sodium azide, followed by 3 washes with PBS. The blots were then incubated for 3-4 hours at 4°C with a 1/200 dilution of pre-immune rabbit serum (lane A, Figure 5) or with the same dilution of anti-fusion protein rabbit antiserum (lane B, The sera was previously absorbed 2x with non-viable desiccated Mycobacterium tuberculosis H-37 RA (Difco Laboratories, Detroit, MI) and were diluted in PBS containing 1% NGS and 5% powdered non-fat bovine milk (Carnation,

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Nestlé Food Company, Glendale, CA). The membranes were then washed with PBS, incubated for 1 hour at room temperature with goat anti-rabbit IgG antibody conjugated with alkaline phosphatase (Promega, Madison, WI), washed once with PBS and 2x with veronal buffer pH 9.4. The reaction was visualized using the substrate mixture 5-bromo-4-chloro-3-indoyl-phosphate and nitroblue tetrazolium (Kirkegaard & Perry Laboratories Inc., Gaithersburg, MD) according to the manufacturer's instructions.

Figure 5 shows that the rabbit anti-recombinant protein antiserum detects a single protein of 23 kDa (Ldp23) in the *Leishmania* crude extract antigen preparation. No bands were observed when an anti-GST antiserum was used (not shown). Moreover, the FACScan analysis (Figure 6) shows that the antibody induced by the recombinant Ldp23 reacts with intact live *L. donovani* promastigotes, thus pointing to a cell surface expression of this molecule on these organisms. The dotted line in Figure 6 shows the indirect immunofluorescence performed using pre-immune mouse serum and the solid line in Figure 6 shows the result obtained with mouse anti-GST-Ldp23 antiserum. Both sera were diluted at 1/100. Parasites were washed with staining buffer and incubated with FITC conjugated goat anti-mouse immunoglobulin antibody. Fluorescence intensity was analyzed by FACScan.

# F. Recognition of Recombinant Ldp23 by Leishmania-Specific Lymph Node T-cells

To test the responsiveness of T-cells to the Ldp23 protein, two sets of experiments were performed. In the first experiment, lymph node T-cells (10<sup>5</sup>/well) from BALB/c mice immunized with *L. donovani* promastigotes (as described above) were stimulated to proliferate with 2 x 10<sup>5</sup> Mitomycin C-treated normal mononuclear spleen cells (APC) and pulsed with the purified recombinant fusion protein. Proliferation of T-cells was measured at 72 hours of culture. Values are expressed in Figure 7 as cpm and represent the mean of [<sup>3</sup>H]TdR incorporation of triplicate cultures. Background cpm of cells (T cells + APC) cultured in the presence of medium alone was 1291. Figure 7 shows that *Leishmania* specific T-cells proliferate well and in a dose response manner to recombinant Ldp23. No response was observed when purified GST

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was added instead of the recombinant fusion protein nor when lymph node T-cells from mice immunized with CFA alone were stimulated to proliferate in the presence of the Leishmanial fusion protein (not shown).

The recognition of the recombinant Ldp23 protein by Leishmania-specific T-cells was also tested using two murine models of leishmaniasis, the L. major highly susceptible BALB/c mice and the L. amazonensis susceptible CBA/J mice as described in Champsi and McMahon-Pratt, Infect. Immun. 56:3272 (1988). These models were selected to investigate the cytokine pattern induced by Ldp23. In the mouse model of leishmaniasis, resistance is associated with Th 1 cytokines while susceptibility is linked to Th 2 responses.

Lymph node cells were obtained 3 weeks after the initiation of infection of BALB/c mice with *L. major* and the ability of these cells to recognize the recombinant Ldp23 was measured by proliferation and by the production of the cytokines IFN-γ and IL-4. 2 x 10<sup>6</sup> cells obtained from the draining popliteal lymph node of infected mice were cultured for 72 hours in the presence of recombinant Ldp23 or *Leishmania* lysate. The levels of IFN-γ and IL-4 in culture supernatants were measured by ELISA as previously described (Chatelain et al., *J. Immunol. 148*:1172 (1992), Curry et al., *J. Immunol. Meth. 104*:137 (1987), and Mossman and Fong, *J. Immunol. Meth. 116*:151 (1989)) using specific anti IFN-γ and IL-4 monoclonal antibodies (PharMingen, San Diego, CA).

Ldp23 did stimulate these cells to proliferate (not shown) and induced a typical Th 1 type of cytokine response as indicated by the production of high levels of IFN-γ (panel A of Figure 8) and no IL-4 (panel B of Figure 8). Stimulation of these cells with a *Leishmania* crude lysate yielded a mixed Th cytokine profile. Exactly the same pattern of cytokine production was obtained from the CBA/J mice infected with *L. amazonensis* (not shown). These results clearly indicate that Ldp23 is a powerful and selective activator of the Th 1 cytokines by mouse cells.

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# EXAMPLE 3

#### PREPARATION OF HSP83

This Example illustrates the preparation of a Leishmania antigen Hsp83, having the sequence provided in SEQ ID NO:6.

A genomic expression library was constructed with sheared DNA from L. braziliensis (MHOM/BR/75/M2903) in bacteriophage λZAP II (Stratagene, La Jolla, CA). The expression library was screened with Escherichia coli preadsorbed serum from an L. braziliensis-infected individual with ML. Immunoreactive plaques were purified, and the pBSK(-) phagemid was excised by protocols suggested by the manufacturer. Nested deletions were performed with exonuclease III to generate overlapping deletions for single-stranded template preparations and sequencing. Single-stranded templates were isolated following infection with VCSM13 helper phage as recommended by the manufacturer (Stratagene, La Jolla, CA) and sequenced by the dideoxy chain terminator method or by the Taq dye terminator system using the Applied Biosystems automated sequencer model 373A.

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Recombinant antigens produced by these clones were purified from 500 ml of isopropyl-β-D-thiogalactopyranoside (IPTG)-induced cultures as described in Skeiky et al., *J. Exp. Med. 176*:201-211 (1992). These antigens were then assayed for the ability to stimulate PBMC from *Leishmania*-infected individuals to proliferate and secrete cytokine. Peripheral blood was obtained from individuals living in an area (Corte de Pedra, Bahia, Brazil) where *L. braziliensis* is endemic and where epidemiological, clinical, and immunological studies have been performed for over a decade, and PBMC were isolated from whole blood by density centrifugation through Ficoll (Winthrop Laboratories, New York, N.Y.). For *in vitro* proliferation assays, 2 X 10<sup>5</sup> to 4 X 10<sup>5</sup> cells per well were cultured in complete medium (RPMI 1640 supplemented with gentamicin, 2-mercaptoethanol, L-glutamine, and 10% screened pooled A+ human serum; Trimar, Hollywood, Calif.) in 96-well flat-bottom plates with or without 10 μg of the indicated antigens per ml or 5 μg of phytohemagglutinin per ml (Sigma Immunochemicals, St. Louis, Mo.) for 5 days. The cells were then pulsed with

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 $1 \mu \text{Ci}$  of [3H]thymidine for the final 18 h of culture. For determination of cytokine production 0.5 to 1 ml of PBMC was cultured at  $1 \times 10^6$  to  $2 \times 10^6$  cells per ml with or without the *Leishmania* antigens for 48 and 72 h.

The supernatants and cells were harvested and analyzed for secreted cytokine or cytokine mRNAs. Aliquots of the supernatants were assayed for gamma interferon (IFN-γ), tumor necrosis factor alpha (TNF-α), interleukin-4 (IL-4), and IL-10 as described in Skeiky et al., J. Exp. Med. 181:1527-1537 (1995). For cytokine mRNA PCR analysis, total RNA was isolated from PBMC and cDNA was synthesized by using poly(dT) (Pharmacia, Piscataway, NJ) and avian mycloblastosis virus reverse transcriptase. Following normalization to β-actin, diluted cDNA was amplified by PCR using Taq polymerase (Perkin-Elmer Cetus, Foster City, CA) with 0.2 µM concentrations of the respective 5' and 3' external primers in a reaction volume of 50 µl. The nucleotide sequences of the primary pairs and the PCR conditions used were as described in Skeiky et al., J. Exp. Med. 181:1527-1537 (1995). We verified that our PCR conditions were within the semiquantitative range by initially performing serial dilutions of the cDNAs and varying the number of cycles used for PCR. Plasmids containing the human sequences for IL-2, IFN-γ, IL-4, IL-10, and β-actin were digested, and the DNA inserts were purified after separation on 1% agarose gels. Radiolabeled <sup>32</sup>P probes were prepared by the random priming method. PCR products were analyzed by electrophoresis on 1.5% agarose gels, transferred to nylon membranes, and probed with the appropriate <sup>32</sup>P-labeled DNA insert.

A recombinant clone was identified in the above assays which, following sequence comparison of its predicted amino acid sequence with sequences of other proteins, was identified as a *Leishmania braziliensis* homolog of the eukaryotic 83 kD heat shock protein (Lbhsp83). The sequence of the clone is provided in SEQ ID NO:5 and the deduced protein sequence is provided in SEQ ID NO:6. On the basis of the homology, this clone, designated Lbhsp83a, appears to lack the first 47 residues of the full length 703 amino acid residues. Lbhsp83 has an overall homology of 94% (91% identity and 3% conservative substitution), 91% (84% identity and 7% conservative substitution) and 77% (61% identity and 16% conservative substitution) with *L*.

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amazonensis hsp83, T. cruzi hsp83 and human hsp89, respectively. A second clone (designated Lbhsp83b), which contained the 43 kD C-terminal portion of hsp83 (residues 331 to 703) was also isolated. Figure 19 presents a comparison of the Lbhsp83 sequence with L. amazonensis hsp83(Lahsp83), T. cruzi hsp83 (Tchsp83) and human hsp89 (Huhsp89).

The results of proliferation assays using Lbhsp83a are shown in Table 1. Cells from all mucosal leishmaniasis (ML) patients proliferated strongly in response to Lbhsp83a, with stimulation indices (SIs) ranging from 19 to 558 (as compared to 20 to 1,634 for parasite lysate). Proliferation of PBMC from cutaneous leishmaniasis (CL) patients was variable and except for levels in two patients (IV and VII), levels were significantly lower than those of ML patients. By comparison, the proliferative responses of individuals with self-healing CL to Lbhsp83a were similar to those of individuals with ML. However, the responses of all six self-healing individuals to Lbhsp83 were consistently higher than those to Lbhsp83b. This suggests that PBMC from self-healing CL patients preferentially recognize one or more T-cell epitopes located within the amino portion of Lbhsp83.

Table 1

In vitro Proliferation of PMBC from L. braziliensis-infected Individuals
in Response to Lbhsp83

Group and Patient	Mean [3H]thymidine incorporation [103 cpm (SD)], SI with:			
	Lysate	Lbhsp83a	Lbhsp83b	
MIL			*	
I	41.3, (1.3), 294	32.5, (6.6), 221	46.7, (1.4), 318	
π	44.2, (0.5), 104	20, (3.7), 47	36.7, (0.76), 86	
Ш	27.4, (1.5), 150	8.1, (1.7), 44	9.9, (0.32), 54	
IV	52.7, (3.3), 138	54.1, (6.2), 142	32.0, (1.3), 84	
v	140.6, (7.6), 308	151.8, (57), 333	150.4, (7.9), 331	
VI	15.8, (1.8), 20	21.3, (4.4), 28	14.4, (1.3), 19	
VII	300.1, (9.4), 1634	102.1, (7.6), 558	41.7, (4.9), 228	

Group and Patient	Mean [3H]thymidine incorporation [103 cpm (SD)], SI with:			
	Lysate	Lbhsp83a	Lbhsp83b	
CL			. (4)	
I	0.26, (0.0), 1.5	0.57, (0.3), 3.3	0.43, (0.17), 3.3	
II	55.63, (8.6), 218	0.42, (0.0), 1.6	0.8, (0.14), 3.2	
III	0.39, (0.5), 4.0	3.4, (0.5), 9	2.6, (0.9), 6.6	
IV	19.14, (1.3), 87	7.17, (0.6), 32	5.9, (0.9), 27	
V	0.32, (0.2), 3.0	1.47, (0.5), 14	0.3, (0.1), 3.0	
VI	0.77, (0.1), 4.7	1.44, (0.2), 9	1.3, (0.6), 8.0	
VII	4.01, (1.0), 2.0	60.3, (8.5), 15	66.7, (3.9), 16.6	
Self-healing CL				
I	19.7, (4.4), 94	61.3, (4.6), 293	5.0, (2.0), 24	
· · II	0.6, (0.1), 6.5	7.0, (2.0), 79	1.2, (0.8), 13	
III	59.6, (7.1), 519	49.4, (3.1), 429	21.4, (3.7), 186	
IV	0.2, (0.1), 1.6	13.1, (1.7), 108	0.6, (0.1), 5	
V	27.1, (2.0), 225	6.3, (2.6), 52	3.0, (1.5), 25	
VI	130.3, (14), 340	28.2, (2.9), 74	7.7, (3.8), 20	
Control (uninfected)	·		· ·	
<b>I</b>	0.19, (0.0), 1.4	0.18, (0.0), 1.3	0.40, (0.16), 2.8	
II .	0.31, (0.1), 1.7	0.19, (0.0), 1.0	0.27, (0.0), 1.5	
Ш	0.44, (0.2), 4.1	0.48, (0.1), 5.0	0.51, (0.2), 5.2	
IV	0.4, (0.1), 3.2	0.52, (0.2), 5.1	0.50, (0.1), 5.0	

A more detailed analysis of cytokine patterns of PBMC from ML patients was performed by reverse transcriptase PCR. Cytokine mRNAs were evaluated in cells prior to culturing (Figure 9, lanes O) or following culturing in the absence (lanes –) or presence of the indicated antigen for 48 and 72 h. Figure 4A shows the results for five of the six ML patients whose PBMC were analyzed. In about half of the ML patients, noncultured (resting) PBMC had detectable levels of mRNA for IFN-γ, IL-2, and IL-4 but not IL-10. CL patient PBMC, however, had IL-10 mRNA in the resting state in addition to mRNAs for the other cytokines tested (Figure 4B). Following *in vitro* culture without antigen, the levels of mRNA for IFN-γ, IL-2, and IL-4 in resting cells from ML patients decreased to background levels while IL-10 mRNA levels increased. In contrast, PBMC of most CL patients had stable or increased IL-10

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mRNA, while the mRNAs for IL-2, IFN-γ, and IL-4 were reduced to barely detectable levels in the absence of antigen stimulation.

In PBMC of three ML patients, stimulation with lysate resulted in increased expression of mRNA for IFN-γ, IL-2, and IL-4 but not IL-10. By comparison, both Lbhsp83 polypeptides elicited the production of mRNA for IFN-γ and IL-2 from all ML patient PBMC tested. In contrast, profiles of mRNA for IL-10 and IL-4 differed for the two hsp83 polypeptides. Lbhsp83a stimulated the production of IL-10 but not IL-4 mRNA (patients I, II, III, and IV), while Lbhsp83b stimulated the production of IL-4 but not IL-10 mRNA in all six patients.

All CL patients tested responded to both Lbhsp83 polypeptides as well as to the parasite lysate by upregulating the synthesis of mRNAs for IL-2 and IFN-γ, and in two of four patients (I and IV), the level of IL-4 mRNA also increased, indicating stimulation of both Th1 and Th2 cytokines. Interestingly and as in the case of ML patient uncultured PBMC which did not have detectable levels of IL-10 mRNA, Lbhsp83a and not Lbhsp83b stimulated PBMC from one CL patient (IV) to synthesize IL-10 mRNA. However, in the other three patients (I, II, and III) with resting levels of IL-10 mRNA, both rLbhsp83 polypeptides as well as the parasite lysate downregulated the expression of IL-10 mRNA.

PBMC supernatants were also assayed for the presence of secreted IFN-γ, TNF-α, IL-4, and IL-10. Cells from all ML and self-healing CL patients (seven and six patients, respectively) and from four of seven CL patients were analyzed for secreted IFN-γ following stimulation with both rLbhsp83 polypeptides, parasite lysate and Lbhsp70, an *L. braziliensis* protein homologous to the eukaryotic 70 kD heat shock protein (Figure 10A). In general, rLbhsp83a stimulated patient PBMC to secrete higher levels of IFN-γ than did rLbhsp83b (0.2 to 36 and 0.13 to 28 ng/ml, respectively). The presence of secreted IFN-γ correlated well with the corresponding mRNA detected by PCR

PBMC from four of five ML patients (I, II, V, and VII) had supernatant TNF- $\alpha$  levels (0.8 to 2.2 ng/ml) higher than those detected in cultures of PBMC from uninfected controls following stimulation with parasite lysate (Figure 10B). Similarly,

the same PBMC were stimulated by rLbhsp83 to produce levels of TNF- $\alpha$  in supernatant ranging from 0.61 to 2.9 ng/ml. Compared with those of uninfected controls, PBMC from three (I, V, and VI), five (I, II, IV, V, and VI), and two (II and V) of six individuals analyzed produced higher levels of TNF- $\alpha$  in response to parasite lysate, rLbhsp83a, and rLbhsp83b, respectively. The levels of TNF- $\alpha$  produced by PBMC from CL patients in response to parasite lysate were comparable to those produced by uninfected controls. However, rLbhsp83 stimulated TNF- $\alpha$  production in the PBMC of two of these patients. rLbhsp83a stimulated higher levels of TNF- $\alpha$  production than did rLbhsp83b. In the absence of antigen stimulation, only PBMC from ML patients (five of six) produced detectable levels of supernatant TNF- $\alpha$  (60 to 190 pg/ml).

In agreement with the IL-10 mRNA, IL-10 was detected by ELISA in the antigen-stimulated PMBC culture supernatants from ML and CL patients. The levels (49 to 190 pg) were significantly higher (up to 10-fold) following stimulation with rLbhsp83a compared with those after parallel stimulation of the same cells with rLbhsp83b (Figure 11). Parasite lysate also stimulated PMBC from some of the patients to produce IL-10. Although rLbhsp83 stimulated PMBC from uninfected individuals to produce IL-10, with one exception, the levels were lower than those observed with patient PMBC. IL-4 was not detected in any of the supernatants analyzed. Therefore, the level of any secreted IL-4 is below the detection limit of the ELISA employed (50 pg/ml). Taken together, the results demonstrate that a predominant Th1-type cytokine profile is associated with PMBC from *L. braziliensis*-infected individuals following stimulation with rLbhsp83 polypeptides.

antibody production to Lbhsp83, we compared the antibody (immunoglobulin G) reactivities to Lbhsp83 in sera from the three patient groups (Figure 12). The ELISA reactivities of ML patient sera with rLbhsp83a were comparable to those observed with parasite lysate, and in general, there was a direct correlation between ML patient anti-Lbhsp83 antibody titer and T-cell proliferation. Of 23 serum samples from ML patients analyzed, 22 were positive (~96%) with absorbance values of 0.20 to >3.0. Eleven of

the ML patient serum samples had optical density values that were >1. In general, CL patients had significantly lower anti-Lbhsp83 antibody titers  $(\bar{x} = 0.74; \text{ standard error of the mean [SEM]} = 0.1)$  compared to those of ML patients. Therefore, ML and CL patient anti-rhsp83 antibody titers correlated with their respective T-cell proliferative responses. Anti-rLbhsp83 antibody titers were significantly higher in patients with ML  $(\bar{x} = 1.5; \text{ SEM} = 0.2)$  than in self-healing CL patients  $(\bar{x} = 0.35; \text{ SEM} = 0.056)$ , although their T-cell proliferative responses were similar. In fact, anti-Lbhsp83 antibody titers in serum from self-healing CL patients were comparable to those from uninfected controls  $(\bar{x} = 0.24; \text{ SEM} = 0.028)$ . By using 2 standard deviations greater than the mean absorbance value of uninfected control (0.484) as a criterion for positive reactivity to Lbhsp83, eight of nine of the self-healing patient serum samples tested were negative.

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#### **EXAMPLE 4**

#### PREPARATION OF CLONES ENCODING LT-210

This Example illustrates the preparation of clones encoding portions of the *Leishmania* antigen Lt-210, and which has the sequence provided in SEQ ID NO:8.

An expression library was constructed from *L. tropica* (MHOM/SA/91/WR1063C) genomic DNA. The DNA was isolated by solubilizing *L. tropica* promastigotes in 10mM Tris-HCl, pH 8.3, 50mM EDTA, 1% SDS and treating with 100μg/ml RNaseA and 100μg/ml proteinase K. The sample was then sequentially extracted with an equal volume of phenol, phenol: chloroform (1:1), and Chloroform. DNA was precipitated by adding 0.1 volume of 3M sodium acetate (pH 5.2) and 2.5 volume 95% ethanol. The precipitate was resuspended in 10μM Tris, 1mM EDTA. DNA was sheared by passage through a 30-gauge needle to a size range of 2-6 kilobase, and was repaired by incubation with DNA polI in the presence of 100 μM each dATP, dCTP, dGTP, and dTTP. *Eco*RI adapters were ligated to the DNA

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fragments. After removal of unligated adapters by passage over a G-25 Sephadex<sup>TM</sup> column, the fragments were inserted in EcoRI cut Lambda ZapII (Stratagene, La Jolla, CA).

Approximately 43,000 pfu were plated and screened with sera isolated from viscerotropic leishmaniasis (VTL) patients. Sera from VTL patients were received from Drs. M. Grogl and A. Magill. The VTL patient group included eight individuals from whom parasites were isolated and cultured, seven of which had confirmed infection with *L. tropica*. Four other patients were culture negative, but were still considered to be infected based on either PCR analysis or a positive monoclonal antibody smear (Dr. Max Grogl, personal communication). Serum samples from the 11 infected patients were pooled and anti-*E. coli* reactivity removed by affinity chromatography (Sambrook et al., *supra*, p. 12.27-12.28). Lambda phage expressing reactive proteins were detected after antibody binding by protein A-horseradish peroxidase and ABTS substrate.

Three clones, Lt-1, Lt-2, and Lt-3, containing a portion of the Lt-210 gene were identified and purified. The clones ranged in size from 1.4 to 3.3 kb and encoded polypeptides of 75 kD, 70 kD, and 120 kD, respectively. These three clones contain partial sequences of the Lt-210 gene. Lt-1 and Lt-2 are overlapping clones and were chosen for further study.

The DNA sequences of Lt-1 and Lt-2 were determined. Exonuclease III digestion was used to create overlapping deletions of the clones (Heinikoff, Gene 28:351-359, 1984). Single strand template was prepared and the sequence determined with Applied Biosystems Automated Sequencer model 373A or by Sanger dideoxy sequencing. The sequence on both strands of the coding portion of Lt-1 clone was determined. The partial sequence of one strand of Lt-2 clone was determined.

SEQ ID NO:7 presents the DNA sequence of Lt-1, and SEQ ID NO:8 provides the predicted amino acid sequence of the open reading frame. The DNA sequence of the coding portion of the Lt-1 clone includes a repeated nucleotide sequence at the 5' portion of the clone containing eight copies of a 99 bp repeat, three copies of a 60 bp repeat unit, which is part of the larger 99 bp repeat, and 800 bp of

non-repeat sequence. The deduced amino acid sequence of the 99 bp repeat contains limited degeneracies. The mass of the predicted recombinant protein is 67,060 Daltons. A database search of PIR with the predicted amino acid sequence of the open reading frame yielded no significant homology to previously submitted sequences. Predicted secondary structure of the repeat portion of the clone is entirely  $\alpha$ -helical.

Sequence analysis of Lt-2 revealed that the 3' portion of the clone consisted of a mixture of 60 and 99 bp repeats that were identical, excepting occasional degeneracies, to the 60 and 99 bp repeats observed in Lt-1. Collectively, the sequencing data suggest that Lt-1 and Lt-2 are different portions of the same gene, Lt-2 being upstream of Lt-1, with possibly a small overlap.

Hybridization analysis confirmed that rLt-2 and rLt-1 contain overlapping sequences. Genomic DNAs of various *Leishmania* species were restricted with a variety of enzymes, separated by agarose gel electrophoresis, and blotted on Nytran membrane filter (Schleicher & Schuell, Keene, NH). Inserts from rLt-1 and rLt-1 2 were labeled with <sup>32</sup>P-CTP by reverse transcriptase from random oligonucleotide primers and used as probes after separation from unincorporated nucleotides on a Sephadex G-50 column. Hybridizations using the rLt-1 or the rLt-2 probe are performed in 0.2M NaH<sub>2</sub>PO<sub>4</sub>/3.6 M NaCl at 65°C, whereas hybridization using the rLt-1r probe is performed in 0.2 M NaH<sub>2</sub>PO<sub>4</sub>/3.6 M NaCl/0.2 M EDTA at 60°C overnight.

Filters are washed in 0.075 M NaCl/0.0075 M sodium citrate pH 7.0 (0.15 M NaCl/0.0150 M sodium citrate for the Lt-1r probe), plus 0.5% SDS at the same temperature as hybridization.

Genomic DNA from a number of Leishmania species including L. tropica were analyzed by Southern blots as described above using the Lt-1, Lt-2, and Lt-1r inserts separately as probes. Collectively, various digests of L. tropica DNA indicate that this gene has a low copy number. A similar, overlapping pattern was observed using either the Lt-1 or Lt-2 insert as a probe, consistent with the premise that these two clones contain sequences near or overlapping one another. In addition, sequences hybridizing with these clones are present in other Leishmania species.

L. tropica isolates have limited heterogeneity. Southern analyses of digested genomic DNA from four L. tropica parasite strains isolated from VTL patients and three L. tropica parasite strains isolated from CL cases (two human, one canine) were performed. The Lt-Ir insert described below was labeled and used as a probe. The seven different L. tropica isolates yielded similar intensities and restriction patterns, with only a single restriction fragment length polymorphism among the isolates. These data, along with Southern analyses with additional enzymes, indicate limited heterogeneity in this region among the L. tropica isolates.

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The recombinant proteins of Lt-1 and Lt-2 were expressed and purified. The nested deletion set of Lt-1 formed for sequencing included a clone referred to as Lt-lr, which contains one and one-third repeats. This polypeptide was also expressed and purified. In vivo excision of the pBluescript SK- phagemid from Lambda Zap II was performed according to the manufacturer's protocol. Phagemid virus particles were used to infect E. coli XL-1 Blue. Production of protein was induced by the addition of IPTG. Protein was recovered by first lysing pellets of induced bacteria in buffer (LB, 50 mM Tris-HCl, pH 8.0, 100 mM NaCl, 10 mM EDTA) using a combination of lysozyme (750µg/mL) and sonication. rLt-1, rLt-2, and rLt-1r, were recovered from the inclusion bodies after solubilization in 8M urea (rLt-1 and rLt-2) or 4M urea (rLt-1r). Proteins rLt-1 and rLt-2 were enriched and separated by precipitation with 25%-40% ammonium sulfate and rLt-lr was enriched by precipitation with 10%-25% ammonium sulfate. The proteins were further purified by preparative gel electrophoresis in 10% SDS-PAGE. Recombinant proteins were eluted from the gels and dialyzed in phosphate-buffered saline (PBS). Concentration was measured by the Pierce (Rockford, IL) BCA assay, and purity assessed by Coomassie blue staining after SDS-PAGE.

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## **EXAMPLE 5**

## PREPARATION OF LBEIF4A

This example illustrates the molecular cloning of a DNA sequence encoding the L. braziliensis ribosomal antigen LbeIF4A.

A genomic expression library was constructed with sheared DNA from L. braziliensis (MHOM/BR/75/M2903) in bacteriophage λZAPII (Stratagene, La Jolla, CA). The expression library was screened with E. coli-preadsorbed patient sera from an L. braziliensis-infected individual with mucosal leishmaniasis. Plaques containing immunoreactive recombinant antigens were purified, and the pBSK(-) phagemid excised using the manufacturer's protocols. Nested deletions were performed with Exonuclease III to generate overlapping deletions for single stranded template preparations and sequencing. Single stranded templates were isolated following infection with VCSM13 helper phage as recommended by the manufacturer (Stratagene, La Jolla, CA) and sequenced by the dideoxy chain terminator method or by the Taq dye terminator system using the Applied Biosystems Automated Sequencer Model 373A.

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The immunoreactive recombinant antigens were then analyzed in patient T-cell assays for their ability to stimulate a proliferative and cytokine production, as described in Examples 7 and 8 below.

A recombinant clone was identified in the above assays which, following sequence comparison of its predicted amino acid sequence with sequences of other proteins, was identified as a *Leishmania braziliensis* homolog of the eukaryotic initiation factor 4A (eIF4A). The isolated clone (pLeIF.1) lacked the first 48 amino acid residues (144 nucleotides) of the full length protein sequence. The pLeIF.1 insert was subsequently used to isolate the full length genomic sequence.

SEQ ID NO:9 shows the entire nucleotide sequence of the full-length LbeIF4A polypeptide. The open reading frame (nucleotides 115 to 1323) encodes a 403 amino acid protein with a predicted molecular weight of 45.3 kD. A comparison of the predicted protein sequence of LbeIF4A with the homologous proteins from tobacco (TeIF4A), mouse (MeIF4A), and yeast (YeIF4A) shows extensive sequence homology,

with the first 20-30 amino acids being the most variable. The lengths (403, 413, 407, and 395 amino acids), molecular weights (45.3, 46.8, 46.4, and 44.7 kDa), and isoelectric points (5.9, 5.4, 5.5, and 4.9) of LbeIF4A, TeIF4A, MeIF4A and YeIF4A, respectively, are similar. LbeIF4A shows an overall homology of 75.5% (57% identity, 18.5% conservative substitution) with TeIF4A, 68.6% (50% identity, 18.6% conservative substitution) with MeIF4A and 67.2% (47.6% identity, 19.6% conservative substitution) with YeIF4A.

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#### EXAMPLE 6

#### PREPARATION OF SOLUBLE LEISHMANIA ANTIGENS

This Example illustrates the preparation of soluble *Leishmania* antigens from an *L. major* culture supernatant. *L. major* promastigotes were grown to late log phase in complex medium with serum until they reached a density of 2-3 x 10<sup>7</sup> viable organisms per mL of medium. The organisms were thoroughly washed to remove medium components and resuspended at 2-3 x 10<sup>7</sup> viable organisms per mL of defined serum-free medium consisting of equal parts RPMI 1640 and medium 199, both from Gibco BRL, Gaithersburg, MD. After 8-12 hours, the supernatant was removed, concentrated 10 fold and dialyzed against phosphate-buffered saline for 24 hours. Protein concentration was then determined and the presence of at least eight different antigens confirmed by SDS-PAGE. This mixture is referred to herein as "soluble *Leishmania* antigens."

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# **EXAMPLE 7**

# COMPARISON OF INTERLEUKIN-4 AND INTERFERON-γ PRODUCTION STIMULATED BY LEISHMANIA ANTIGENS

This Example illustrates the immunogenic properties of the antigens prepared according to Examples 1, 2, 5 and 6, as determined by their ability to stimulate IL-4 and IFN-γ in lymph node cultures from infected mice and in human PBMC preparations. Lymph node cultures for use in these studies were prepared from *L. major*-infected BALB/c mice 10 days after infection, as described in Example 2. PBMC were prepared using peripheral blood obtained from individuals with cured *L. donovani* infections who were immunologically responsive to *Leishmania*. Diagnosis of the patients was made by clinical findings associated with at least one of the following: isolation of parasite from lesions, a positive skin test with *Leishmania* lysate or a positive serological test. Uninfected individuals were identified based on a lack of clinical signs or symptoms, a lack of history of exposure or travel to endemic areas, and the absence of a serological or cellular response to *Leishmania* antigens. Peripheral blood was collected and PBMC isolated by density centrifugation through Ficoll<sup>TM</sup> (Winthrop Laboratories, New York).

Culture supernatants were assayed for the levels of secreted IL-4 and IFN-γ. IFN-γ was quantitated by a double sandwich ELISA using mouse anti-human IFN-γ mAb (Chemicon, Temucula, CA) and polyclonal rabbit anti-human IFN-γ serum. Human rIFN-γ (Genentech Inc., San Francisco, CA) was used to generate a standard curve. IL-4 was quantitated in supernatants by a double sandwich ELISA using a mouse anti-human IL-4 mAb (M1) and a polyclonal rabbit anti-human IL-4 sera (P3). Human IL-4 (Immunex Corp., Seattle, WA) was used to generate a standard curve ranging from 50 pg/ml to 1 ng/ml.

Figures 13A and 13B, illustrate the mean level of secreted IL-4 and IFN- $\gamma$ , respectively, 72 hours after addition of 10  $\mu$ g/mL of each of the following antigens to a lymph node culture prepared as described above: soluble *Leishmania* antigen (i.e., an extract prepared from ruptured promastigotes which contains membrane and internal

antigens (SLA)), Ldp23, LbeIF4A (LeIF), Lbhsp83, M15 and LmeIF (the *L. major* homolog of LbeIF4A). The levels of secreted IL-4 and IFN-γ in medium alone (*i.e.*, unstimulated) are also shown. While SLA elicits a predominantly Th2 response from lymph node cells of Leishmania-infected mice, Ldp23, LbeIF4A, Lbhsp83 and M15 elicited relatively little IL-4 and large amounts of IFN-γ, consistent with a Th1 response profile.

Figure 14 shows the level of secreted IFN-γ in culture filtrate from infected and uninfected human PBMC preparations 72 hours after addition of 10 μg/mL *L. major* lysate, M15 or L-Rack, an immunodominant leishmanial antigen in murine leishmaniasis. Similarly, Figure 15 illustrates the level of secreted IFN-γ in culture filtrate from infected and uninfected human PBMC preparations 72 hours after addition of 10μg/mL *L. major* lysate, soluble *Leishmania* antigens (prepared as described in Example 6) or L-Rack. These results indicate that M15 and soluble *Leishmania* antigens, but not L-Rack, are potent stimulators of IFN-γ production in patient PBMC, but not in PBMC obtained from uninfected individuals. Thus, M15 and soluble *Leishmania* antigens elicit a dominant Th1 cytokine profile in both mice and humans infected with *Leishmania*.

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#### **EXAMPLE 8**

# COMPARISON OF PROLIFERATION STIMULATED BY LEISHMANIA ANTIGENS

This Example illustrates the immunogenic properties of the antigens prepared according to Examples 1, 2, 5 and 6, as determined by their ability to stimulate proliferation in lymph node cultures from infected mice and in human PBMC preparations.

For *in vitro* proliferation assays,  $2 - 4 \times 10^5$  cells/well were cultured in complete medium (RPMI 1640 supplemented with gentamycin, 2-ME, L-glutamine, and 10% screened pooled A+ human serum; Trimar, Hollywood, CA) in 96-well flat bottom plates with or without 10  $\mu$ g/ml of the indicated antigens or 5  $\mu$ g/ml PHA

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(Sigma Immunochemicals, St. Louis, MO) for five days. The cells were then pulsed with 1  $\mu$ Ci of [<sup>3</sup>H] thymidine for the final 18 hours of culture.

Figure 16 illustrates the proliferation observed after addition of 10  $\mu$  g/mL or 20  $\mu$ g/mL of each of the following antigens to a lymph node culture prepared as described in Example 7: SLA, Ldp23, LbeIF4A, Lbhsp83, and M15. The level of proliferation without the addition of antigen is also shown. Data are represented as mean cpm. These results demonstrate that a variety of leishmanial antigens are capable of stimulatory lymph node cell proliferation from *Leishmania*-infected mice.

Figures 17 and 18 illustrate the proliferation observed in human PBMC preparations from Leishmania-immune and uninfected individuals following the addition of 10 µg/mL M15 and soluble Leishmania antigens, respectively. These values are compared to the proliferation observed following the addition of culture medium, L. major lysate or L-Rack. The results show that M15 and soluble Leishmania antigens stimulate proliferation in Leishmania-immune PBMC, but not in PBMC obtained from uninfected individuals, demonstrating that M15 and soluble antigens (but not L-Rack) are recognized by PBMC from individuals immune to Leishmania due to a previous infection.

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## EXAMPLE 9

## PREPARATION OF LMSP1A AND LMSP9A

This Example illustrates the preparation of two soluble Leishmania antigens, Lmspla and Lmsp9a.

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# A. <u>Purification of Lmsp1a and Lmsp9a from a mixture of soluble L. major antigens</u>

A high titer rabbit sera was raised against *L. major* soluble antigens, prepared as described above in Example 6. Specifically, a New Zealand white rabbit was immunized subcutaneously at multiple sites with 180 µg of *L. major* soluble antigens in a suspension containing 100 µg muramyl dipeptide and 50 % incomplete

Freund's adjuvant. Six weeks later the rabbit was given a subcutaneous boost of  $100 \mu g$  of the same soluble antigen preparation in incomplete Freund's adjuvant. This was followed by two intravenous boosts spaced two weeks apart, each with  $100 \mu g$  of the soluble antigen preparation. Sera was collected from the rabbit 11 days after the final boost.

Anti *E. coli* antibody reactivities were removed from the rabbit sera by pre-adsorbing on nitrocellulose filters containing lysed *E. coli*. Adsorbed sera were evaluated by Western blot analysis using 10 µg *Leishmania* promastigote lysate (lane 1) and 1 µg soluble *L. major* antigen mixture (lane 2). As shown in Figure 20, the rabbit sera was found to be reactive with seven dominant antigens of the soluble *L. major* antigen mixture with molecular weights ranging from 18 to >200 kDa. A four times longer exposure of the same blot revealed three additional immunoreactive species with molecular weights less than 18 kDa. The same sera reacted with approximately 10 antigens of the promastigote lysate, but with a pattern significantly different from that observed with the soluble *L. major* antigens (Figure 20). This is suggestive of potential post-translational modification of the same antigen before (intracellular localization) and after secretion/shedding. Such modifications may include cleavage of a leader sequence and/or the addition of carbohydrate molecules to the secreted/shed antigens.

The rabbit sera described above was subsequently used to screen an L major cDNA expression library prepared from L. major promastigote RNA using the unidirectional Lambda ZAP (uni-ZAP) kit (Stratagene) according to the manufacturer's protocol. A total of 70,000 pfu of the amplified cDNA library was screened with the rabbit sera at a 1:250 dilution. Nineteen positive clones were confirmed in the tertiary screening. The phagemid were excised and DNA from each of the 19 clones was sequenced using a Perkin Elmer/Applied Biosystems Division automated sequencer Model 373A. All 19 clones were found to represent two distinct sequences, referred to as Lmsp1a and Lmsp9a. The determined cDNA sequences for Lmsp1a and Lmsp9a are provided in SEQ ID NO: 19 and 21, respectively, with the corresponding amino acid sequences being provided in SEQ ID NO: 20 and 22, respectively.

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# B. Characterization of Lmspla and Lmsp9a

Fig. 21 shows the full-length cDNA (SEQ ID NO: 19) and predicted amino acid sequence (SEQ ID NO: 20) for the antigen Lmsp1a. The EcoRI/XhoI insert is 1019 bp long and contains the following features: a) the last 17 nt of the spliced leader sequence characteristic of all trypanosoma nuclearly encoded mRNA; b) 39 nt of 5' untranslated sequence; c) an open reading frame of 453 nt long coding for a 151 deduced amino acid sequence with a predicted molecular mass of 16.641 kDa; and d) 471 nt of 3' untranslated sequence terminating with a poly A tail. The predicted amino acid sequence contains three potential phosphorylation sites at amino acid residues 3, 85 and 102. In addition, Lmsp1a contains an RGD sequence at residue 104, a sequence that may play a role in parasite invasion of the macrophage. RGD sequences have been shown to mediate the binding of various adhesion proteins to their cell surface receptors. There is no obvious leader sequence (secretory signal) at the amino terminal portion suggesting that the protein might be shed or excreted. Lmsp1a appears to be one of the most abundant antigens found in the culture supernatant of live promastigote, since 17 of the 19 clones contain sequences of variable lengths identical to Lmsp1a.

Comparison of the amino acid sequence of Lmps1a with known sequences using the DNA STAR system (Version 87) revealed that Lmsp1a shares between 65% to 70% homology with the eukaryotic nucleoside diphosphate kinase protein, also referred to in the mouse and human as a tumor metastasis inhibitor gene.

Southern blot analysis of genomic DNA from *L. major* (Friedlander strain) digested with a panel of restriction enzymes (lanes 1 to 7) and six other *Leishmania* species of different geographic locations digested with PstI (lanes 8 to 13) using the full-length cDNA insert of Lmps1a, demonstrated that Lmsp1a is present in all the species characterized with a high degree of conservation (Fig. 22). This suggests evolutionary significance for the maintenance of Lmsp1a and the existence of homologous species among all the *Leishmania* species.

The remaining two cDNA clones isolated from the soluble *L. major* antigen mixture represent identical sequences (referred to as Lmsp9a; SEQ ID NO: 21), suggesting that the two copies resulted from amplification of the primary library.

Sequencing of the Lmsp9a cDNA revealed that the clone does not contain the full length 5' sequence since it is lacking both the spliced leader and 5' untranslated sequences. The 3' end of the cDNA contains a poly A stretch, as would be expected for a *Leishmania* mRNA. Of the predicted translated sequence (SEQ ID NO: 22), 34 of the 201 amino acids (17%) represent cysteine residues. Comparison of the predicted protein sequence with those of known proteins as described above, revealed some homology with other cysteine rich proteins such as the major surface trophozoite antigen of *Giardia lamblia* and furin proteases.

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#### EXAMPLE 10

# PREPARATION AND CHARACTERIZATION OF MAPS-1A

This Example illustrates the preparation and characterization of the Leishmania antigen MAPS-1A (SEQ ID NO: 24).

A pool of sera was obtained from 5 BALB/c mice that had been given a primary immunization and two boosts with crude *L. major* promastigote culture supernatant as described below in Example 12. These mice were subsequently shown to be protected when challenged with a dose of live *L. major* promastigotes generally found to be lethal. The mouse sera thus obtained were used to screen an *L. major* amastigote cDNA expression library prepared as described in Example 1. Several seroreactive clones were isolated and sequenced using a Perkin Elmer/Applied Biosystems Division automated sequencer Model 373A (Foster City, CA).

One of these clones, referred to herein as MAPS-1A, was found to be full-length. Comparison of the cDNA and deduced amino acid sequences for MAPS-1A (SEQ ID Nos: 23 and 24, respectively) with known sequences in the gene bank using the DNA STAR system revealed no significant homologies to known *Leishmania* sequences, although some sequence similarity was found to a group of proteins, known as thiol-specific antioxidants, found in other organisms.

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For example, supernatants from suitable host/vector systems which secrete recombinant protein into culture media may be first concentrated using a commercially available filter. Following concentration, the concentrate may be applied to a suitable purification matrix such as an affinity matrix or an ion exchange resin. Finally, one or more reverse phase HPLC steps can be employed to further purify a recombinant protein.

In general, any of a variety of expression vectors known to those of ordinary skill in the art may be employed to express recombinant polypeptides of this invention. Expression may be achieved in any appropriate host cell that has been transformed or transfected with an expression vector containing a DNA molecule that encodes a recombinant polypeptide. Suitable host cells include prokaryotes, yeast and higher eukaryotic cells. Preferably, the host cells employed are *E. coli*, yeast or a mammalian cell line such as COS or CHO. The DNA sequences expressed in this manner may encode naturally occurring antigens, portions of naturally occurring antigens, or other variants thereof. For example, variants of a native antigen may generally be prepared using standard mutagenesis techniques, such as oligonucleotide-directed site-specific mutagenesis, and sections of the DNA sequence may be removed to permit preparation of truncated polypeptides.

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In another aspect, the present invention provides epitope repeat sequences, or antigenic epitopes, of a *Leishmania* antigen, together with polypeptides comprising at least two such contiguous antigenic epitopes. As used herein an "epitope" is a portion of an antigen that reacts with sera from *Leishmania*-infected individuals (i.e. an epitope is specifically bound by one or more antibodies present in such sera). As discussed above, epitopes of the antigens described in the present application may be generally identified using techniques well known to those of skill in the art.

In one embodiment, antigenic epitopes of the present invention comprise an amino acid sequence provided in SEQ ID NO:43, 56, 57 or 58. As discussed in more detail below, antigenic epitopes provided herein may be employed in the diagnosis and treatment of *Leishmania* infection, either alone or in combination with other

San Diego, CA, and may generally be used according to the manufacturer's instructions. The level of mRNA encoding one or more specific cytokines may be evaluated by, for example, amplification by polymerase chain reaction (PCR). In general, a polypeptide that is able to induce, in a preparation of about 1-3 x 10<sup>5</sup> cells, the production of 30 pg/mL of IL-12, IL-4, IFN-γ, TNF-α or IL-12 p40, or 10 pg/mL of IL-12 p70, is considered able to stimulate production of a cytokine.

Immunogenic portions of the antigens described herein may be prepared and identified using well known techniques, such as those summarized in Paul, Fundamental Immunology, 3rd ed., 243-247 (Raven Press, 1993) and references cited therein. Such techniques include screening polypeptides derived from the native antigen for immunogenic properties using, for example, the representative techniques described herein. An immunogenic portion of a polypeptide is a portion that, within such representative assays, generates an immune response (e.g., proliferation and/or cytokine production) that is substantially similar to that generated by the full length antigen. In other words, an immunogenic portion of an antigen may generate at least about 25%, and preferably at least about 50%, of the response generated by the full length antigen in the model assays described herein.

Portions and other variants of immunogenic *Leishmania* antigens may be generated by synthetic or recombinant means. Synthetic polypeptides having fewer than about 100 amino acids, and generally fewer than about 50 amino acids, may be generated using techniques well known to those of ordinary skill in the art. For example, such polypeptides may be synthesized using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where amino acids are sequentially added to a growing amino acid chain. *See* Merrifield, *J. Am. Chem. Soc.* 85:2149-2146, 1963. Equipment for automated synthesis of polypeptides is commercially available from suppliers such as Perkin Elmer/Applied BioSystemsDivision, Foster City, CA, and may be operated according to the manufacturer's instructions.

Recombinant polypeptides containing portions and/or variants of a native antigen may be readily prepared from a DNA sequence encoding the antigen.

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immunizing BALB/c mice (e.g., in the rear foot pad) with Leishmania promastigotes emulsified in complete Freund's adjuvant. The draining lymph nodes may be excised following immunization and T-cells may be purified in an anti-mouse Ig column to remove the B cells, followed by a passage through a Sephadex G10 column to remove the macrophages. Similarly, lymph node cells may be isolated from a human following biopsy or surgical removal of a lymph node.

The ability of a polypeptide (e.g., a Leishmania antigen or a portion or other variant thereof) to induce a response in PBMC or lymph node cell cultures may be evaluated by contacting the cells with the polypeptide and measuring a suitable response. In general, the amount of polypeptide that is sufficient for the evaluation of about 2 x 10<sup>5</sup> cells ranges from about 10 ng to about 100 µg, and preferably is about 1-10 µg. The incubation of polypeptide with cells is typically performed at 37°C for about 1-3 days. Following incubation with polypeptide, the cells are assayed for an appropriate response. If the response is a proliferative response, any of a variety of techniques well known to those of ordinary skill in the art may be employed. For example, the cells may be exposed to a pulse of radioactive thymidine and the incorporation of label into cellular DNA measured. In general, a polypeptide that results in at least a three fold increase in proliferation above background (i.e., the proliferation observed for cells cultured without polypeptide) is considered to be able to induce proliferation.

Alternatively, the response to be measured may be the secretion of one or more cytokines (such as interferon- $\gamma$  (IFN- $\gamma$ ), interleukin-4 (IL-4), interleukin-12 (p70 and/or p40), interleukin-2 (IL-2) and/or tumor necrosis factor- $\alpha$  (TNF- $\alpha$ )) or the change in the level of mRNA encoding one or more specific cytokines. In particular, the secretion of interferon- $\gamma$ , interleukin-2, tumor necrosis factor- $\alpha$  and/or interleukin-12 is indicative of a Th1 response, which is responsible for the protective effect against *Leishmania*. Assays for any of the above cytokines may generally be performed using methods known to those of ordinary skill in the art, such as an enzyme-linked immunosorbent assay (ELISA). Suitable antibodies for use in such assays may be obtained from a variety of sources such as Chemicon, Temucula, CA and PharMingen,

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4H6-41 and 8G3-100 may be isolated by means of CD4+ T cell expression cloning as described below. DNA sequences encoding these antigens are provided in SEQ ID NO: 72-79, respectively, with the corresponding predicted amino acid sequences being provided in SEQ ID NO: 80-87. The immunogenic properties of the isolated Leishmania antigens may be evaluated using, for example, the representative methods described herein.

Regardless of the method of preparation, the antigens described herein are immunogenic. In other words, the antigens (and immunogenic portions thereof) are capable of eliciting an immune response in cultures of lymph node cells and/or peripheral blood mononuclear cells (PBMC) isolated from presently or previously Leishmania-infected individuals. More specifically, the antigens, and immunogenic portions thereof, have the ability to induce T-cell proliferation and/or to elicit a dominantly Th1-type cytokine response (e.g., IL-2, IFN-γ, and/or TNF-α production by T-cells and/or NK cells; and/or IL-12 production by monocytes, macrophages and/or B cells) in cells isolated from presently or previously *Leishmania*-infected individuals. A Leishmania-infected individual may be afflicted with a form of leishmaniasis (such as subclinical, cutaneous, mucosal or active visceral) or may be asymptomatic. Such individuals may be identified using methods known to those of ordinary skill in the art. Individuals with leishmaniasis may be identified based on clinical findings associated with at least one of the following: isolation of parasite from lesions, a positive skin test with Leishmania lysate or a positive serological test. Asymptomatic individuals are infected individuals who have no signs or symptoms of the disease. Such individuals can be identified based on a positive serological test and/or skin test with Leishmania lysate.

The term "PBMC," which refers to a preparation of nucleated cells consisting primarily of lymphocytes and monocytes that are present in peripheral blood, encompasses both mixtures of cells and preparations of one or more purified cell types. PBMC may be isolated by methods known to those in the art. For example, PBMC may be isolated by density centrifugation through, for example, Ficoll<sup>TM</sup> (Winthrop Laboratories, New York). Lymph node cultures may generally be prepared by

medium with serum until they reach a density of 2-3 x 10<sup>7</sup> viable organisms per mL of medium. The organisms are thoroughly washed to remove medium components and resuspended at 2-3 x 10<sup>7</sup> viable organisms per mL of defined serum-free medium consisting of equal parts RPMI 1640 and medium 199, both from Gibco BRL, Gaithersburg, MD. After 8-12 hours, the supernatant containing soluble *Leishmania* antigens is removed, concentrated 10 fold and dialyzed against phosphate-buffered saline for 24 hours. The presence of at least eight different antigens within the mixture of *Leishmania* antigens may be confirmed using SDS-PAGE (i.e., through the observation of at least 8 different bands). The immunogenic properties of the soluble *Leishmania* antigens may be confirmed by evaluating the ability of the preparation to elicit an immune response in cultures of lymph node cells and/or peripheral blood mononuclear cells (PBMC) isolated from presently or previously *Leishmania*-infected individuals. Such an evaluation may be performed as described below.

Individual antigens present within the mixture of soluble Leishmania antigens may be isolated by immunizing mice or rabbits with Leishmania culture supernatant, containing soluble antigens, and employing the resultant sera to screen a Leishmania cDNA expression library as described in detail below. This procedure may be used to isolate recombinant DNA molecules encoding the L. major antigens referred to herein as Lmsp1a, Lmsp9a and MAPS-1A. DNA sequences encoding Lmsp1a, Lmsp9a and MAPS-1A are provided in SEQ ID NO: 19, 21 and 23, respectively, with the corresponding predicted amino acid sequences being presented in SEQ ID NO: 20, 22 and 24, respectively. Similarly, sera from mice or rabbits immunized with L. major culture supernatant may be used to screen an L. major genomic DNA library. As detailed below, this procedure may be used to isolate DNA molecules encoding the L. major antigens referred to herein as LmgSP1, LmgSP3, LmgSP5, LmgSP8, LmgSP9, LmgSP13, LmgSP19, and DNA molecules encoding the L. chagasi antigens LcgSP1, LcgSP3, LcgSP4, LcgSP8, and LcgSP10. The DNA sequences encoding these antigens are provided in SEQ ID NO:29-35 and 44-48, respectively, with the corresponding amino acid sequences being provided in SEQ ID NO: 36-42 and 49-53. The L. major antigens referred to herein as 1G6-34, 1E6-44, 4A5-63, 1B11-39, 2A10-37, 4G2-83,

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PBMC isolated from *L. braziliensis*-infected patients. Accordingly, Lbhsp83 is an immunogenic *Leishmania* antigen. Regions of Lbhsp83 that are not conserved with the mammalian gene have been found to be particularly potent for T-cell stimulation and antibody binding. Such regions may be identified, for example, by visual inspection of the sequence comparison provided in Figure 19.

This approach may also be used to isolate a DNA molecule encoding a 210 kD immunogenic *L. tropica* antigen, referred to herein as Lt-210. The preparation and characterization of Lt-210, and immunogenic portions thereof (such as Lt-1 and immunogenic repeat and non-repeat sequences), is described in detail in U.S. Patent Application Serial No. 08/511,872, filed August 4, 1995. The sequence of a DNA molecule encoding Lt-1 is provided in SEQ ID NO:7 and the encoded amino acid sequence is presented in SEQ ID NO:8.

The above approach may further be used to isolate a DNA molecule encoding a *L. braziliensis* antigen referred to herein as LbeIF4A. Briefly, such a clone may be isolated by screening a *L. braziliensis* expression library with sera obtained from a patient afflicted with mucosal leishmaniasis, and analyzing the reactive antigens for the ability to stimulate proliferative responses and preferential Th1 cytokine production in PBMC isolated from *Leishmania*-infected patients, as described below. The preparation and characterization of LbeIF4A is described in detail in U.S. Patent Application Serial Nos. 08/454,036 and 08/488,386, which are continuations-in-part of U.S. Patent Application Serial No. 08/232,534, filed April 22, 1994. The sequence of a DNA molecule encoding LbeIF4A is provided in SEQ ID NO:9 and the encoded amino acid sequence is presented in SEQ ID NO:10. Homologs of LbeIF4A, such as that found in *L. major*, may also be isolated using this approach, and are within the scope of the present invention.

Compositions of the present invention may also, or alternatively, contain soluble Leishmania antigens. As used herein, "soluble Leishmania antigens" refers to a mixture of at least 8 different Leishmania antigens that may be isolated from the supernatant of Leishmania promastigotes of any species grown for 8-12 hours in protein-free medium. Briefly, the organisms are grown to late log phase in complex

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This approach may be used to identify a 23 kD *Leishmania donovani* antigen (referred to herein as Ldp23). The sequence of a DNA molecule encoding Ldp23 is provided in SEQ ID NO:3 and the amino acid sequence of Ldp23 is provided in SEQ ID NO:4. Using the methods described herein, Ldp23 has been shown to induce a Th1 immune response in T-cells prepared from *Leishmania*-infected mice.

Alternatively, a *Leishmania* cDNA or genomic expression library may be screened with serum from a *Leishmania*-infected individual, using techniques well known to those of ordinary skill in the art. DNA molecules encoding reactive antigens may then be used to express the recombinant antigen for purification. The immunogenic properties of the purified *Leishmania* antigens may then be evaluated using, for example the representative methods described herein.

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For example, sera from Leishmania-infected mice may be used to screen a cDNA library prepared from Leishmania amastigotes. Reactive clones may then be expressed and recombinant proteins assayed for the ability to stimulate T-cells or NK cells derived from Leishmania-immune individuals (i.e., individuals having evidence of infection, as documented by positive serological reactivity with Leishmania-specific antibodies and/or a Leishmania-specific DTH response, without clinical symptoms of leishmaniasis). This procedure may be used to obtain a recombinant DNA molecule encoding the Leishmania antigen designated M15. The sequence of such a DNA molecule is provided in SEQ ID NO:1, and the amino acid sequence of the encoded protein is provided in SEQ ID NO:2.

A similar approach may be used to isolate a genomic DNA molecule encoding an immunogenic *Leishmania braziliensis* antigen, referred to herein as Lbhsp83. More specifically, a genomic clone encoding Lbhsp83 may be isolated by screening a *L. braziliensis* expression library with sera from a *Leishmania*-infected individual. The DNA encoding Lbhsp83 is homologous to the gene encoding the eukaryotic 83 kD heat shock protein. The sequence of a DNA molecule encoding nearly all of Lbhsp83 is presented in SEQ ID NO:5, and the encoded amino acid sequence is provided in SEQ ID NO:6. Using the methods described below, Lbhsp83 has been found to stimulate proliferation, and a mixed Th1 and Th2 cytokine profile, in

joined by way of a linker sequence (e.g., Gly-Cys-Gly) that does not significantly diminish the immunogenic properties of the component polypeptides.

In general, Leishmania antigens having immunogenic properties, and DNA sequences encoding such antigens, may be prepared using any of a variety of procedures from one or more Leishmania species including, but not limited to, L. donovani, L. chagasi, L. infantum, L. major, L. amazonensis, L. braziliensis, L. panamensis, L. mexicana, L. tropica, and L. guyanensis. Such species are available, for example, from the American Type Culture Collection (ATCC), Rockville, MD. For example, peptides isolated from MHC class II molecules of macrophages infected with a Leishmania species may be used to rescue the corresponding Leishmania donor antigens. MHC class II molecules are expressed mainly by cells of the immune system, including macrophages. These molecules present peptides, which are usually 13-17 amino acids long, derived from foreign antigens that are degraded in cellular vesicles. The bound peptide antigens are then recognized by CD4 T-cells. Accordingly, foreign peptides isolated from MHC class II molecules of, for example, Leishmania-infected murine macrophages may be used to identify immunogenic Leishmania proteins.

Briefly, peptides derived from Leishmania antigens may be isolated by comparing the reverse phase HPLC profile of peptides extracted from infected macrophages with the profile of peptides extracted from uninfected cells. Peptides giving rise to distinct HPLC peaks unique to infected macrophages may then be sequenced using, for example, Edman chemistry as described in Edman and Berg, Eur J. Biochem, 80:116-132 (1967). A DNA fragment corresponding to a portion of a Leishmania gene encoding the peptide may then be amplified from a Leishmania cDNA library using an oligonucleotide sense primer derived from the peptide sequence and an oligo dT antisense primer. The resulting DNA fragment may then be used as a probe to screen a Leishmania library for a full length cDNA or genomic clone that encodes the Leishmania antigen. Such screens may generally be performed using techniques well known to those of ordinary skill in the art, such as those described in Sambrook et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratories, Cold Spring Harbor, NY (1989).

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asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe; (4) lys, arg, his; and (5) phe, tyr, trp, his.

Variants may also (or alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the immunogenic properties, secondary structure and hydropathic nature of the polypeptide. For example, a polypeptide may be conjugated to a signal (or leader) sequence at the N-terminal end of the protein which co-translationally or post-translationally directs transfer of the protein. The polypeptide may also be conjugated to a linker or other sequence for ease of synthesis, purification or identification of the polypeptide (e.g., poly-His), or to enhance binding of the polypeptide to a solid support. For example, a polypeptide may be conjugated to an immunoglobulin Fc region.

A nucleotide "variant" is a sequence that differs from the recited nucleotide sequence in having one or more nucleotide deletions, substitutions or additions. Such modifications may be readily introduced using standard mutagenesis techniques, such as oligonucleotide-directed site-specific mutagenesis as taught, for example, by Adelman et al. (*DNA*, 2:183, 1983). Nucleotide variants may be naturally occurring allelic variants, or non-naturally occurring variants. Variant nucleotide sequences preferably exhibit at least about 70%, more preferably at least about 80% and most preferably at least about 90% identity to the recited sequence. Such variant nucleotide sequences will generally hybridize to the recited nucleotide sequence under stringent conditions. As used herein, "stringent conditions" refers to prewashing in a solution of 6X SSC, 0.2% SDS; hybridizing at 65 °C, 6X SSC, 0.2% SDS overnight; followed by two washes of 30 minutes each in 1X SSC, 0.1% SDS at 65 °C and two washes of 30 minutes each in 0.2X SSC, 0.1% SDS at 65 °C.

"Polypeptides" as described herein also include combination polypeptides. A "combination polypeptide" is a polypeptide comprising at least one of the above immunogenic portions and one or more additional immunogenic *Leishmania* sequences, which are joined via a peptide linkage into a single amino acid chain. The sequences may be joined directly (i.e., with no intervening amino acids) or may be

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sequence is an antigen that contains, within its full length sequence, the recited sequence. The native antigen may, or may not, contain additional amino acid sequence.

An immunogenic portion of a *Leishmania* antigen is a portion that is capable of eliciting an immune response (*i.e.*, cellular and/or humoral) in a presently or previously *Leishmania*-infected patient (such as a human or a dog) and/or in cultures of lymph node cells or peripheral blood mononuclear cells (PBMC) isolated from presently or previously *Leishmania*-infected individuals. The cells in which a response is elicited may comprise a mixture of cell types or may contain isolated component cells (including, but not limited to, T-cells, NK cells, macrophages, monocytes and/or B cells). In particular, immunogenic portions are capable of inducing T-cell proliferation and/or a dominantly Th1-type cytokine response (*e.g.*, IL-2, IFN-γ, and/or TNF-α production by T-cells and/or NK cells; and/or IL-12 production by monocytes, macrophages and/or B cells). Immunogenic portions of the antigens described herein may generally be identified using techniques known to those of ordinary skill in the art, including the representative methods provided herein.

The compositions and methods of the present invention also encompass variants of the above polypeptides. A polypeptide "variant," as used herein, is a polypeptide that differs from the native antigen only in conservative substitutions and/or modifications, such that the ability of the polypeptide to include an immune response is retained. Polypeptide variants preferably exhibit at least about 70%, more preferably at least about 90% and most preferably at least about 95% identity to the identified polypeptides. Alternatively, such variants may be identified by modifying one of the above polypeptide sequences and evaluating the immunogenic properties of the modified polypeptide using, for example, the representative procedures described herein.

A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide chemistry would expect the secondary structure and hydropathic nature of the polypeptide to be substantially unchanged. In general, the following groups of amino acids represent conservative changes: (1) ala, pro, gly, glu, asp, gln,

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Figure 29 illustrates the effectiveness of immunization with MAPS-1A plus adjuvant in conferring protection against infection (as measured by footpad swelling) in a murine leishmaniasis model system, as compared to the administration of adjuvant alone.

Figures 30A and B illustrate the proliferation of murine lymph node cultures stimulated with either LcgSP8, LcgSP10 or LcgSP3.

# DETAILED DESCRIPTION OF THE INVENTION

As noted above, the present invention is generally directed to compositions and methods for preventing, treating and detecting leishmaniasis, as well as for stimulating immune responses in patients. The compositions of the subject invention include polypeptides that comprise at least an immunogenic portion of a *Leishmania* antigen, or a variant of such an antigen that differs only in conservative substitutions and/or modifications. In one preferred embodiment, compositions of the present invention include multiple polypeptides selected so as to provide enhanced protection against a variety of *Leishmania* species.

Polypeptides within the scope of the present invention include, but are not limited to, polypeptides comprising immunogenic portions of *Leishmania* antigens comprising the sequences recited in SEQ ID NO:2 (referred to herein as M15), SEQ ID NO:4 (referred to herein as Ldp23), SEQ ID NO:6 (referred to herein as Lbhsp83), SEQ ID NO:8 (referred to herein as Lt-210), SEQ ID NO:10 (referred to herein as LbeIF4A), SEQ ID NO: 20 (referred to herein as Lmsp1a), SEQ ID NO: 22 (referred to herein as Lmsp9a), SEQ ID NOs: 24 and 26 (referred to herein as MAPS-1A), and SEQ ID NO: 36-42, 49-53 and 55. As used herein, the term "polypeptide" encompasses amino acid chains of any length, including full length proteins (*i.e.*, antigens), wherein the amino acid residues are linked by covalent bonds. Thus, a polypeptide comprising an immunogenic portion of one of the above antigens may consist entirely of the immunogenic portion, or may contain additional sequences. The additional sequences may be derived from the native *Leishmania* antigen or may be heterologous, and such sequences may (but need not) be immunogenic. An antigen "having" a particular

Figure 19 presents a comparison of a Lbhsp83 sequence (SEQ ID NO:6) with homologous sequences from L. amazonensis (Lahsp83) (SEQ ID NO:16), T. cruzi (Tchsp83) (SEQ ID NO:17) and humans (Huhsp89) (SEQ ID NO:18).

Figure 20 illustrates the reactivity of rabbit sera raised against soluble Leishmania antigens with Leishmania promastigote lysate (lane 1) and soluble Leishmania antigens (lane 2).

Figure 21 shows the cDNA and predicted amino acid sequence for the Leishmania antigen Lmsp1a.

Figure 22 shows a Southern blot of genomic DNA from L. major digested with a panel of restriction enzymes (lanes 1 to 7) and six other Leishmania species digested with PstI (lanes 8 to 13) probed with the full-length cDNA insert of Lmsp1a.

Figure 23 shows a Southern blot of genomic DNA from *L. major* digested with a panel of restriction enzymes, six other *Leishmania* species digested with PstI and the infectious pathogens *T. cruzi* and *T. brucei*, probed with the full-length cDNA insert of the *Leishmania* antigen MAPS-1A.

Figure 24 illustrates the proliferation of PBMC isolated from uninfected-individuals, patients with active mucosal leishmaniasis and patients post kala-azar infection, stimulated by MAPS-1A.

Figure 25 illustrates the proliferation of murine lymph node cultures stimulated by MAPS-1A.

Figure 26 illustrates the reactivity of MAPS-1A with sera from human leishmaniasis patients.

Figure 27 illustrates the reactivity of MAPS-1A with sera from mice immunized against and/or infected with leishmaniasis.

Figure 28 illustrates the effectiveness of immunization with either soluble Leishmania antigens or a mixture of Ldp23, LbeiF4A and M15 plus adjuvant in conferring protection against infection (as measured by footpad swelling) in a murine leishmaniasis model system, as compared to the administration of adjuvant alone.

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infected and control individuals in response to stimulation with parasite lysate or the indicated polypeptides.

Figure 11 illustrates the levels of IL-10 p40 (in pg/mL) in the supernatant of PBMC cultures from *L. braziliensis*-infected individuals and uninfected controls 72 hours following stimulation with parasite promastigate lysate (Lb), Lbhsp83a or Lbhsp83b.

Figure 12 presents the reactivities of sera from *L. braziliensis* infected-patients with representative polypeptides of the present invention in a standard ELISA. Values are expressed as absorbance at 405 nm.

Figures 13A and 13B illustrate the level of secreted IL-4 and IFN-γ (in pg/mL) stimulated in mouse lymph node cultures by the addition of representative polypeptides of the present invention.

Figure 14 shows the level of IFN-γ (in pg/mL) secreted by *Leishmania*-infected and uninfected human PBMC stimulated by the *Leishmania* antigen M15, as compared to the levels stimulated by *L. major* lysate and L-Rack, an antigen that does not appear to be recognized by *Leishmania*-infected humans.

Figure 15 shows the level of IFN-γ (in pg/mL) secreted by infected and uninfected human PBMC stimulated by soluble *Leishmania* antigens (S antigens), as compared to the levels stimulated by *L. major* lysate and L-Rack.

Figure 16 illustrates the proliferation of murine lymph node cultures stimulated by the addition of representative polypeptides of the present invention. Values are expressed as cpm.

Figure 17 shows the proliferation of human PBMC, prepared from Leishmania-immune and uninfected individuals, stimulated by M15 as compared to the proliferation stimulated by L. major lysate and L-Rack. Values are expressed as cpm.

Figure 18 illustrates the proliferation of human PBMC, prepared from Leishmania-infected and uninfected individuals, stimulated by soluble Leishmania antigens as compared to the proliferation stimulated by culture medium, L. major lysate and L-Rack. Values are expressed as cpm.

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and 3 refer to RNA obtained from promastigotes at the logarithmic growth phase, promastigotes at the stationary growth phase and amastigote forms, respectively.

Figure 5 shows a Western blot analysis of *L. donovani* promastigote antigens incubated with pre-immune rabbit serum (lane A) or with anti-Ldp23 rabbit antiserum (lane B).

Figure 6 illustrates the surface expression of Ldp23 on live *L. donovani* promastigotes. The dotted line shows the indirect immunofluorescence performed using pre-immune mouse serum and the solid line shows the result obtained with mouse anti-GST-Ldp23 antiserum. Fluorescence intensity was analyzed by FACScan.

Figure 7 shows the stimulation of Leishmania-specific T-cell proliferation by Ldp23. The results are presented as relative cell number as a function of fluorescence intensity. T-cells (10<sup>5</sup>/well) were purified from lymph nodes of BALB/c mice immunized in the foot pad with L. donovani promastigotes in CFA and were cultured with various concentrations of the purified recombinant Ldp23 in the presence of 2 x 10<sup>5</sup> Mitomycin C-treated normal BALB/c spleen mononuclear cells. Proliferation of T-cells was measured at 27 hours of culture. Values are expressed as cpm and represent the mean of [3H]TdR incorporation of triplicate cultures.

Figure 8 illustrates Ldp23-induced cytokine production by lymph node cells of BALB/c mice. Cultures were incubated with varying amounts of Ldp23 or *Leishmania* lysate, presented as μg/mL, and were assayed by ELISA for the production of interferon-γ (panel A) or interleukin-4 (panel B), both of which are shown as ng/mL.

Figure 9 shows the PCR amplification of cytokine mRNAs isolated from mucosal leishmaniasis (Panel A) and cutaneous leishmaniasis (panel B) patient PBMC before and after stimulation with representative polypeptides of the present invention. Lanes O and - indicate the level of PCR products at the initiation of culture and after 72 hours of culture, respectively, in the absence of added polypeptide; lanes Lb, 83a and 83b indicate the level of PCR products following culturing of PBMC with L. braziliensis lysate, and the Leishmania antigens Lbhsp83a and Lbhsp83b, respectively.

Figure 10 presents a comparison of the levels of interferon-γ (panel A) and TNF-α (panel B) in the supernatants of 72 hour PBMC cultures from *Leishmania*-

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above; and (b) an apparatus sufficient to contact the pharmaceutical composition with the dermal cells of a patient.

In further aspects, the present invention provides methods for stimulating a cellular and/or humoral immune response in a patient, comprising administering to a patient a pharmaceutical composition or vaccine as described above.

In a related aspect, methods are provided for treating a patient afflicted with a disease responsive to IL-12 stimulation, comprising administering to a patient a pharmaceutical composition or vaccine as described above.

These and other aspects of the present invention will become apparent upon reference to the following detailed description and attached drawings. All references disclosed herein are hereby incorporated by reference in their entirety as if each was incorporated individually.

#### BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the stimulation of proliferation of T-cells obtained from L. donovani-immunized BALB/c mice (represented by stimulation index) by L. donovani-infected macrophages after incubation for 24, 48 and 72 hours.

Figure 2 illustrates representative HPLC profiles of peptides isolated from MHC class II molecules of P388D1 macrophages. Panel A shows peptides isolated from uninfected macrophages and panel B shows peptides isolated from L. donovani infected macrophages. The arrows in panel B indicate peptide peaks present only in the infected macrophage preparation.

Figure 3 illustrates the expression and purification of the *Leishmania* antigen Ldp23 as a recombinant fusion protein. Panel A shows a Coomassie blue-stained SDS-PAGE gel of lysed *E. coli* without (lane 1) and with (lane 2) IPTG induction of Ldp23 expression. Arrow indicates the recombinant fusion protein. Panel B shows the fusion protein following excision from a preparative SDS-PAGE gel, electroelution, dialysis against PBS and analytical SDS-PAGE.

Figure 4 presents a Northern blot analysis of total RNA prepared from L.

30 donovani, L. major, L. amazonensis and L. pifanoi with a <sup>32</sup>P labeled Ldp23 gene. 1, 2

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sequence selected from the group consisting of SEQ ID Nos: 2, 4, 20, 22, 24, 26, 36-38, 41, 50-53 and 82. DNA sequences encoding the above polypeptides, recombinant expression vectors comprising these DNA sequences and host cells transformed or transfected with such expression vectors are also provided.

In related aspects, the present invention provides pharmaceutical compositions which comprise one or more of the polypeptides described herein, or a DNA molecule encoding such polypeptides, and a physiologically acceptable carrier. Vaccines which comprise one or more such polypeptides or DNA molecules, together with a non-specific immune response enhancer are also provided. In specific embodiments of these aspects, the *Leishmania* antigen has an amino acid sequence selected from the group consisting of SEQ ID Nos: 2, 4, 20, 22, 24, 26, 36-38, 41, 50-53 and 82.

In still further related embodiments, the pharmaceutical compositions and vaccines comprise at least two different polypeptides, each polypeptide comprising an immunogenic portion of a *Leishmania* antigen having an amino acid sequence selected from the group consisting of sequences recited in SEQ ID Nos: 2, 4, 6, 8, 10, 20, 22, 24, 26, 36-38, 41, 50-53, 82, and variants thereof that differ only in conservative substitutions and/or modifications. In other embodiments, the inventive pharmaceutical compositions comprise one or more of the inventive polypeptides in combination with a known *Leishmania* antigen.

In yet other related embodiments, the pharmaceutical compositions and vaccines comprise soluble *Leishmania* antigens.

In another aspect, the present invention provides methods for inducing protective immunity against leishmaniasis in a patient, comprising administering to a patient a pharmaceutical composition or vaccine as described above.

In further aspects, methods and diagnostic kits are provided for detecting Leishmania infection in a patient. The methods comprise: (a) contacting dermal cells of a patient with a pharmaceutical composition as described above; and (b) detecting an immune response on the patient's skin, therefrom detecting Leishmania infection in the patient. The diagnostic kits comprise: (a) a pharmaceutical composition as described

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Leishmaniasis is a serious problem in much of the world, including Brazil, China, East Africa, India and areas of the Middle East. The disease is also endemic in the Mediterranean region, including southern France, Italy, Greece, Spain, Portugal and North Africa. The number of cases of leishmaniasis has increased dramatically in the last 20 years, and millions of cases of this disease now exist worldwide. About 2 million new cases are diagnosed each year, 25% of which are visceral leishmaniasis. There are, however, no vaccines or effective treatments currently available.

Accurate diagnosis of leishmaniasis is frequently difficult to achieve. There are 20 species of Leishmania that infect humans, including L. donovani, L. chagasi, L. infantum, L. major, L. amazonensis, L. braziliensis, L. panamensis, L. mexicana, L. tropica, and L. guyanensis, and there are no distinctive signs or symptoms that unambiguously indicate the presence of Leishmania infection. Parasite detection methods have been used, but such methods are neither sensitive nor clinically practical. Current skin tests typically use whole or lysed parasites. Such tests are generally insensitive, irreproducible and prone to cross-reaction with a variety of other diseases. In addition, the preparations employed in such tests are often unstable. Thus, there is a need for improved methods for the detection of Leishmania infection.

Current experimental vaccines consisting of whole organisms have not proven effective in humans. Accordingly, there remains a need in the art for vaccines to prevent leishmaniasis in humans and dogs, and for improved therapeutic compositions for the treatment of leishmaniasis.

#### SUMMARY OF THE INVENTION

Briefly stated, the present invention provides compositions and methods for preventing, treating and detecting leishmaniasis, as well as for stimulating immune responses in patients. In one aspect, polypeptides are provided which comprise at least an immunogenic portion of a *Leishmania* antigen, or a variant of such an antigen that differs only in conservative substitutions and/or modifications. In specific embodiments of the invention, the *Leishmania* antigen comprises an amino acid

## LEISHMANIA ANTIGENS FOR USE IN THE THERAPY AND DIAGNOSIS OF LEISHMANIASIS

#### REFERENCE TO RELATED APPLICATIONS

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### TECHNICAL FIELD

The present invention relates generally to compositions and methods for preventing, treating and detecting leishmaniasis, and for stimulating immune responses in patients. The invention is more particularly related to polypeptides comprising an immunogenic portion of a *Leishmania* antigen or a variant thereof, and to vaccines and pharmaceutical compositions comprising one or more such polypeptides. The vaccines and pharmaceutical compositions may be used, for example, for the prevention and therapy of leishmaniasis, as well as for the detection of *Leishmania* infection.

#### 15 BACKGROUND OF THE INVENTION

Leishmania organisms are intracellular protozoan parasites of macrophages that cause a wide range of clinical diseases in humans and domestic animals, primarily dogs. In some infections, the parasite may lie dormant for many years. In other cases, the host may develop one of a variety of forms of leishmaniasis. For example, the disease may be asymptomatic or may be manifested as subclinical visceral leishmaniasis, which is characterized by mild symptoms of malaise, diarrhea and intermittent hepatomegaly. Patients with subclinical or asymptomatic disease usually have low antibody titers, making the disease difficult to detect with standard techniques. Alternatively, leishmaniasis may be manifested as a cutaneous disease, which is a severe medical problem but is generally self-limiting, or as a highly destructive mucosal disease, which is not self-limiting. Finally, and most seriously, the disease may be manifested as an acute visceral infection involving the spleen, liver and lymph nodes, which, untreated, is generally a fatal disease. Symptoms of acute visceral fever, leukopenia, anemia leishmaniasis include hepatosplenomegaly, hypergammaglobulinemia.

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AL	Albania	ES	Spain	LS	Lesotho	· sı	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania :	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
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ΑZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad .
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## WORLD INTELLECTUAL PROPERTY ORGANIZATION International Bureau



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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C12N 15/30, C07K 14/44, C12N 1	5/63
5/10, A61K 39/008, A61K 48/00	

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**08/798**,841 **08/920,6**09 12 February 1997 (12.02.97) US 27 August 1997 (27.08.97) US

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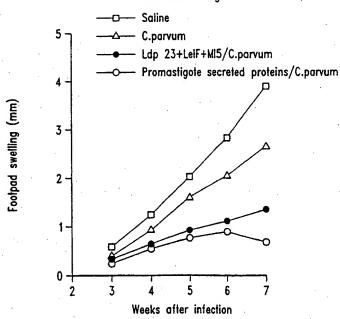
(74) Agents: MAKI, David, J. et al.; Seed and Berry LLP, 6300 Columbia Center, 701 Fifth Avenue, Seattle, WA 98104-7092 (US).

(54) Title: LEISHMANIA ANTIGENS FOR USE IN THE THERAPY AND DIAGNOSIS OF LEISHMANIASIS

#### (57) Abstract

Compositions and methods for preventing, treating and detecting leishmaniasis and stimulating immune responses in patients are disclosed. The compounds provided include polypeptides that contain at least an immunogenic portion of one or more *Leishmania* antigens, or a variant thereof. Vaccines and pharmaceutical compositions comprising such polypeptides, or DNA molecules encoding such polypeptides, are also provided and may be used, for example, for the prevention and therapy of leishmaniasis, as well as for the detection of *Leishmania* infection.

Protection against infection with L. major in BALB/c mice immunized leishmanial antigens.



Information on patent family members

Interna II Application No
PCT/US 98/03002

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9529239 A	02-11-1995	AU 2427695 A BR 9507505 A CA 2188543 A CN 1149887 A EP 0804581 A JP 9512172 T NO 964468 A	16-11-1995 02-09-1997 02-11-1995 14-05-1997 05-11-1997 09-12-1997 20-12-1996
WO 9711180 A	27-03-1997	AU 7242796 A EP 0854924 A	09-04-1997 29-07-1998

Idem as subject 18, for SEQ ID NO:85

28. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:86

29. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:87

Leishmania antigen comprising SEQ ID NO:20, variant thereof, DNA molecule encoding it, expression vector and host cell comprising the same, pharmaceutical compositions and vaccines comprising the polypeptide, vaccine comprising the DNA molecule, diagnostic kit

18. Claims: Partially 56-62

Pharmaceutical compositions and vaccines comprising a polypeptide comprising an immunogenic portion of a Leishmania antigen comprising SEQ ID NO:39 or a variant thereof

19. Claims: Partially 56-62
Idem as subject 18, for SEQ ID NO:42

20. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:55

21. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:61

22. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:62

23. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:80

24. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:81

25. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:83

26. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:84

27. Claims: Partially 56-62

Idem as subject 3, for SEQ ID NO:38

- 9. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:41
- 10. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:49
- 11. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:50
- 12. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:51
- 13. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:52
- 14. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55
  Idem as subject 3, for SEQ ID NO:53
- 15. Claims: Partially 13, 16-19, 25, 27-29, 31, 33, 35-40, 42-55

Idem as subject 3, for SEQ ID NO:82

16. Claims: 14, 15, and partially 16, 25, 27-29, 31, 33, 35-40, 42-55

Antigenic epitope of Leishmania antigen comprising SEQ ID NO:43, polypeptide comprising at least two such contiguous epitopes, DNA molecule encoding it, pharmaceutical compositions and vaccines comprising the polypeptide, vaccine comprising the DNA molecule, diagnostic kit

17. Claims: 20-24, 26, 32, 41 and partially 27-30, 33-39, 42-55

Polypeptide comprising an immunogenic portion of a

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-6, and partially 25, 27-29, 31, 33, 35-40, 42-55

Polypeptide comprising an immunogenic portion of a Leishmania antigen having SEQ ID NO:2, variant thereof, DNA molecule encoding it, expression vector and host cell comprising the same, pharmaceutical compositions and vaccines comprising the polypeptide, vaccine comprising the DNA molecule, diagnostic kit

2. Claims: 7-12, and partially 25, 27-29, 31, 33, 35-40, 42-55

Polypeptide comprising an immunogenic portion of a Leishmania antigen having SEQ ID NO:4, variant thereof, DNA molecule encoding it, expression vector and host cell comprising the same, pharmaceutical compositions and vaccines comprising the polypeptide, vaccine comprising the DNA molecule, diagnostic kit

3. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

Polypeptide comprising an immunogenic portion of a Leishmania antigen comprising SEQ ID NO:22, variant thereof, DNA molecule encoding it, expression vector and host cell comprising the same, pharmaceutical compositions, and vaccines comprising the polypeptide, vaccine comprising the DNA molecule, diagnostic kit

- Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55
   Idem as subject 3, for SEQ ID NO:24
- 5. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:26
- 6. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:36
- 7. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:37
- 8. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

Inter. .onal application No.

PCT/US 98/03002

International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:  Claims Nos.:	ox I Observations where ertain	n claims wire tound unse	archabi (Continuati noi	Rem 1 of mat sheet)
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Internat I Application No PCT/US 98/03002

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C.(Continua Category *	ation) DOCUMENTS CONSIDERED TO BE RELEVANT  Citation of document, with indication, where appropriate, of the relevant passages		Relevant to claim No.
	one control of the majorator, while appropriate, of the relevant passages		nesevant to claim No.
, X	wo 97 11180 A (CORIXA CORPORATION) 27 March 1997  see page 2, line 20 - page 3, line 37 see page 6, line 16 - page 7, line 20 see page 9, line 4 - line 19 see page 12, line 20 - page 14, line 17 see page 15, line 3 - page 16, line 26 see page 17, line 8 - line 30; claims; examples 1,7,8		1-6,25, 27-29, 31, 35-40, 42-55
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Internati Application No PCT/US 98/03002

A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12N15/30 C07K14/44 A61K39/008 C12N15/63 C12N5/10 A61K48/00

According to International Patent Classification (IPC) or to both national classification and IPC

#### **B. FIELDS SEARCHED**

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Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

Category *	Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No.
A	WO 95 29239 A (CORIXA CORPORATION) 2 1-6,25, November 1995 27-29, 31, 35-40, 42-55
	see page 2, line 17 - page 3, line 37 see page 5, line 8 - page 23, line 9; examples
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Date of the actual completion of the international search	Date of mailing of the international search report
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29 October 1998 (29.10.98)

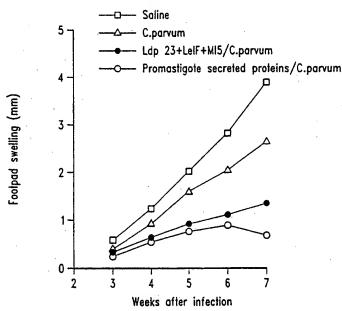
- (71) Applicant: CORIXA CORPORATION [US/US]; Suite 200, 1124 Columbia Street, Seattle, WA 98104 (US).
- (72) Inventors: REED, Steven, G.; 2843 122nd Place N.E., Bellevue, WA 98005 (US). CAMPOS-NETO, Antonio; 9308 Midship Court N.E., Bainbridge Island, WA 98110 (US). WEBB, John, R.; 15314 Silverfirs Drive, Everett, WA 98208 (US). DILLON, Davin, C.; 21607 N.E. 24th Street, Redmond, WA 98053 (US). SKEIKY, Yasir, A.; 8327 - 25th Avenue N.W., Seattle, WA 98107 (US).
- (74) Agents: MAKI, David, J. et al.; Seed and Berry LLP, 6300 Columbia Center, 701 Fifth Avenue, Seattle, WA 98104-7092 (US).

(54) Title: LEISHMANIA ANTIGENS FOR USE IN THE THERAPY AND DIAGNOSIS OF LEISHMANIASIS

#### (57) Abstract

Compositions and methods for preventing, treating and detecting leishmaniasis and stimulating immune responses in patients are disclosed. The compounds provided include polypeptides that contain at least an immunogenic portion of one or more Leishmania antigens, or a variant thereof. Vaccines and pharmaceutical compositions comprising such polypeptides, or DNA molecules encoding such polypeptides, are also provided and may be used, for example, for the prevention and therapy of leishmaniasis, as well as for the detection of Leishmania infection.

Protection against infection with L. major in BALB/c mice immunized leishmanial antigens.



### Proliferation rLcgSP Antigens

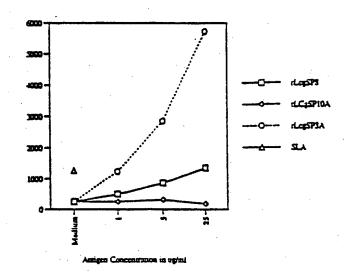


FIGURE 30A

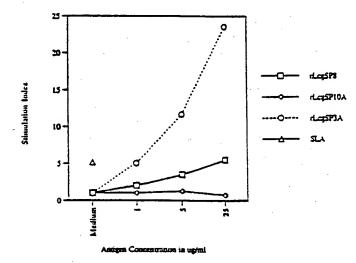


Figure 30B

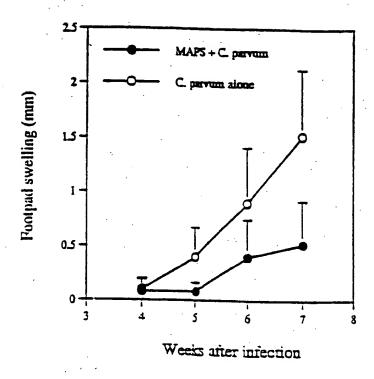


FIG. 29

27/29

FIG. 28

Protection against infection with L. major in BALB/c mice immunized leishmanial antigens.

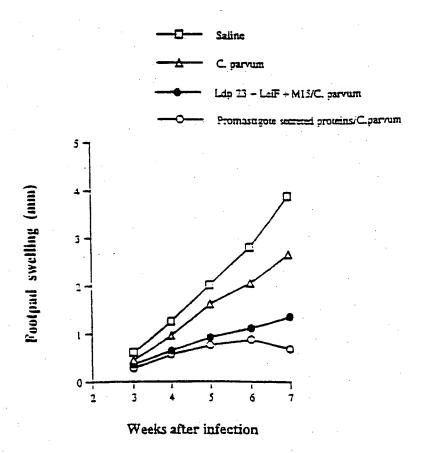
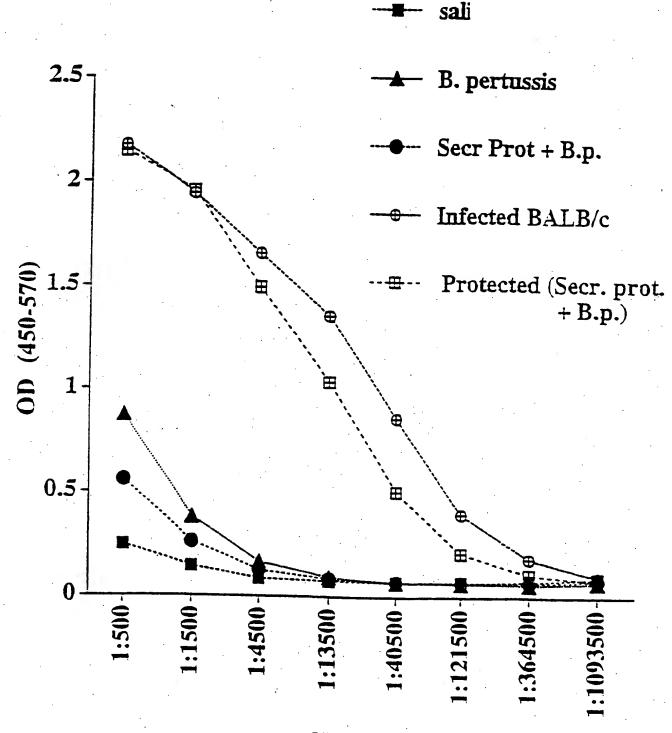


Figure 28

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# Antibody Response of Immunized/II BALB/c Mice to MAPS



serum dilution

ELISA analysis of MAPS-specific antibody titre human leishmaniasis patient sera (8/27/96)

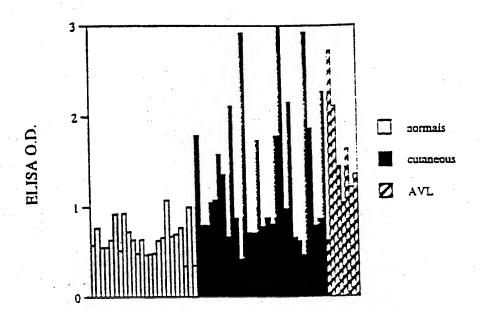


FIG. 26

Proliferative Responses of L. major-infected BALE (20 days post-infection) to recombinant MAPS protein

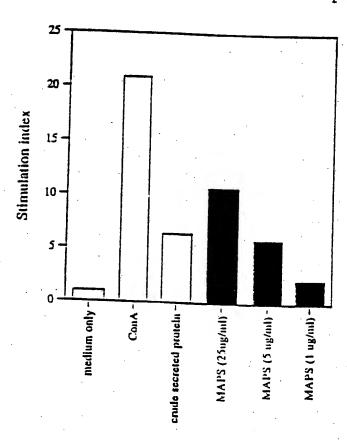
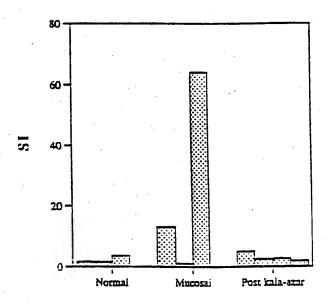


FIG. 25

FIG. 24



PBMC donor

Proliferative response of human PBMC to MAPS

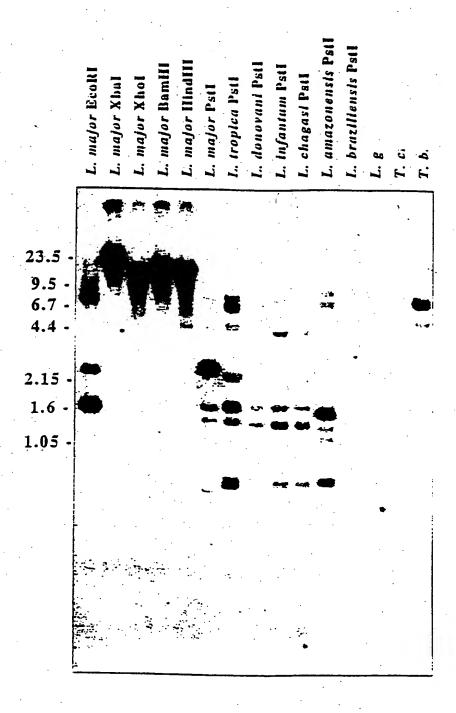
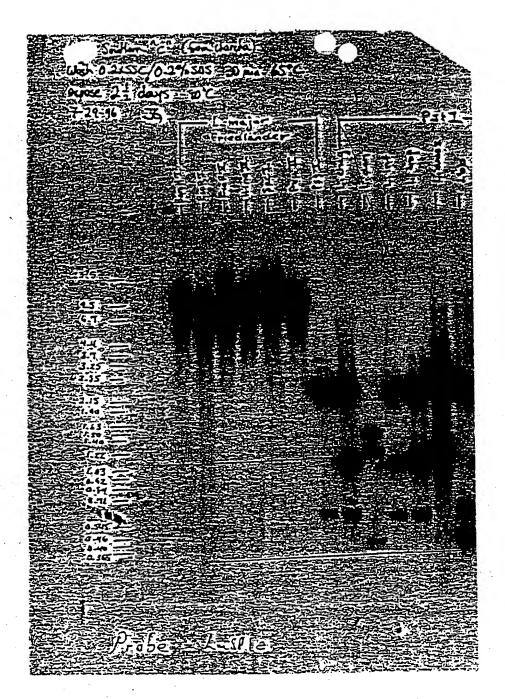
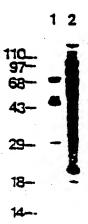


FIG. 23



## 20/24

	GETTTCTGTACTTTATTGC	TTCCAGCCTTTATTCACTCTTCGATTTCCTCTAACACCATGTCCTCGAGCGCACCTTTATTGCCGTC	- 100
5- Adaptor	Spliced-leader	S-UT	100
3 - Adaptor	1 Spilead-Idadai 1	M S S E R T F L A V	
	: Ter vererserffsttes	CGAGATCATCGCCCGCTTCGAGCGCAAGGGCTACAAGCTGGTCGCCTTGAAGATACTGCAGCCGACGA	
ICCCOUNCESCO	11 GEAGERED GET. CO. 1 GG	CHARLET COLOURS I CONSESCANDOS I ACTUANO	- 200
		E E E A R F E R K G Y K L V A L K ! L Q P T	•
K P U G			
CAGCAGGCCCA	GGGTCACTATAAGGACCTT	TGCTCCAAGCCGTTTTTCCCGGCCCTTGTGAAGTACTTCTCCTCTGGCCCGATCGTGTGTATGGTGTG	- 300
		No.	•
E Q A Q	3 G H Y K O L	CSKPFFPALVKYFSSGPLVCHVW	•
GAGGG TAAGAAC	STGGTGAAGAGCGGCCGCC	TGCTGCTCGCCGCGACGAACCCGGCCGACTCACAGGCCCGGCACGATCCGTGGCGACTTTGCCGTGGAT	- 400
E G K N	Y Y K S G R	V L L G A T N P A O S Q P G T I R G O F A V O	
COSCOCIONACÓ	TGTGCCACGGGTCCGACTC	DOTTODAGDTOGAGGEGGGGGGAGATTTGGTTTTGGTTGAGAGGCGGGAGAGAGCGGAGAGAGGTGGAGATGGAGATGGAGAGAGA	:
			- 500 •
r G R N	v c H G S O S	VESAEREIAFWFXAOEIASWTS	
		TTGC3GACAC3C7TTGAGGAC3TAGCTGTACCSSSAATGAATTCTTCTCGAAAACCACATCATAAGC	
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s v s c	1 Y S .	<b>३-</b> -पर	-
TOTTAAGAGGTT	'ATTTTTCTTGATCGATGCC	ADTDDDDTATTATAADAACAACAATAACAATAAACAACTAACAACAACAACAACAA	700
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			_ 300
-41646675.60		7.117	-
		3- UT	<b>-</b> -
		T- UT TTTTTTTTTDTDTDTDTDADGCDAAACDACAGAGAGAGAGAGAGAGAAACAACAACAACAACAACAAC	_ 300 _ _
			- - - -
TGCAAAGTTTT	AAASDESTABTSSTSAATAT	TTYTTTTTDTDTTDDTDTDADDCDTTTTCTTTCDTAAACDAAACDAAATACAAAAAAAA	т - - -
TGCAAAGTTTT	AAASDESTABTSSTSAATAT	AAACAACAACAACCOCCA TACACCAAGAGCAAATGCTTTCTTCTGCGGGACTGCTCTTCTTTTTTTT	- - aòi
TGCAAAGTTTT	AAASDESTABTSSTSAATAT	TTYTTTTTDTDTTDDTDTDADDCDTTTTCTTTCDTAAACDAAACDAAATACAAAAAAAA	т - - -
FGCAAAGTTTT	FA FAAC TOOTGA TOGGCAAA	AAACAACAACAACCOCCA TACACCAAGAGCAAATGCTTTCTTCTGCGGGACTGCTCTTCTTTTTTTT	т - - -



HTETFAFQAEINQLHSLIINTFYSHKEIFLANDVISUASDACDKIRYQDA.R.CE	
ADLVIIII.GTTARSGTKAFHEALEAGGDHSHIGGEGVGFYSAYLVANNÁVTVYSKHHSDEAY-WESSAGGTFTTTSVQESDHKÄGTSTTLIII.KEDQQEYLEE 184 .E	
RRVKELIKKUISEFIGYDIELHVEKTAEKEVTDEDEEEDESKKRSCUDEGEPKVEBVTEGG-ED-KKKKTKKVKEVKKT-YBVKNKUKPLHTRD 274	
TKDVTKEEYNAFYKAIBHDWEUTAATKHPSVEGQI.EPRAIAFVPKHAPFHHFBPHKKRIMIKLYVHHVFIHHHCEDLCPDWLUFVKGVVDSEDLFLHISR 374 P	
ENLOQUKILKVIRKUIVKKCLELFEBIAENKEDYKQFYEQFGRUIKLOIMEDTANRKLHELLHFYSTESGEEHTTLKUYVTHHKPEQKSIYYITGDSKK 414	٠.
KI.ESSPFIEKARRCOLEVLFHTEPIDEYVHQQVKDFEDKKFACI.TKEGVIIFEESEEKKQHEEKKAACEKI.CKTHKEVLGDKVEKVTVSERLI.TSPCILV 574TQ.K.R.F	• •
ТSEFGWSANNEQ1HRNQALÂD38HAQYHVSKKTHEVHPUHF11KELRRNVKANERHKAVKDI.VF1.LFUTSL.TSUFQL.NDPTGYAERINNHKLGI.SI.DE 674	
EEEEVA-EAPPAEAAPAEVTAGTSSHEQVD 703 1.Dhuph3EE.V.AV.T	

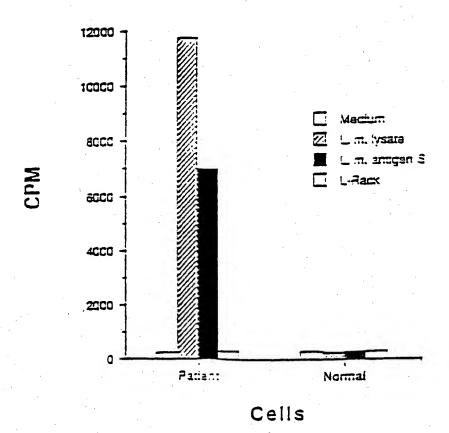


FIG. 18

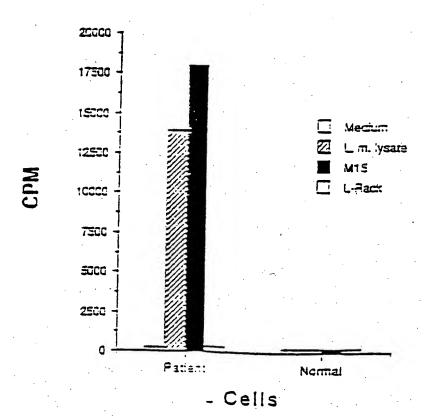
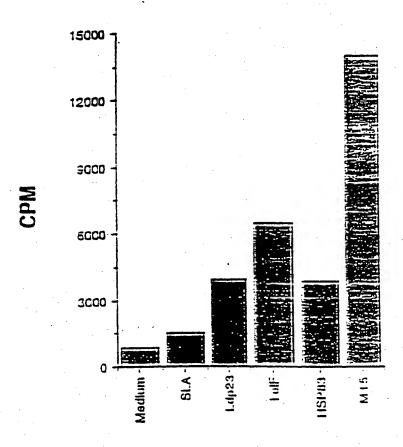


FIG. 17



Antigen (10 µg/ml)

FIG. 16

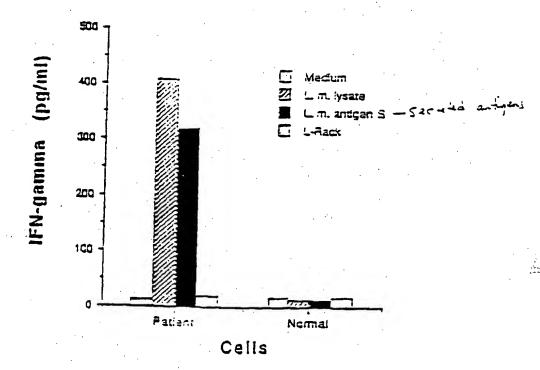


FIG. 15

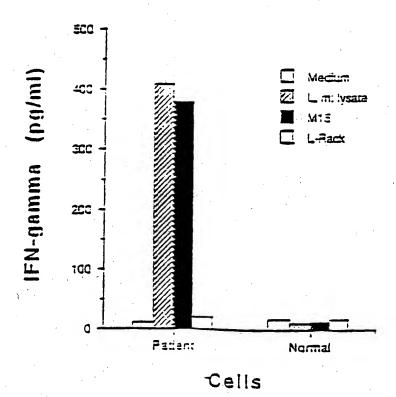
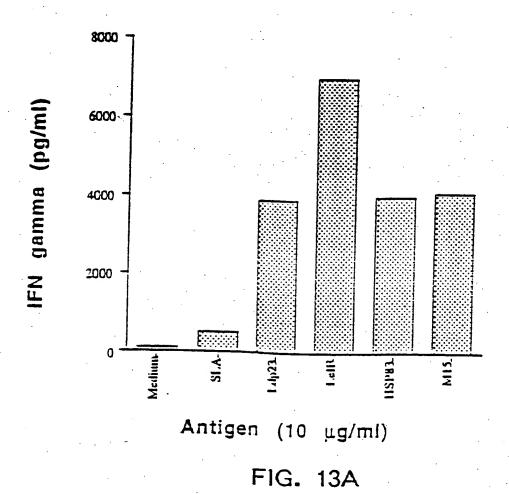


FIG. 14



IL-4 (pg/ml)

Madlum
SIA
Lap23
Lap23
ILSP03
M15-

Antigen (10 µg/ml)

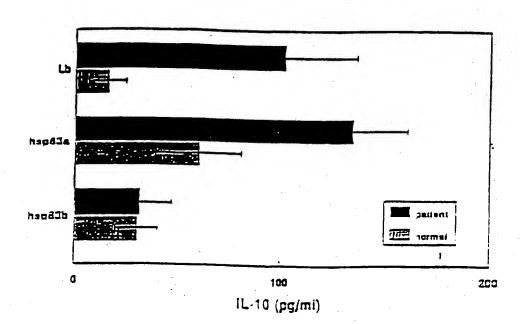


FIG. 11

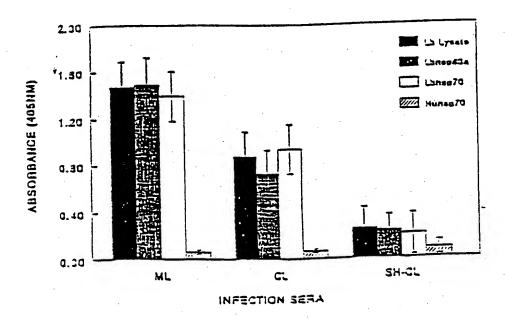
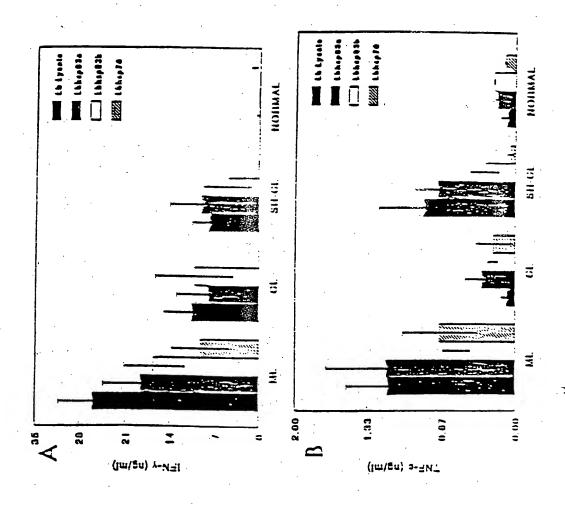


FIG. 12





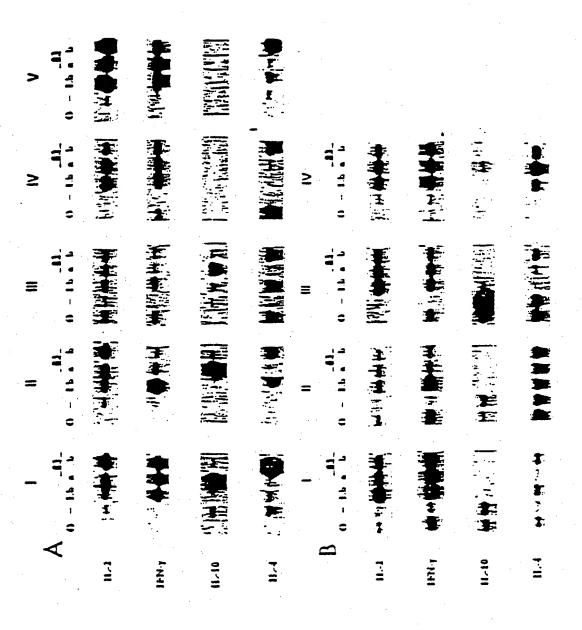


FIG. 9

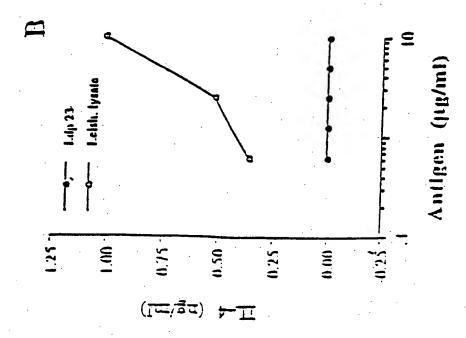
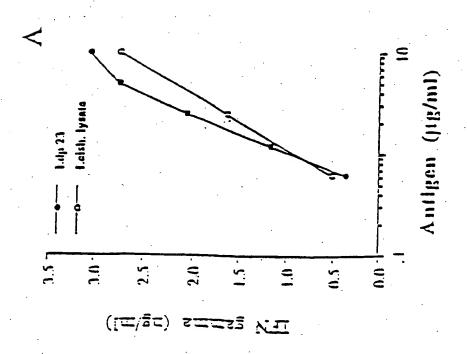


FIG. 8



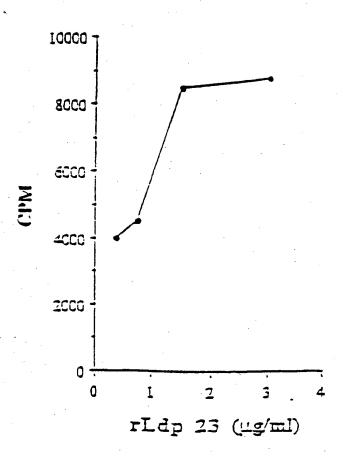


FIG. 7

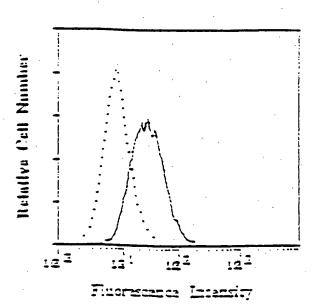
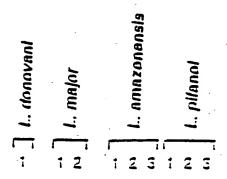


FIG. 6



FIG. 5



Wails -

# · 11 1112=11

C.24 -

FIG. 4

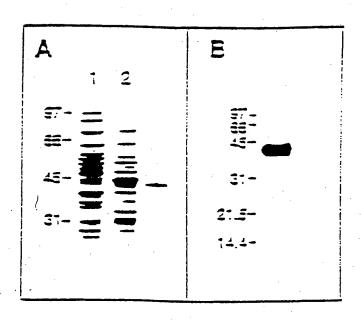


FIG. 3

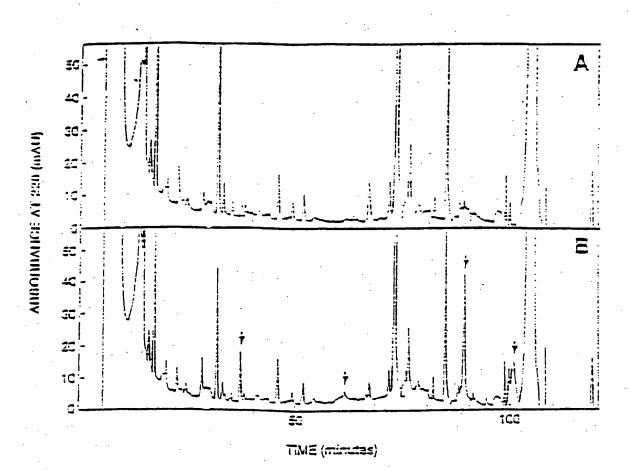
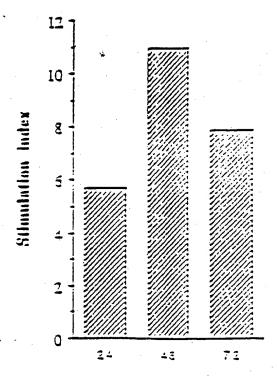


FIG. 2



Time of infection (hours)

FIG. 1

- 63. A diagnostic kit comprising:
- (a) a pharmaceutical composition according to any one of claims 56 and 57; and
- (b) apparatus sufficient to contact dermal cells of a patient with the pharmaceutical composition.

and/or modifications, wherein said antigen comprises an amino acid sequence selected from the group consisting of SEQ ID NO:39, 42, 55, 61, 62, 80, 81, and 83-87.

- 57. A pharmaceutical composition according to claim 56 further comprising a K39 *Leishmania* antigen.
- 58. A vaccine comprising a polypeptide and a non-specific immune response enhancer, the polypeptide comprising an immunogenic portion of a *Leishmania* antigen or a variant of said antigen that differs only in conservative substitutions and/or modifications, wherein said antigen comprises an amino acid sequence selected from the group consisting of SEQ ID NO:39, 42, 55, 61, 62, 80, 81 and 83-87.
- 59. A pharmaceutical composition according to any one of claims 56 and 57, for use in the manufacture of a medicament for inducing protective immunity against leishmaniasis in a patient.
- 60. A vaccine according to claim 58, for use in the manufacture of a medicament for inducing protective immunity against leishmaniasis in a patient.
- 61. A pharmaceutical composition according to any one of claims 56 or 57, for use in a method for detecting *Leishmania* infection in a patient.
- 62. A pharmaceutical composition according to claim 56, and a composition comprising a K39 *Leishmania* antigen, for use in a method for detecting *Leishmania* infection in a patient.

- 48. A pharmaceutical composition according to any one of claims 25, 26, 27, 29 and 30, for use in the manufacture of a medicament for stimulating a cellular and/or humoral immune response in a patient.
- 49. The composition of claim 48 wherein said response is a Th1 immune response.
- 50. The composition of claim 48 wherein said response is IL-12 production.
- 51. A vaccine according to any of claims 31-34, 40 and 41, for use in the manufacture of a medicament for stimulating a cellular and/or humoral immune response in a patient.
  - 52. The vaccine of claim 51 wherein said response is a Th1 response.
  - 53. The method of claim 51 wherein said response is IL-12 production.
- 54. A pharmaceutical composition according to any one of claims 25, 26, 27, 29 and 30, for use in the manufacture of a medicament for treating a patient afflicted with a disease responsive to IL-12 stimulation.
- 55. A vaccine according to any one of claims 31-34. 40 and 41, for use in the manufacture of a medicament for treating a patient afflicted with a disease responsive to IL-12 stimulation.
- 56. A pharmaceutical composition comprising a polypeptide and a physiologically acceptable carrier, the polypeptide comprising an immunogenic portion of a *Leishmania* antigen or a variant of said antigen that differs only in conservative substitutions

- 41. A vaccine comprising a DNA molecule according to claim 21.
- 42. A pharmaceutical composition according to any one of claims 25, 26, 27, 29 and 30, for use in the manufacture of a medicament for inducing protective immunity against leishmaniasis in a patient.
- 43. The composition of claim 42 wherein the leishmaniasis is caused by a Leishmania species selected from the group consisting of L. donovani, L. chagasi, L. infantum, L. major, L. amazonensis, L. braziliensis, L. panamensis, L. tropica and L. guyanensis.
- 44. A vaccine according to any one of claims 31-34, 40 and 41, for use in a method for inducing protective immunity against leishmaniasis in a patient comprising administering.
- 45. The vaccine of claim 44 wherein the leishmaniasis is caused by a Leishmania species selected from the group consisting of L. donovani, L. chagasi, L. infantum, L. major, L. amazonensis, L. braziliensis, L. panamensis, L. tropica and L. guyanensis.
- 46. A pharmaceutical composition according to any one of claims 25, 26, 27, 29 and 30, for use in a method for detecting *Leishmania* infection in a patient.
  - 47. A diagnostic kit comprising:
- (a) a pharmaceutical composition according to any one of claims 25, 26, 27, 29 and 30; and
- (b) apparatus sufficient to contact dermal cells of a patient with the pharmaceutical composition.

(h) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:10, or a variant of said antigen that differs only in conservative substitutions and/or modifications:

and a non-specific immune response enhancer.

- 34. A vaccine comprising soluble *Leishmania* antigens and a non-specific immune response enhancer.
- 35. A vaccine according to any one of claims 31, 32, 33 and 34 wherein the non-specific immune response enhancer is an immunogenic portion of a *Leishmania* antigen having the amino acid sequence recited in SEQ ID NO:10, or a variant of said antigen that differs only in conservative substitutions and/or modifications.
- 36. A vaccine according to any one of claims 31, 32 and 33, further comprising soluble *Leishmania* antigens.
- 37. A vaccine according to claim 36 wherein the non-specific immune response enhancer is an immunogenic portion of a *Leishmania* antigen having the amino acid sequence recited in SEQ ID NO:10, or a variant of said antigen that differs only in conservative substitutions and/or modifications.
- 38. A vaccine according to any one of claims 31, 32, 33 and 34, further comprising a delivery vehicle.
- 39. The vaccine of claim 38 wherein the delivery vehicle is a biodegradable microsphere.
- 40. A vaccine comprising a DNA molecule according to any one of claims 3, 9 and 16.

- 28. A pharmaceutical composition according to any one of claims 25, 26 and 27, further comprising soluble *Leishmania* antigens.
- 29. A pharmaceutical composition according to any one of claims 25, 26 and 27, further comprising a K39 Leishmania antigen.
- 30. A pharmaceutical composition comprising soluble *Leishmania* antigens and a physiologically acceptable carrier.
- 31. A vaccine comprising a polypeptide according to any one of claims 1, 7, 13 and 15, and a non-specific immune response enhancer.
- 32. A vaccine comprising a polypeptide according to claim 20 and a non-specific immune response enhancer.
- 33. A vaccine comprising at least two different polypeptides selected from the group consisting of:
  - (a) a polypeptide according to claim 1;
  - (b) a polypeptide according to claim 7;
  - (c) a polypeptide according to claim 13;
  - (d) a polypeptide according to claim 15
  - (e) a polypeptide according to claim 21;
- (f) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:6, or a variant of said antigen that differs only in conservative substitutions and/or modifications;
- (g) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:8, or a variant of said antigen that differs only in conservative substitutions and/or modifications; and

- 23. An expression vector comprising the DNA molecule of claim 21.
- 24. A host cell transformed or transfected with the expression vector of claim 23.
- 25. A pharmaceutical composition comprising a polypeptide according to any one of claims 1, 7, 13 and 15, and a physiologically acceptable carrier.
- 26. A pharmaceutical composition comprising a polypeptide according to claim 20 and a physiologically acceptable carrier.
- 27. A pharmaceutical composition comprising at least two different polypeptides selected from the group consisting of:
  - (a) a polypeptide according to claim 1;
  - (b) a polypeptide according to claim 7;
  - (c) a polypeptide according to claim 13
  - (d) a polypeptide according to claim 15;
  - (e) a polypeptide according to claim 20;
- (f) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:6, or a variant of said antigen that differs only in conservative substitutions and/or modifications;
- (g) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:8, or a variant of said antigen that differs only in conservative substitutions and/or modifications; and
- (h) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:10, or a variant of said antigen that differs only in conservative substitutions and/or modifications;

and a physiologically acceptable carrier.

- 16. An isolated DNA molecule comprising a nucleotide sequence encoding a polypeptide according to claims 13 or 15.
- 17. The DNA molecule of claim 16 wherein the nucleotide sequence is selected from the group consisting of:
- (a) sequences recited in SEQ ID NO: 21, 23, 25, 29-31, 34, 45-48 and 74; and
- (b) DNA sequences that hybridize to a nucleotide sequence complementary to a sequence recited in SEQ ID NO: 21, 23, 25, 29-31, 34, 45-48 and 74 under moderately stringent conditions.
  - 18. An expression vector comprising the DNA molecule of claim 16.
- 19. A host cell transformed or transfected with the expression vector of claim 18.
- 20. A polypeptide comprising an immunogenic portion of a *Leishmania* antigen or a variant of said antigen that differs only in conservative substitutions and/or modifications, wherein said antigen comprises an amino acid sequence provided in SEQ ID NO: 20.
- 21. An isolated DNA molecule comprising a nucleotide sequence encoding a polypeptide according to claim 20.
- 22. The DNA molecule of claim 21 wherein the nucleotide sequence is selected from the group consisting of:
  - (a) a sequence recited in SEQ ID NO: 19; and
- (b) DNA sequences that hybridize to a nucleotide sequence complementary to the sequence of SEQ ID NO: 19 under moderately stringent conditions.

- 8. The polypeptide of claim 7, comprising amino acids 1-175 of SEQ ID NO:4.
- 9. An isolated DNA molecule comprising a nucleotide sequence encoding a polypeptide according to claim 7.
- 10. The DNA molecule of claim 9 wherein the nucleotide sequence is selected from the group consisting of:
  - (a) nucleotides 25 through 549 of SEQ ID NO:3; and
- (b) DNA sequences that hybridize to a nucleotide sequence complementary to nucleotides 25 through 549 of SEQ ID NO:3 under moderately stringent conditions.
- 11. A recombinant expression vector comprising the DNA molecule of claim 9.
- 12. A host cell transformed or transfected with the expression vector of claim 11.
- 13. A polypeptide comprising an immunogenic portion of a *Leishmania* antigen or a variant of said antigen that differs only in conservative substitutions and/or modifications, wherein said antigen comprises an amino acid sequence selected from the group consisting of sequences recited in SEQ ID NO: 22, 24, 26, 36-38, 41, 49-53 and 82.
- 14. An antigenic epitope of a *Leishmania* antigen comprising an amino acid sequence recited in SEQ ID NO:43.
- 15. A polypeptide comprising at least two contiguous antigenic epitopes according to claim 14.

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- 1. A polypeptide comprising an immunogenic portion of a *Leishmania* antigen having the amino acid sequence recited in SEQ ID NO:2, or a variant of said antigen that differs only in conservative substitutions and/or modifications.
- 2. The polypeptide of claim 1, comprising amino acids 1 564 of SEQ ID NO:2.
- 3. An isolated DNA molecule comprising a nucleotide sequence encoding a polypeptide according to claim 1.
- 4. The DNA molecule of claim 3 wherein the nucleotide sequence is selected from the group consisting of:
  - (a) nucleotides 421 through 2058 of SEQ ID NO:1; and
- (b) DNA sequences that hybridize to a nucleotide sequence complementary to nucleotides 421 through 2058 of SEQ ID NO:1 under moderately stringent conditions.
- 5. A recombinant expression vector comprising the DNA molecule of claim 3.
- 6. A host cell transformed or transfected with the expression vector of claim 5.
- 7. A polypeptide comprising an immunogenic portion of a *Leishmania* antigen having the amino acid sequence recited in SEQ ID NO:4, or a variant of said antigen that differs only in conservative substitutions and/or modifications.

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154

100

105

110

Glu Tyr Pro Asp Arg Ile Met Met Thr Phe Ser Val Ile Pro Ser Pro 115 120 125

Arg Val Ser Asp Thr Val Val Xaa Pro Tyr Asn Thr Thr Leu Ser Val 130 135 140

His Gln Leu Val Glu 145

- (2) INFORMATION FOR SEQ ID NO:87:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 69 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:87:

Asn Pro Leu Val Tyr Ala Tyr Val Asp Thr Asp Gly Gln His Glu Thr 1 5 10 15

Thr Phe Leu Ala Ile Pro Val Val Leu Gly Met Asn Gly Ile Glu Lys
20 25 30

Arg Leu Pro Ile Gly Pro Leu His Ser Thr Glu Glu Thr Leu Leu Lys 35 40 45

Ala Ala Leu Pro Val Ile Lys Lys Asn Ile Val Lys Gly Ser Glu Phe 50 55 60

Ala Arg Ser His Leu

- Gly Leu Lys Arg Leu Ala Lys Ser Asp Pro Leu Val Val Cys Ser Ile 260 265 270
- Glu Glu Ser Gly Glu His Ile Val Ala Gly Ala Gly Glu Leu His Leu 275 280 285
- Glu Ile Cys Leu Lys Asp Leu Gln Glu Asp Phe Met Asn Gly Ala Pro 290 295 300
- Leu Lys Ilè Ser Glu Pro Val Val Ser Phe Arg Glu Thr Val Thr Asp 305 310 315 320
- Val Ser Ser Gln Gln Cys Leu Ser Lys Ser Ala Asn Lys His Asn Arg 325 330 335
- Leu Phe Cys Arg Gly Ala Pro Leu Thr Glu Xaa Leu Ala Leu Ala Xaa 340 345 350
- Xaa Glu Gly Thr Ala Gly Pro Xaa Ala 355 360

#### (2) INFORMATION FOR SEQ ID NO:86:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 149 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:86:

- Arg Ile Asn Val Tyr Phe Asp Xaa Ser Thr Gly Gly Arg Tyr Val Pro 1 5 10 15
- Arg Ala Val Leu Met Asp Leu Glu Pro Gly Thr Met Asp Ser Val Arg
  20 25 30
- Ala Gly Pro Tyr Gly Gln Leu Phe Arg Pro Asp Asn Phe Ile Phe Gly
  35 40 45
- Gln Ser Gly Ala Gly Asn Asn Trp Ala Lys Gly His Tyr Thr Glu Gly 50 55 60
- Ala Glu Leu Ile Asp Ser Val Leu Asp Val Cys Arg Lys Glu Ala Glu 65 70 75 80
- Ser Cys Asp Cys Leu Gln Gly Phe Gln Leu Ser His Ser Leu Gly Gly 85 90 95
- Gly Thr Gly Ser Gly Met Gly Thr Leu Leu Ile Ser Xaa Leu Arg Xaa

# (D) TOPOLOGY: linear

(xi)	SEQ	UENC:	E DE	SCRI	PTIO	N: S	EQ I	D NO	:85:			-			
Lys 1	Lys	Trp	Ile	Lys 5	Gln	Glu	Thr	Asn	Ala 10	Asp	Gly	Glu	Arg	Val	Arg
Arg	Ala	Phe	Cys 20	Gln	Phe	Cys	Leu	Asp 25	Pro	Ile	Tyr	Gln	Ile 30	Phe	Asp
Ala	Val	Met 35	Asn	Glu	Lys	Lys	Asp 40	Lys	Val	Asp	Lys	Met 45	Leu	Lys	Ser
Leu	His 50	Val	Thr	Leu	Thr	Ala 55	Glu	Glu	Arg	Glu	Gln 60.	Val	Pro	Xaa	Lys
Leu 65	Leu	Lys	Thr	Val	Met 70	Met	Xaa	Phe	Leu	Pro 75	Ala	Ala	Glu	Thr	Leu 80
Leu	Gln	Met	Ile	Val 85	Ala	His	Leu	Pro	Ser 90	Pro	Lys	Lys	Ala	Gln 95	Ala
Tyr	Arg	Ala	Glu 100	Met	Leu	Tyr	Ser	Gly 105	Glu	Ala	Ser	Pro	Glu 110	Asp	Lys
Tyr	Phe	Met 115	Gly	Ile	Lys	Asn	Cys 120	Asp	Pro	Ala	Ala	Pro 125	Leu	Met	Leu
Tyr	Ile 130	Ser	Lys	Met	Val	Pro 135	Thr	Ala	Asp	Arg	Gly 140	Arg	Phe	Phe	Ala
Phe 145	Gly	Arg	Ile	Phe	Ser 150	Gly	Lys	Val	Arg	Ser 155	Gly	Gln	Lys	Val	Arg 160
Ile	Met	Gly	Asn	Asn 165	Tyr	Val	Tyr	Gly	Lys 170	Lys	Gln	Asp	Leu	Tyr 175	Glu
Asp	Lys	Pro	Val 180	Gln	Arg	Ser	Va1	Leu 185	Met	Met	Gly	Arg	Tyr 190	Gln	Glu
Ala	Val	Glu 195	Asp	Met	Pro	Cys	Gly 200	Asn	Val	Val	Gly	Leu 205	Val	Gly	Val
Asp	Lys 210	Tyr	Ile	Val	Lys	Ser 215	Ala	Thr	Ile	Thr	Asp 220	Asp	Gly	Glu	Ser
Pro 225	His	Pro	Leu	Arg	Asp 230	Met	Lys	Tyr	Ser	Val 235	Ser	Pro	Val	Val	Arg 240
Val	Ala	Val	Glu	Ala 245	Lys	Asn	Pro	Ser	Asp 250	Leu	Pro	Lys	Leu	Val 255	Glu

- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:84:

Leu Thr Glu Phe Gln Thr Asn Leu Val Pro Tyr Pro Arg Ile His Phe 1 5 10 15

Val Leu Thr Ser Tyr Ala Pro Val Val Ser Ala Glu Lys Ala Tyr His 20 25 30

Glu Gln Leu Ser Val Ala Asp Ile Thr Asn Ser Val Phe Glu Pro Ala 35 40 45

Gly Met Leu Thr Lys Cys Asp Pro Arg His Gly Lys Tyr Met Ser Cys 50 55 60

Cys Leu Met Tyr Arg Gly Asp Val Val Pro Lys Asp Val Asn Ala Ala 65 70 75 80

Ile Ala Thr Ile Lys Thr Lys Arg Thr Ile Gln Phe Val Asp Trp Cys
85 90 95

Pro Thr Gly Phe Lys Cys Gly Ile Asn Tyr Gln Pro Pro Thr Val Val 100 105 110

Pro Gly Gly Asp Leu Ala Lys Val Gln Arg Ala Val Cys Met Ile Ala 115 120 125

Asn Ser Thr Ala Ile Ala Glu Val Phe Ala Arg Ile Asp His Lys Phe 130 135 140

Asp Leu Met Tyr Ser Lys Arg Ala Phe Val His Trp Tyr Val Gly Glu
145 150 155 160

Gly Met Glu Glu Gly Glu Phe Ser Glu Ala Arg Glu Asp Leu Ala Ala 165 170 175

Leu Glu Lys Asp Tyr Glu Glu Val Gly Ala Glu Ser Ala Asp Asp Met 180 185 190

Gly Glu Glu Asp Val Glu Glu Tyr 195 200

#### (2) INFORMATION FOR SEQ ID NO:85:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 361 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:

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150

Leu Ser Lys Leu Thr Gly Val Phe Ala Pro Arg Pro Arg Pro Gly Pro 20 25 30

His Lys Leu Arg Glu Cys Leu Pro Leu Leu Val Ile Ile Arg Asn Arg 35 40 45

Leu Lys Tyr Ala Leu Asn Ala Arg Glu Gly Glu Met Ile Leu Arg Gln 50 55 60

Gly Leu Val His Val Asp Asn His Pro Arg Arg Asp Gly Lys Tyr Pro 65 70 75 80

Ala Gly Phe Met Asp Val Val Glu Ile Pro Lys Thr Gly Asp Arg Phe 85 90 95

Arg Leu Met Tyr Asp Val Lys Gly Arg Phe Ala Leu Val Asn Leu Ser 100 105 110

Glu Ala Glu Ala Gln Ile Lys Leu Met Lys Val Val Asn Leu Tyr Thr 115 120 125

Ala Thr Gly Arg Val Pro Val Ala Val Thr His Asp Gly His Arg Ile 130 135 140

Arg Tyr Pro Asp Pro His Thr Ser Ile Gly Asp Thr Ile Val Tyr Asn 145 150 155 160

Val Lys Glu Lys Lys Cys Val Asp Leu Ile Lys Asn Arg Gln Gly Lys
165 170 175

Ala Val Ile Val Thr Gly Gly Ala Asn Arg Gly Arg Ile Gly Glu Ile 180 185 190

Val Lys Val Glu Cys His Pro Gly Ala Phe Asn Ile Ala His Leu Lys 195 200 205

Asp Ala Ser Gly Ala Glu Phe Ala Thr Arg Ala Ala Asn Ile Phe Val 210 215 220

Ile Gly Lys Asp Leu Asn Asn Leu Gln Val Thr Val Pro Lys Gln Gln 225 230 235 240

Gly Leu Arg Met Asn Val Ile Gln Glu Arg Glu Glu Arg Leu Ile Ala 245 250 255

Ala Glu Ala Arg Lys Asn Ala Pro Ala Arg Gly Ala Arg Arg Ala Arg 260 265 270

Lys

#### (2) INFORMATION FOR SEQ ID NO:84:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 200 amino acids

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:82:

Met Ser Ile Ile Lys Glu Asp Asp Ala Val Gly Cys Tyr Met Thr Val

1 5 10 15

Thr Leu Val Asp Asp Thr Lys Val Glu Gly Thr Ile Phe Thr Tyr Asn 20 25 30

Ser Lys Glu Gly Ile Ile Val Leu Leu Ser Leu Arg Asp Asp Gln Thr 35 40 45

Asn Met Lys Leu Ile Arg Thr Pro Tyr Ile Lys Asp Phe Ser Leu Ser 50 55 60

His Ala Glu Glu Gly Ala His Leu Pro Pro Ala Leu Asp Ser Phe Asn 65 70 75 80

Glu Leu Pro Ser Met His Ala Gly Arg Asp Lys Ser Ile Phe Lys His 85 90 95

Ala Ser Thr Gln Leu Lys Asn Ala Glu Ala Asn Arg Glu Lys His Phe 100 105 110

Asn Ser Val Thr Thr Asp Thr Pro Ile Ala Thr Leu Asp Ala Tyr Leu 115 120 125

Lys Leu Leu Arg Leu Tyr Pro Leu Ile Glu Trp Asn Ser Asp Glu Gly
130 135 140

Val Ile Gln Val Ser Asp Thr Val Ile Val Val Gly Asp Pro Asp Trp 145 150 155 160

Arg Thr Pro Lys Ala Met Leu Val Asp Gly Ala Pro Glu Lys Asp Arg 165 170 175

Pro Leu Val Asp Arg Leu Gln Val Ala Leu Gly Asn Gly Lys Lys 180 185 190

#### (2) INFORMATION FOR SEQ ID NO:83:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 273 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:83:

Met Ala Lys Lys His Leu Lys Arg Leu Tyr Ala Pro Lys Asp Trp Met 1 5 10 15

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Gln	Val	Thr	Asp	Val 165	Val	Glu	Lys	Ala	Ala 170	Glu	His	Tyr	Lys	Val 175	
Pro	Val	Asp	Gly 180	Val	Leu	Ser	His	Met 185	Met	Lys	Arg	Tyr	Ile 190	Ile	Asp
Xaa	Tyr	Arg 195	Cys	Ile	Pro	Gln	Arg 200	Arg	Val	Ala	Glu	His 205	Met	Val	His
Asp	Tyr 210	Asp	Leu	Glu	Lys	Ala 215	Gln	Val	Trp	Thr	Leu 220	Asp	Ile	Val	Met
Thr 225	Ser	Gly	Lys	Gly	Lys 230	Leu	Lys	Glu	Arg	Asp 235	Ala	Arg	Pro	Cys	Val 240
Phe	Lys	Val	Ala	Leu 245	Asp	Ser	Asn	Tyr	Ser 250	Val	Lys	Met	Glu	Ser 255	Ala
Lys	Glu	Val	Gln 260	Lys	Glu	Ile	Asp	Ser 265	Xaa	Tyr	Ala	Thr	Phe 270	Pro	Phe
Ala	Ile	Arg 275	Asn	Leu	Ğlu	Ala	Lys 280	Lys	Ala	Arg	Leu	Gly 285	Leu	Asn	Glu
Met	Ala 290	Lys	His	Gly	Ala	Val 295	Ile	Pro	Tyr	Pro	Ile 300	Leu	Phe	Glu	Lys
Glu 305	Gly	Glu	Val	Val	Ala 310	His	Phe	Lys	Ile	Thr 315	Val	Leu	Ile	Ser	Asn 320
Lys	Lys	Ile	Glu	Pro 325	Ile	Thr	Gly		1330	Pro	Gln	Lys	Ala	Pro 335	Ala
Leu	Glu	Pro	Tyr 340	Thr	Asp	Glu	Met	Leu 345	Leu	Ala	Thr	Asn	Lys 350	Leu	Phe
Ala	Val	Ala 355	Arg	Glu	Glu	Gly	Gly 360	Glu	Val	Asp	Gly	Arg 365	Gly	Ile	Arg
Asp	Ala 370	Val	Leu	Arg	Ala	Phe 375	Val	Gly	Val	Arg	Leu 380	Leu			

# (2) INFORMATION FOR SEQ ID NO:82:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 191 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

Tyr Val Asn Asp Val Met Glu Arg Ile Cys Thr Glu Ala Ala Ser Ile 50 55 60

Val Arg Ala Asn Lys Lys Arg Thr Leu Gly Ala Arg Glu Val Gln Thr 65 70 75 80

Ala Val Arg Ile Val Leu Pro Ala Glu Leu Ala Lys His Ala Met Ala 85 90 95

Glu Gly Thr Lys Ala Val Ser Ser Ala Ser Arg 100 105

#### (2) INFORMATION FOR SEQ ID NO:81:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 381 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:81:

Asp Glu Glu Glu Glu Asp Thr Thr Ile Asn Asn Ser Asp Val Val Val 1 5 10 15

Arg Tyr Lys Lys Ala Ala Thr Trp Cys Asn Glu Thr Leu Arg Val Leu 20 25 30

Ile Asp Ala Thr Lys Pro Gly Ala Lys Val Cys Asp Leu Cys Arg Leu 35 40 45

Gly Asp Asp Thr Ile Thr Ala Xaa Val Lys Thr Met Phe Lys Gly Thr 50 55 60

Glu Lys Gly Ile Ala Phe Pro Thr Cys Ile Ser Val Asn Asn Cys Val 65 70 75 80

Cys His Asn Ser Pro Gly Val Ser Asp Glu Thr Thr Gln Gln Glu Ile 85 90 95

Ala Met Gly Asp Val Val His Tyr Asp Leu Gly Ile His Val Asp Gly
100 105 110

Tyr Cys Ala Val Val Ala His Thr Ile Gln Val Thr Glu Asp Asn Glu
115 120 125

Leu Gly Lys Asp Glu Lys Ala Ala Arg Val Ile Thr Ala Ala Tyr Asn 130 135 140

Ile Leu Asn Thr Ala Leu Arg Gln Met Arg Pro Gly Thr Thr Ile Tyr 145 150 155 160

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. 140	
ACCTTCTCCG TCATCCCGTC CCCCCGCGTG TCGGATACCG TTGTGGANCC GTACAACACG	420
ACCCTCTCTG TGCACCAGCT CGTGGAA	447
(2) INFORMATION FOR SEQ ID NO:79:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 375 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:79:	
GTAACCCGCT GGTGTACGCA TATGTAGACA CAGACGGGCA GCACGAGACG ACGTTCCTCG	60
CGATCCCTGT GGTGCTTGGC ATGAATGGAA TCGAGAAGCG CCTGCCGATT GGTCCGCTGC	120
ACTCGACGGA GGAAACGCTG CTGAAGGCGG CACTGCCGGT GATCAAGAAG AATATCGTGA	1,80
AGGGCAGCGA GTTCGCGCGC TCACACCTGT AGCACCTCAG CTTTTTTTT TTGCGTTAAA	240
CGGGCGTGGG AAGCACCTCG ATACTTCGCT TCGCGCTGAC GGACCCGCAC GACATCGTTC	300
GTCATCCCCC TCCCCCTCTT CGGCCCTATA CGCATGAAGG AGTGGAATTA TGCAACAGCA	360.
TGTTNATATC AAGTG	375
(2) INFORMATION FOR SEQ ID NO:80:	
(i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 107 amino acids  (B) TYPE: amino acid  (C) STRANDEDNESS:  (D) TOPOLOGY: linear	
	٠
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:80:	
Met Ala Ser Ser Arg Lys Ala Ser Asn Pro His Lys Ser His Arg Lys  1 5 10 15	
Pro Lys Arg Ser Trp Asn Val Tyr Val Gly Arg Ser Leu Lys Ala Ile 20 25 30	
Asn Ala Gln Met Ser Met Ser His Arg Thr Met Lys Ile Val Asn Ser	

45

35

GATGCTGTAC	TCTGGCGAGG	CGTCGCCGGA	GGACAAGTAC	TTCATGGGTA	TCAAGAACTG	360
CGACCCCGCT	GCGCCGCTCA	TGCTGTACAT	CAGCAAGATG	GTGCCGACGG	CCGACCGCGG	420
CCGCTTCTTC	GCCTTTGGCC	GCATCTTCTC	CGGTAAGGTG	CGCAGCGGCC	AGAAGGTGCG	480
CATCATGGGT	AACAACTACG	TCTACGGCAA	GAAGCAGGAC	CTGTACGAGG	ACAAGCCTGT	540
GCAGCGCTCC	GTGCTGATGA	TGGGCCGCTA	CCAGGAGGCC	GTGGAGGACA	TGCCGTGCGG	600
TAACGTGGTG	GGCCTTGTGG	GCGTGGACAA	GTACATCGTG	AAGTCCGCGA	CGATCACGGA	660
CGATGGCGAG	AGCCCGCACC	CGCTGCGCGA	CATGAAGTAC	TCTGTGTCGC	CCGTCGTGCG	720
TGTGGCCGTG	GAGGCGAAGA	ACCCGTCCGA	CCTGCCGAAG	CTTGTGGAGG	GCCTGAAGCG	780
CCTTGCCAAG	TCCGACCCGC	TGGTGGTGTG	CAGCATTGAG	GAGTCTGGCG	AGCACATTGT	840
TGCCGGCGCT	GGCGAGCTTC	ACCTTGAGAT	TTGCCTGAAG	GATCTCCAGG	AGGACTTCAT	900
GAACGGCGCG	CCGCTNAAGA	TCTCCGAGCC	GGTGGTGTCG	TTCCGCGAGA	CGGTGACGGA	960
TGTGTCGTCG	CAGCAGTGCC	TGTCGAAGTC	TGCGAACAAG	CACAACCGTC	TCTTCTGCCG	1020
CGGTGCGCCG	CTNACAGAGG	ANCTGGCGCT	GGCGATNGAN	GAAGGCACCG	CTGGTCCCGA	1080
NGCGGA						1086

### (2) INFORMATION FOR SEQ ID NO:78:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 447 base pairs
  - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:78:

CG	CATCAACG	TCTACTTCGA	TNAGTCGACG	GGAGGCCGCT	ACGTGCCGCG	CGCCGTGCTG	60
ΑΊ	GGACCTCG	AGCCCGGCAC	TATGGACTCC	GTTCGCGCCG	GCCCGTACGG	CCAGCTGTTC	120
CG	CCCGGACA	ACTTCATCTT	TGGTCAGTCC	GGCGCTGGCA	ACAACTGGGC	CAAGGGCCAC	180
T	CACTGAGG	GCGCGGAGCT	GATCGACTCC	GTGCTTGATG	TGTGCCGCAA	GGAGGCGGAG	240
AC	CTGCGACT	GCCTGCAGGG	CTTCCAGCTG	TCTCACTCCC	TCGGCGGCGG	CACGGGCTCC	300
GC	CATGGGCA	CGCTGCTCAT	TTCCAANCTG	CGCGANGAGT	ACCCGGACCG	GATCATGATG	360

# (D) TOPOLOGY: linear

(X1) S	SEQUENCE	DESCRIPTION:	SEQ	ID	NO:76:

CTGACGGAGT	TCCAGACGAA	CCTTGTGCCG	TACCCGCGCA	TCCACTTCGT	GCTGACAAGC	60
TACGCTCCGG	TGGTGTCTGC	CGAGAAGGCG	TACCACGAGC	AGCTNTCCGT	CGCGGACATC	120
ACGAACTCGG	TNTTTGAGCC	TGCTGGCATG	CTNACAAAGT	GCGATCCTCG	CCACGGCAAG	180
TACATGTCGT	GCTGCCTCAT	GTACCGCGGT	GATGTCGTGC	CGAAGGATGT	CAACGCCGCG	240
ATTGCGACGA	TCAAGACGAA	GCGCACAATT	CAGTTCGTGG	ACTGGTGCCC	GACCGGCTTC	300
AAGTGCGGCA	TCAACTACCA	GCCGCCGACC	GTTGTGCCCG	GCGGTGACCT	CGCGAAGGTG	3 6.0
CAGCGCGCCG	TGTGCATGAT	TGCCAACTCG	ACCGCGATCG	CTGAGGTGTT	TGCCCGCATC	420
GACCACAAGT	TCGACCTGAT	GTACAGCAAG	CGCGCGTTTG	TGCACTGGTA	CGTGGGTGAG	480
GGCATGGAGG	AGGGCGAGTT	CTCCGAGGCG	CGCGAGGATC	TCGCTGCGCT	GGAGAAGGAC	540
TACGAGGAGG	TTGGCGCCGA	GTCCGCCGAC	GACATGGGCG	AGGAGGACGT	CGAGGAGTAC	600
TAAGGTAGAC	TCGTGCCGCG	CGCTGATGAT	GTAGGTGCAC	GCGTGCGTGT	GCTGCAGCGG	660
AGCCGCCGCC	ACCGCGACTG	TGTGTGTG	CGCGCGTGAC	GACCGGCTCG	AG	712

# (2) INFORMATION FOR SEQ ID NO:77:

# (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1086 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:77:

CAAGAAGTGG	ATCAAGCAGG	AGACGAACGC	CGATGGCGAG	CGCGTGCGCC	GCGCGTTCTG	60
CCAGTTCTGC	CTAGACCCCA	TCTACCAGAT	CTTCGACGCT	GTGATGAACG	AGAAGAAGGA	120
CAAGGTGGAC	AAGATGCTCA	AGTCGCTGCA	CGTGACGCTN	ACGGCTGAGG	AGCGCGAGCA	180
GGTGCCGAAN	AAGCTTCTGA	AGACGGTGAT	GATGAANTTC	CTGCCGGCTG	CTGAGACGCT	240
GCTACAGATG	ATCGTGGCGC	ACCTGCCGTC	GCCCAAGAAG	GCGCAGGCGT	ACCGTGCGGA	300

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:75:

TTTCTC	STACT	TTATTGAACA	TCAGTAGAAC	ACGTTCTTCC	CGCAAAGATG	GCCAAGAAGC	60
ACCTC	AAGCG	CTTGTATGCG	CCCAAGGACT	GGATGCTGAG	CAAGCTGACC	GGCGTGTTCG	120
CGCCG	CGTCC	GCGTCCGGGT	CCGCACAAGC	TGCGCGAGTG	CCTGCCGCTN	CTGGTGATCA	180
TCCGC	AACCG	GCTGAAGTAC	GCGCTGAACG	CGCGCGAGGG	TGAGATGATC	CTGCGCCAGG	240
GTCTG	GTGCA	CGTGGACAAC	CACCCGCGCC	GCGACGGCAA	GTATCCCGCC	GGTTTCATGG	300
ACGTG	GTCGA	GATCCCGAAG	ACGGGCGACC	GCTTCCGCCT	GATGTACGAC	GTCAAGGGCC	360
GCTTC	GCGTT	GGTGAACCTG	TCCGAGGCGG	AGGCGCAGAT	CAAGCTGATG	AAGGTTGTGA	420
ACCTG'	TACAC	GGCCACCGGC	CGCGTGCCGG	TCGCTGTGAC	GCACGACGGC	CACCGCATCC	480
GCTAC	CCGGA	CCCGCACACC	TCCATTGGTG	ACACCATCGT	GTACAACGTC	AAGGAGAAGA	540
AGTGC	GTGGA	CCTGATCAAG	AACCGCCAGG	GCAAGGCCGT	GATCGTGACC	GGTGGCGCCA	600
ACCGC	GGCCG	CATCGGCGAG	ATCGTGAAGG	TGGAGTGCCA	CCCCGGTGCG	TTCAACATTG	660
CGCAC	CTGAA	GGACGCGTCC	GGCGCCGAGT	TCGCCACCCG	CGCCGCGAAC	ATCTTCGTGA	720
TCGGC	AAGGA	CCTGAACAAC	CTGCAGGTAA	CGGTGCCGAA	GCAGCAGGGC	CTGCGCATGA	780
ACGTG	ATCCA	GGAGCGCGAG	GAGCGCCTGA	TCGCGGCGGA	GGCCCGCAAG	AACGCGCCGG	840
CTCGT	GGTGC	CCGCAGGGCC	CGCAAGTGAG	GAGGCGATTA	CACGCATGCG	TGTTTGTGGC	900
TCTGA	AGCGA	CTTGGCGGGT	CGGCTGTGAG	GGTTTGAGAG	GAGGTGTGTG	ATGCGTGTGA	960
AGTCC	TTCTC	CGTTCTCAGC	TCTCTCTGTG	CTGTAGCTGT	GCCTTTCCCC	AGATCGCTTT	1020
ACCGC	ATTTG	CATACATCTG	TGTAGTCGCA	TGTGCGTGTT	TCTGTCTCTC	GGTGGGTCTC	1080
CCTCT	CCCTC	CCTTTCTGCC	TCTCTCTTTG	AGTGGGTGTG	CATGCGTCGC	GCGCGACGGG	1140
CTCCG	CTTNA	GTGATTCTCT	CGTGTTTTAN	GGCTGTTTTY	TTTCTYAGTT	NAGCGTTTTY	1200
GTTCA	TGATT	TCCTCAGACC	САААААААА	ААААААА			1238

### (2) INFORMATION FOR SEQ ID NO:76:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 712 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single

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TGTGTGTGTG	TGGGGGGGCG	TGTTACGAGT	ACAAAAGAGG	CTCGATCTTT	GCGATCTTTT	1380
CTTTCTGTAA	ACAGGAACAT	AAGTAACCAA	АААААААА	AAAAAACTCG	AG	1432

# (2) INFORMATION FOR SEQ ID NO:74:

# (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 873 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:74:

CTTTATTGTC	ATCACTGTAA	AGCACTGTTT	TTTCTTTCAC	TTTTTCTTGA	GTGTTTTCTT	60
CTATTCACCA	TGAGCATTAT	CAAGGAGGAC	GACGCCGTGG	GCTGCTACAT	GACGGTGACC	120
CTCGTGGACG	ACACCAAGGT	GGAGGGTACC	ATCTTCACCT	ACAATTCCAA	GGAGGGCATC	180
ATAGTACTCC	TGTCCCTCCG	CGACGATCAG	ACGAACATGA	AGCTAATCCG	CACTCCGTAC	240
ATCAAAGACT	TCAGCCTTTC	ACACGCTGAG	GAGGGAGCGC	ACCTGCCCCC	GGCACTGGAC	300
TCCTTCAACG	AGCTTCCGTC	CATGCACGCC	GGCCGCGACA	AGTCCATCTT	CAAGCACGCC	360
AGCACGCAGC	TCAAGAACGC	CGAGGCGAAC	CGCGAAAAGC	ACTTCAACTC	TGTCACGACC	420
GACACACCGA	TTGCCACACT	TGATGCGTAC	CTCAAGCTCC	TGCGGCTATA	CCCCTTAATT	480
GAGTGGAACA	GCGACGAGGG	TGTCATCCAG	GTCTCGGACA	CCGTCATTGT	CGTAGGAGAC	540
CCCGACTGGC	GGACGCCCAA	GGCAATGCTG	GTGGACGGCG	CCCCTGAGAA	GGACAGACCG	600
CTTGTAGATC	GCCTGCAGGT	TGCGCTCGGM	AACGGCAAGA	AGTĢATTCAG	TGTGTAGCGG	660
ACAGAACATC	GTGTGCTTGT	GTGTCTGTTT	GANGTTTGTT	TGTTTTCTCT	TTGTGGTACT	720
GCGTACGACG	GCGCCTTCTC	CCGGTGGTGG	GTGAGTCCAT	AAGCAGTTGA	GTTCTYGGTT	780
GTAGNAAVGC	CTYACYGCCG	ACCATATGGG	AGAGGGCGAA	CAAATNTTTG	ATAGAAGTTG	840
AAAATCCCAA	AGTYAAAAGA	AAAAAAAA	<b>AAA</b>			873

# (2) INFORMATION FOR SEQ ID NO:75:

### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1238 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(A) LENGTH: 1432 base pairs(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:73:

GATGAAGAAG AGGAGGACAC CACCATCAAC AACTCCGACG TGGTGGTGCG CTACAAGAAG 60 GCCGCAACGT GGTGCAATGA AACGTTGCGC GTGCTTATCG ATGCCACAAA ACCTGGCGCC 120 AAGGTGTGCG ACCTGTGCCG CCTCGGTGAT GACACCATCA CCGCCNAGGT CAAGACAATG 180 TTCAAAGGCA CGGAAAAAGG CATCGCTTTC CCGACCTGCA TCTCGGTCAA CAACTGCGTA 240 TGCCACAACA GCCCTGGCGT GTCGGACGAG ACGACGCAGC AAGAGATCGC GATGGGTGAC 300 GTCGTGCACT ACGACCTGGG CATCCACGTG GACGGCTACT GCGCCGTCGT CGCGCACACC 360 ATTCAGGTGA CAGAGGACAA TGAGCTTGGC AAGGACGAGA AGGCGGCGCG CGTCATTACA 420 GCGGCGTACA ACATCCTGAA CACGGCGCTG CGCCAGATGC GTCCCGGTAC GACCATCTAC 480 CAGGTGACAG ACGTAGTTGA GAAGGCTGCG GAGCACTACA AGGTGACTCC GGTAGACGGC 540 GTCCTCTCGC ATATGATGAA GCGCTACATC ATAGACNGAT ACCGCTGTAT CCCGCAGCGC 600 AGGGTCGCGG AGCACATGGT GCACGACTAC GATCTCGAGA AAGCGCAGGT GTGGACGCTA 660 GACATTGTCA TGACCTCCGG CAAGGGCAAG CTGAAGGAGC GCGATGCGCG GCCGTGCGTG 720 TTCAAGGTGG CTCTGGACTC CAACTACTCT GTGAAAATGG AAAGCGCGAA GGAGGTTCAG 780 AAGGAAATCG ACTCCNAGTA TGCCACCTTC CCCTTTGCCA TCCGCAACCT GGAGGCCAAG 840 AAGGCCCGCC TCGGTCTCAA CGAGATGGCG AAGCACGGTG CTGTCATCCC GTACCCTATT 900 CTCTTCGAAA AGGAAGGCGA GGTCGTCGCC CATTTCAAGA TTACGGTGCT CATCAGCAAC 960 AAGAAGATTG AGCCGATTAC CGGCCTGAAG CCGCAGAAGG CCCCGGCGCT CGAGCCATAC 1020 ACGGACGAGA TGCTGCTTGC GACGAACAAG CTCTTCGCTG TCGCTAGAGA AGAAGGCGGC 1080 GAAGTAGACG GCCGTGGCAT CCGTGACGCT GTACTGCGAG CTTTCGTAGG CGTACGCCTC 1140 TTGTGAGGCG TACACGTGTG CTGTTTGCGG ACGAGGAGGC ACCCATTCTG TTCCCCTTCT 1200 TCGCTAATCT TCGCGTTTCC TCTGACGCTG GCTTCTYTGC CGGAGTGTGG TGAGGCGCGT 1260 GGGGGAGAAA CGGCCCACTY GCATGCCTGT GCATACGCGA GCACGGTAGG GAGCGCGGTG 1320

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(xi)	SEQUENCE	DESCRIPTION:	SEQ	ID	NO:71	:
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Gly Cys Gly Pro Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp 1 10 15

Gly Pro Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp Gly Pro
20 25 30

Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp Gly 35 40 45

# (2) INFORMATION FOR SEQ ID NO:72:

# (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 664 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:72:

GCTGCAGGAA	TTCGGCACGA	GATTGCTTCC	CAGCCCACCT	TCGCTATCCA	GCCACTCTCG	60
CTCTTCTACA	TCTCCCACCC	CCTCACACCG	CCATGGCTTC	TTCCCGCAAG	GCTTCCAACC	120
CGCACAAGTC	GCACCGCAAG	CCGAAGCGCT	CGTGGAACGT	GTACGTGGGC	CGCTCGCTGA	180
AGGCGATCAA	CGCCCAGATG	TCGATGTCGC	ACCGCACGAT	GAAGATCGTG	AACTCGTACG	240
TGAACGACGT	GATGGAGCGC	ATCTGCACTG	AGGCCGCGTC	GATTGTTCGC	GCGAACAAGA	300
AGCGCACGTT	GGGTGCGCGC	GAGGTGCAGA	CGGCGGTGCG	CATTGTGCTG	CCGGCGGAGC	360
TCGCGAAGCA	TGCCATGGCT	GAGGGCACGA	AGGCCGTGTC	GAGCGCGTCC	CGCTAAAGCG	420
GCTTGCCGGA	TGCCGTGTGA	GTAGGAGGGT	GGCTTGCCGC	AAACGCTGAC	CTCGGCGATT	480
GCGGCGTGGC	GCTCCCCTTC	TCCTCCTTGT	CCGGCGGTGT	GTGTCATGCA	TTTGCGTGAC	540
TCCTCCCTCT	TATAGATGCA	AGCTTTTTTT	TTCTCTTGAC	GTTTTATTTT	CTCCTCCCC	600
TCCCTTAACG	TGAAGTGTAT	ATGANAGCGT	ACTGGACATG	AAAAAAAA	AAAANAAACT	660
CGAG	f	•	.•			664

# (2) INFORMATION FOR SEQ ID NO:73:

# (i) SEQUENCE CHARACTERISTICS:

Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp Gly Pro
20 25 30

Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp Gly 35 40 45

- (2) INFORMATION FOR SEQ ID NO:69:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 17 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:69:

Gly Cys Gly Pro Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp

1 10 15

Gly

- (2) INFORMATION FOR SEQ ID NO:70:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 31 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:70:

Gly Cys Gly Pro Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp 1 5 10 15

Gly Pro Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp Gly 20 25 30

- (2) INFORMATION FOR SEQ ID NO:71:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 45 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 17 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:
- Gly Cys Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp 1 5 10 15

Gly

- (2) INFORMATION FOR SEQ ID NO:67:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 31 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:
  - Gly Cys Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp 1 5 10 15
  - Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp Gly
    20 25 30
- (2) INFORMATION FOR SEQ ID NO:68:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 45 amino acids
      - (B) TYPE: amino acid
      - (C) STRANDEDNESS:
      - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:68:
  - Gly Cys Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp 1 5 10 15

Pro Lys Glu Asp Gly Xaa Thr Gln Lys Asn Asp Xaa Xaa Gly
1 5 10

- (2) INFORMATION FOR SEQ ID NO:64:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 7 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (ix) FEATURE:
    - (A) NAME/KEY: Modified-site
    - (B) LOCATION: 6
    - (D) OTHER INFORMATION: /note= "Xaa can be either His or

Arg"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:64:

Pro Lys Glu Asp Gly Xaa Thr 1 5

- (2) INFORMATION FOR SEQ ID NO:65:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 7 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (ix) FEATURE:
    - (A) NAME/KEY: Modified-site
    - (B) LOCATION: 5
    - (D) OTHER INFORMATION: /note= "Xaa can be either Gly or

Asp"

- (ix) FEATURE:
  - (A) NAME/KEY: Modified-site
  - (B) LOCATION: 6
  - (D) OTHER INFORMATION: /note= "Xaa can be either Asp or

Gly"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:65:

Gln Lys Asn Asp Xaa Xaa Gly

(2) INFORMATION FOR SEQ ID NO:66:

Gly Asp Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp 130 135 140

Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp Gly Pro 145 150 155 160

Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp Gly Pro Lys Glu 165 170 175

Asp Gly His Thr Gln Lys Asn Asp Gly Asp Gly Pro Lys Glu Asp Gly 180 185 190

Arg Thr Gln Lys Asn Asp Gly Gly Gly Pro Lys Glu Asp Glu Asn Leu 195 200 205

Gln Gln Asn Asp Gly Asn Ala Gln Glu Lys Asn Glu Asp Gly His Asn 210 215 220

Val Gly Asp Gly Ala Asp Gly Asp Gly Asp Asp Asp Gln Pro 225 230 235 240

Lys Glu Gln Val Ala Gly Asn 245

#### (2) INFORMATION FOR SEQ ID NO:63:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 14 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

#### (ix) FEATURE:

- (A) NAME/KEY: Modified-site
- (B) LOCATION: 6
- (D) OTHER INFORMATION: /note= "Xaa can be either His or

# (ix) FEATURE:

Arg"

- (A) NAME/KEY: Modified-site
- (B) LOCATION: 12
- (D) OTHER INFORMATION: /note= "Xaa can be either Gly or Asp"

#### (ix) FEATURE:

- (A) NAME/KEY: Modified-site
- (B) LOCATION: 13
- (D) OTHER INFORMATION: /note= "Xaa can be either Asp or Gly"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:

Met Gly Ser Ser Cys Thr Lys Asp Ser Ala Lys Glu Pro Gln Lys Arg

1 10 15

Ala Asp Asn Ile Asp Thr Thr Thr Arg Ser Asp Glu Lys Asp Gly Ile
20 25 30

His Val Gln Glu Ser Ala Gly Pro Val Gln Glu Asn Phe Gly Asp Ala 35 40 45

Gln Glu Lys Asn Glu Asp Gly His Asn Val Gly Asp Gly Ala Asn Asp 50 55 60

Asn Glu Asp Gly Asn Asp Asp Gln Pro Lys Glu Gln Val Ala Gly Asn 65 70 75 80

#### (2) INFORMATION FOR SEQ ID NO:62:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 247 amino acids
  - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:

Met Gly Ala Tyr Cys Thr Lys Asp Ser Ala Lys Glu Pro Gln Lys Arg
1 5 10 15

Ala Asp Asn Ile His Lys Thr Thr Glu Ala Asn His Arg Gly Ala Ala
20 25 30

Gly Val Pro Pro Lys His Ala Gly Gly Ala Met Asn Asp Ser Ala Pro 35 40 45

Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp Gly Pro Lys Glu 50 55 60

Asp Gly Arg Thr Gln Lys Asn Asp Asp Gly Gly Pro Lys Glu Asp Gly 65 70 75 80

His Thr Gln Lys Asn Asp Gly Asp Gly Pro Lys Glu Asp Gly Arg Thr 85 90 95

Gln Lys Asn Asn Gly Asp Gly Pro Lys Glu Asp Gly His Thr Gln Lys
100 105 110

Asn Asp Gly Asp Ala Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp 115 120 125

### AAGGAGCAGG TTGCCGGCAA CTAG

264

# (2) INFORMATION FOR SEQ ID NO:60:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 744 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:

ATGGGAGCCT	ACTGCACGAA	GGACTCCGCA	AAGGAGCCCC	AGAAGCGTGC	TGATAACATC	60
CATAAAACCA	CTGAGGCCAA	TCACAGAGGC	GCCGCCGGTG	TGCCCCGAA	GCACGCCGGC	120
GGTGCGATGA	ACGACTCTGC	CCCGAAGGAG	GATGGCCATA	CACAGAAAAA	TGACGGCGAT	180
GGCCCTAAGG	AGGACGGCCG	TACACAGAAA	AACGACGACG	GTGGCCCTAA	GGAGGACGGC	240
CATACACAGA	AAAATGACGG	CGATGGCCCT	AAGGAGGACG	GCCGTACACA	GAAAAATAAC	300
GGCGATGGCC	CTAAGGAGGA	CGGCCATACA	CAGAAAAATG	ACGGCGATGC	CCCTAAGGAG	360
GACGGCCGTA	CACAGAAAAA	TGACGCCGAT	GGCCCTAAGG	AGGACGGCCG	TACACAGAAA	420
AATGACGGCG	ATGGCCCTAA	GGAGGACGGC	CGTACACAGA	AAAATGACGG	CGATGGCCCT	480
AAGGAGGACG	GCCGTACACA	GAAAAATGAC	GGCGATGGCC	CTAAGGAGGA	CGGCCATACA	540
CAGAAAAATG	ACGGCGATGG	CCCTAAGGAG	GACGGCCGTA	CACAGAAAA	TGACGGCGGT	600
GGCCCTAAGG	AGGATGAGAA	TCTGCAGCAA	AACGATGGGA	ATGCGCAGGA	GAAGAACGAA	660
GATGGACACA	ACGTGGGGGA	TGGAGCTAAC	GGCAATGAGG	ATGGTAACGA	TGATCAGCCG	720
AAGGAGCAGG	TTGCCGGCAA	CTAG				744

# (2) INFORMATION FOR SEQ ID NO:61:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 80 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

(ii) MOLECULE	TYPE:	peptide
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(v) FRAGMENT TYPE: internal

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:

Pro Lys Glu Asp Gly His Ala 1 5

- (2) INFORMATION FOR SEQ ID NO:58:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 7 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
    - (v) FRAGMENT TYPE: internal.
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:

Pro Lys Asn Asp Asp His Ala 1 5

- (2) INFORMATION FOR SEQ ID NO:59:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 264 base pairs
    - (B) TYPE: nucleic acid
      - (C) STRANDEDNESS: single
      - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:

ATGCACCATC ATCACCATCA CATGGGAAGC TCCTGCACGA AGGACTCCGC AAAGGAGCCC 60
CAGAAGCGTG CTGATAACAT CGATACGACC ACTCGAAGCG ATGAGAAGGA CGGCATCCAT 120
GTCCAGGAGA GCGCCGGTCC TGTGCAGGAG AACTTCGGGG ATGCGCAGGA GAAGAACGAA 180
GATGGACACA ACGTGGGGGA TGGAGCTAAC GACAATGAGG ATGGTAACGA TGATCAGCCG 240

- Arg Leu Ser Asp Ala Cys Ser Val Pro Asn Cys Lys Lys Cys Glu Thr 180 185 190
- Gly Thr Ser Arg Leu Cys Ala Glu Cys Asp Thr Gly Tyr Ser Leu Ser 195 200 205
- Ala Asp Ala Thr Ser Cys Ser Ser Pro Thr Thr Gln Pro Cys Glu Val 210 215 220
- Glu His Cys Asn Thr Cys Val Asn Gly Asp Ser Thr Arg Cys Ala Tyr 225 230 235 240
- Cys Asn Thr Gly Tyr Tyr Val Ser Asp Gly Lys Cys Lys Ala Met Gln 245 250 255
- Gly Cys Tyr Val Ser Asn Cys Ala Gln Cys Met Leu Leu Asp Ser Thr 260 265 270
- Lys Cys Ser Thr Cys Val Lys Gly Tyr Leu Leu Thr Ser Ser Tyr Ser 275 280 285
- Cys Val Ser Gln Lys Val Ile Asn Ser Ala Ala Ala Pro Tyr Ser Leu 290 295 300
- Trp Val Ala Ala Ala Val Leu Leu Thr Ser Phe Ala Met His Leu Ala 305 310 315 320
- (2) INFORMATION FOR SEQ ID NO:56:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 14 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
    - (v) FRAGMENT TYPE: internal
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:

Pro Lys Glu Asp Gly His Ala Pro Lys Asn Asp Asp His Ala 1 5 10

- (2) INFORMATION FOR SEQ ID NO:57:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 7 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear

- (2) INFORMATION FOR SEQ ID NO;55:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 320 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (v) FRAGMENT TYPE: internal
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:

Val Leu Pro Asp Met Thr Cys Ser Leu Thr Gly Leu Gln Cys Thr Asp

10 15

Pro Asn Cys Lys Thr Cys Thr Thr Tyr Gly Gln Cys Thr Asp Cys Asn
20 25 30

Asp Gly Tyr Gly Leu Thr Ser Ser Ser Val Cys Val Arg Cys Ser Val
35 40 45

Ala Gly Cys Lys Ser Cys Pro Val Asp Ala Asn Val Cys Lys Val Cys 50 55 60

Leu Gly Gly Ser Glu Pro Ile Asn Asn Met Cys Pro Cys Thr Asp Pro 65 70 75 80

Asn Cys Ala Ser Cys Pro Ser Asp Ala Gly Thr Cys Thr Gln Cys Ala 85 90 95

Asn Gly Tyr Gly Leu Val Asp Gly Ala Cys Val Arg Cys Gln Glu Pro 100 105 110

Asn Cys Phe Ser Cys Asp Ser Asp Ala Asn Lys Cys Thr Gln Cys Ala 115 120 125

Pro Asn Tyr Tyr Leu Thr Pro Leu Leu Thr Cys Ser Pro Val Ala Cys
130 135 140

Asn Ile Glu His Cys Met Gln Cys Asp Pro Gln Thr Pro Ser Arg Cys 145 150 155 160

Gln Glu Cys Val Ser Pro Tyr Val Val Asp Ser Tyr Asp Gly Leu Cys 165 170 175

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:

						•
60	CCGGGCTGCA	AGTGGATCC	G CTCTAGAACT	GTGGCGGCCG	CTCCACCGCG	AAAGCTGGAG
120	CAGTGCACAG	GACCGGACTI	ACATGCTCGCT	CCCGACATGA	ACGAGTGCTG	GGAATTCGGC
180	GACGGCTACG	AGACTGCAAC	GTCAGTGCAC	ACAACTTACG	CAAGACCTGC	ACCCGAACTG
240	AGCTGCCCCG	GGGCTGCAAG	GCAGTGTAGC	TGCGTGCGCT	CTCCAGCGTT	GTCTCACCTC
300	AATATGTGCC	GCCGATCAAC	GCGGCAGCGA	GTGTGTCTCG	CGTCTGCAAA	TCGACGCTAA
360	ACTCAGTGCG	TGGCACGTGC	CCAGCGACGC	GCCAGCTGCC	CCCCAACTGC	CCTGCACCGA
420	AACTGCTTCA	CCAGGAGCCC	GTGTGAGATG	GACGGCGCCT	CGGTCTCGTG	CGAACGGCTA
480	CTCACCCCGC	GAACTACTAC	AATGTGCGCC	AAGTGCACAC	CGACGCGAAT	GCTGCGACAG
540	GACCCACAGA	CATGCAGTGC	TCGAGCACTG	GCCTGCAACA	CTCCCCGGTG	TCTTGACCTG
600	GACGGCCTCT	TGACAGCTAC	CCTACGTGGT	TGCGTGTCCC	CTGCCAGGAG	CGCCGTCGCG
660	GGTACCTCCA	GTGCGAGACC	ACTGCAAGAA	TCCGTGCCCA	CGATGCCTGC	GCAGGCTCTC
720	AGCTGCAGCA	CGACGCGACG	GTCTCTCCGC	ACCGGCTACA	CGAGTGCGAC	GGCTCTGCGC
780	GGCGATAGCA	ATGTGTGAAC	ACTGCAACAC	GAGGTGGAGC	GCAGCCGTGC	GTCCAACCAC
840	AAGGCCATGC	TGGCAAGTGC	ACGTCTCCGA	ACCGGCTACT	CTACTGCAAC	CCCGCTGTGC
900	AAGTGCTCCA	TGACAGCACC	GCATGCTGCT	TGCGCGCAGT	CGTGTCGAAC	AGGGCTGCTA
960	AAAGTCATCA	CGTCTCGCAG	CCTACAGTTG	CTCACGTCGT	AGGGTACCTG	CGTGCGTGAA
1020	ACCTCTTTTG	CGTGCTCCTC	TGGCCGCCGC	TCTCTGTGGG	CGCGCCCTAC	ACAGTGCGGC
1080	TCTCCAACAT	CCACTCTCAT	GCGAACAACC	GCAGCGGCAT	AGCATAGTGC	CCATGCACCT
1140	ACACACGCAC	CCCCACACAC	AGCACCCCCT	ACAGCGGGGC	ACACACACAG	GTGCATACAC
1200	GCGCTGGCGC	TTTCTCTCGT	TTCGCATTTC	CTTTCCTCGN	TCTTGTTCTT	TTCCCCCTTG
1260	CTCTCTCTCG	CTCTCTCTCT	AACCTCTATT	CCCTCCCCCT	CACGTCGCTC	CGGCCTCCTG
1320	CACTGCCACA	CGTGGGCGGA	TCCTTGCTCG	CCTTTTCTGA	TGCTTCTTAC	CCGGCATCAT
1380	TCACTTCATT	TCCCTCCCTA	.GGCGCAGGCA	GTGTTTAAAC	GCAGACACAC	GTCCCACAGC
1440	CCTTCCGGGC	CATCGGCCGC	CGCCCTCCCC	AGTCGCACAC	CCACTCACCA	TCTCCTAAAG
1500	CGCATGTGTA	GCAGCTCACT	CTCGTTCCTG	TGTGCTCGAC	GGAATGGGTG	GCAGCTGTGC

Asp	Ile	Arg	Ala 100	Arg	Lys	Gly	Ser	Val 105	Pro	Cys	Gly	Ala	Ser 110	Ser	Met

Glu Asn Ser Thr Pro Leu Asp Ser Ala Val Glu Pro Phe Glu Ser Asp 115 120 125

Asp Gly Asp Asp Val Val Asp Lys Thr Gly Leu Asp Pro Asn Glu Leu 130 135 140

Gln Gly Ile Ile Pro Arg Ala Cys Thr Asp Leu Phe Asp Gly Leu Arg 145 150 155 160

Ala Lys Arg Ala Lys Asp Ser Asp Phe Thr Tyr Arg Val Glu Val Ser 165 170 175

Tyr Tyr Glu Ile Tyr Asn Glu Lys Val Phe Asp Leu Ile Arg Pro Gln 180 185 190

Arg Asn Thr Asp Leu Arg Ile Arg Asn Ser Pro Asn Ser Gly Pro Phe 195 200 205

Ile Glu Gly Leu Thr Trp Lys Met Val Ser Lys Glu Glu Asp Val Ala 210 215 220

Arg Val Ile Arg Lys Gly Met Gln Glu Arg His Thr Ala Ala Thr Lys 225 230 235 240

Phe Asn Asp Arg Ser Ser Arg Ser His Ala Ile Leu Thr Phe Asn Ile 245 250 255

Val Gln Leu Ser Met Asp Asp Ser Asp Asn Ala Phe Gln Met Arg Ser 260 265 270

Lys Leu Asn Leu Val Asp Leu Ala Gly Ser Glu Arg Thr Gly Ala Ala 275 280 285

Gly Ala Glu Gly Asn Glu Phe His Asp Gly Val Lys Ile Asn His Ser 290 295 300

Leu Thr Val Leu Gly Arg Val Ile Asp Arg Leu Ala Asp Leu Ser Gln 305 310 315 320

Asn Lys Gly Gly

#### (2) INFORMATION FOR SEQ ID NO:54:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1585 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)

- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania chagasi
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:

Val Arg Glu Thr Arg Thr Cys Met Leu Leu Val Glu Gly Ala Glu Leu 1 5 10 15

Val Pro Leu Met His Lys His Glu Val Val His Cys Leu Val Arg His
20 25 30

Phe Pro Glu His Asn Glu Gln Arg His His Thr Ala Ser Asn Ser Arg 35 40 45

Gln Ser Glu Cys Asp Ala His Thr Lys Ser Lys Val Val Asp Gln
50 55 60

- (2) INFORMATION FOR SEQ ID NO:53:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 324 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania chagasi
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:

Phe Arg Gly Ile Pro Arg Asn Ser Val Arg Arg Gly Thr Arg Thr Gly
1 5 10 15

Val Asp Ser Asp Ala Thr Ser Ser Thr Thr Ala Ala Tyr Asp Gly Ala  $20^{\circ}$  25 30

Gly Ser Ala Pro Val Met Val Asp Ala Asn Val Ser His Pro Pro Tyr 35 40 45

Ala Gly His Asp Gln Val Tyr Met His Val Gly Lys Pro Ile-Val Gly 50 55 60

Asn Thr Leu Asp Gly Tyr Asn Gly Cys Val Phe Ala Tyr Gly Xaa Thr 65 70 75 80

Gly Ser Gly Lys Thr Phe Thr Met Leu Gly Tyr Ala Pro Ser Thr Xaa 85 90 95

465

470

475

- Ala Cys Ala Pro Val Ser Ser Ala Pro Ser Trp Lys Ser Ser Arg Arg 485 490 495
- Arg Ser Gly Ile Met Gly Ala Cys Thr Arg Arg Thr Thr Ser 500 505 510
- (2) INFORMATION FOR SEQ ID NO:51:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 107 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania chagasi
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:
  - Gly Arg Arg Met Ala Ile His Arg Lys Met Thr Ala Met Ala Leu Arg

    1 10 15
  - Arg Thr Ala Val His Arg Lys Thr Thr Thr Val Ala Leu Arg Arg Thr 20 25 30
  - Ala Ile His Arg Lys Met Thr Ala Met Ala Leu Arg Arg Thr Ala Val
  - His Arg Lys Ile Thr Ala Met Ala Leu Arg Arg Thr Ala Ile His Arg 50 55 60
  - Lys Met Thr Ala Met Pro Leu Arg Arg Thr Ala Val His Xaa Lys Met 65 70 75 80
  - Thr Ala Met Ala Leu Arg Arg Thr Ala Val His Arg Lys Met Thr Ala 85 90 95
  - Met Ala Leu Arg Xaa Thr Pro Tyr Thr Glu Lys
    100 105
- (2) INFORMATION FOR SEQ ID NO:52:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 63 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide

Gly	Ser	Tyr	Leu 180		Ala	Gly	Thr	Ser 185		i Sei	Pro	туг	Glu 190		ı Lei
Met	Asn	Glu 195	Tyr	Glu	Pro	Gly	11e 200		Thr	Glm	Lys	205	Asp	Glu	ı Val
Tyr	Ala 210	Asn	Val	Lys	Ser	Trp 215		Pro	Gln	Leu	Leu 220		Asp	Ile	· Va]
Gln 225	Lys	Gln	Ser	Gly	Glu 230	Ser	Val	Ile	Ala	Phe 235		His	Lys	Phe	Pro 240
Gln	Asp	Lys	Gln	Glu 245	Ala	Leu	Cys	Lys	Glu 250		Met	Lys	Ile	Trp 255	
Phe	Asp	Thr	Asp 260	Ala	Gly	Arg	Leu	Asp 265	Val	Ser	Pro	His	Pro 270	Phe	Thr
Gly	Met	Thr 275	Lys	Glu	Asp	Cys	Arg 280		Thr	Thr	Asn	Tyr 285	Ile	Glu	Asp
Thr	Phe 290	Val	Gln	Ser	Leu	Tyr 295	Gly	Val	Ile	His	Glu 300	Ser	Gly	His	Gly
Lys 305	Tyr	Glu	Gln	Asn	Cys 310	Gly	Pro	Arg	Glu	His 315	Ile	Thr	Gln	Pro	Val 320
Cys	Asn	Ala	Arg	Ser 325	Leu	Gly	Leu	His	Glu 330	Ser	Gln	Ser	Leu	Phe 335	Ala
Glu	Phe	Gln	Ile 340	Gly	His	Ala	Thr	Pro 345	Phe	Ile	Asp	Tyr	Leu 350	Thr	Thr
Arg	Leu	Pro 355	Glu	Phe	Phe	Glu	Ala 360	Gln	Pro	Ala	Phe	Ser 365	Gln	Asp	Asn
Met	Arg 370	Lys	Ser	Leu	Gln	Gln 375	Val	Lys	Pro	Gly	Tyr 380	Ile	Arg	Val	Asp
Ala 385	Asp	Glu	Val	Суѕ	Tyr 390	Pro	Leu	His	Val	Ile 395	Leu	Arg	Tyr	Glu	Ile 400
Glu	Arg	Asp	Leu	Met 405	Glu	Gly	Lys	Met	Glu 410	Val	Glu	Asp	Val	Pro 415	Arg
Ala	Trp	Asn.	Ala 420	Lys	Met	Gln	Glu	Tyr 425	Lėu	Gly	Leu	Ser	Thr 430	Glu	Gly
Arg	Asp	Asp 435	Val	Gly	Cys	Leu	Gln 440	Asp	Val	His	Trp	Ser 445	Met	Val	Arg
Ser	Ala 450	Thr .	Leu	Arg		Thr 455	Arg	Ser	Ala	Pro	Cys 460	Met	Arg	Arg	Arg

Ser Trp Arg Ala Ser Glu Arg Ser Trp Glu Thr Thr Arg Trp Met Ser

120

125

Arg Lys Thr Gly Lys Ala Gly Thr 130 · 135

- (2) INFORMATION FOR SEQ ID NO:50:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 510 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania chagasi
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:

Tyr Leu Leu Pro Leu Leu Gly Arg Arg Thr Thr Thr Thr Phe Lys Thr

1 10 15

Thr Pro Arg Leu Leu Val Pro His Leu Leu Leu Ser Thr Phe Asn Pro 20 25 30

Cys Leu Ala Asn Met Glu Ala Tyr Lys Lys Leu Glu Thr Ile Phe Thr 35 40 45

Lys Val Tyr Arg Leu Asp His Phe Leu Gly Leu Gly Asn Trp Asp Met 50 55 60

Asn Thr Asn Met Pro Pro Lys Gly Glu Glu Ser Arg Gly Glu Ala Met 65 70 75 80

Ala Met Leu Ser Glu Leu Arg Phe Gly Phe Ile Thr Ala Pro Glu Val 85 90 95

Lys Ser Leu Ile Glu Ser Ala Thr Lys Gly Ser Glu Glu Leu Asn Ala 100 105 110

Val Gln Arg Ala Asn Leu Arg Glu Met Arg Arg Ala Trp Lys Ser Ala 115 120 125

Thr Ala Leu Pro Ala Glu Phe Val Gly Arg Lys Met Arg Leu Thr Thr 130 135 140

His Ala His Ser Val Trp Arg Asp Ser Arg Lys Ala Asn Asp Phe Ala 145 150 155 160

Lys Phe Leu Pro Val Leu Arg Asp Leu Val Ala Leu Ala Arg Glu Glu 165 170 175

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GGTGTCTTAC	TACGAGATCT	ACAACGAGAA	GGTGTTCGAT	CTCATCCGGC	CGCAGCGCAA	660
CACGGACCTG	AGGATACGTA	ACTCGCCCAA	CTCCGGTCCA	TTTATCGAAG	GCCTGACGTG	720
GAAGATGGTG	TCCAAGGAGG	AAGACGTCGC	CCGCGTGATT	CGCAAGGGCA	TGCAGGAGCG	780
CCACACGGCT	GCGACCAAGT	TCAACGACCG	CAGCAGCCGC	AGCCACGCCA	TCCTCACCTT	840
CAACATTGTG	CAGCTGTCGA	TGGACGACTC	CGACAACGCG	TTCCAGATGC	GCAGCAAGCT	900
GAACCTGGTG	GACCTTGCTG	GGTCGGAGCG	CACTGGTGCG	GCCGGAGCCG	AGGGCAATGA	960
GTTCCACGAC	GGTGTGAAGA	TCAACCACTC	GCTGACGGTG	CTGGGGCGCG	TGATCGACCG	1020
TCTGGCGGAC	CTCTCGCAGA	ACAAGGGAGG	GGG .			1053

## (2) INFORMATION FOR SEQ ID NO:49:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 136 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania chagasi
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:
- Gly Arg Leu Ser Glu Glu Glu Ile Glu Arg Met Val Arg Glu Ala Ala 1 5 10 15
- Glu Phe Glu Asp Glu Asp Arg Lys Val Arg Glu Arg Val Glu Ala Lys 20 25 30
- Asn Ser Leu Glu Ser Ile Ala Tyr Ser Leu Arg Asn Gln Ile Asn Asp 35 40 45
- Lys Asp Lys Leu Gly Asp Lys Leu Ala Ala Asp Asp Lys Lys Ala Ile 50 55 60
- Glu Glu Ala Val Lys Asp Ala Leu Asp Phe Val His Glu Asn Pro Asn 65 70 75 80
- Ala Asp Arg Glu Glu Phe Glu Ala Ala Arg Thr Lys Leu Gln Ser Val 85 90 95
- Thr Asn Pro Ile Ile Gln Lys Val Tyr Gln Gly Ala Ala Gly Ser Gly
  100 105 110
- Ala Glu Glu Ala Asp Ala Met Asp Asp Leu Leu Val Gly Arg Val Lys

TCATCATACA GCATCCAATA GCCGCCAGAG TGAATGCGAT GCGCACACCA AGTCGAAAGT 180 GGTCGACCAG TAGGGGAATG TGACCCTGGC TGGCGTGCAA CATGATCGCC ACGCCAGCGG 240 TGGGCCACAC CACAACAGAG GCGACGAAAG AGAACATGAA CTTGCTCACG AAGCTNACAA 300 TAAGGGCGTC GCTNGTGATG CTAAGAACCA CGCCNAGGTA GACGGCGAAG ANCAAACTAA 360 ACACAGCGT GACGATCCCG AAAAGAAGGA TCTCTGCGGA ATTTTCGTGA GATAGANAAT 420 GCCCGTACTG GAAAAANAAG CCGGCAGGCG CGCGATAACG CTGCAACTTG CCGCTCCTCG 480 CGGGCGCGTT TTCGCTCCTT CTCCGACTTG ATGGCGCNGT CNGNCTTGAC AAAACGGTTA 540 AGCTCCTCAT GCCCCAGCCG ATTCCCAGCT CACGGTCCAC TTCCGGCCAT GCCCACGGAC 600

#### (2) INFORMATION FOR SEQ ID NO:48:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1053 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania chagasi

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

GGGAAAAAG TGGAGCTCCA CCGCGGTGGC GGCCGCTCTA GAACTAGTGG ATCCCCCGGG 60 CTGCAGGAAT TCCGCGGAAT TCCGCGGAAT TCCGTCCGAC GCGGCACCCG 120 CACAGGGGTC GACAGTGACG CAACCTCCTC CACCACTGCG GCCTACGACG GCGCCGGCTC 180 CGCGCCAGTG ATGGTTGACG CCAATGTGAG CCACCCTCCG TACGCGGGGC ATGACCAAGT 240 GTACATGCAC GTCGGCAAGC CCATCGTGGG CAACACCCTC GACGGATACA ACGGGTGCGT 300 GTTCGCCTAC GGGCANACGG GCAGCGGCAA AACCTTCACG ATGCTCGGNT ACGCGCCGAG 360 CACGANCGAC ATCCGCGCTC GCAAAGGGTC CGTCCCCTGC GGGGCCAGCA GCATGGAGAA 420 480 CAGCACTCCT CTTGACAGCG CTGTGGAGCC GTTTGAGAGC GATGACGGCG ACGACGTGGT GGACAAGACG GGGCTGGATC CGAACGAGCT GCAAGGCATC ATCCCGCGCG CGTGCACGGA 540 CCTGTTCGAT GGTCTCCGTG CGAAGCGCGC CAAGGACTCC GACTTCACGT ACCGCGTGGA 600 (i) SEQUENCE CHARACTERISTICS:

(ii) MOLECULE TYPE: DNA (genomic)

(A) ORGANISM: Leishmania chagasi

(vi) ORIGINAL SOURCE:

(A) LENGTH: 560 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear

60

120

180

240

300

360

420

480

540

560

60

120

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ı	7.7

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(xi)	SEQUENCE DES	SCRIPTION: S	EQ ID NO:46	:	
CGGAAGGAG	G ATGGCCATAC	ACAGAAAAT	GACGGCGATG	GCCCTAAGGA	GGACGGCCGT
ACACAGAAA	A ACGACGACGG	G TGGCCCTAAG	GAGGACGCC	ATACACAGAA	AAATGACGGC
GATGGCCCT	A AGGAGGACGG	CCGTACACAG	AAAAATAACG	GCGATGGCCC	TNAGGAGGAC
GGCCATACAG	C AGAAAAATGA	CGGCGATGCC	CCTNAGGAGG	ACGGCCGTAC	ACANAAAAAT
GACGGCNATO	GCCCTNAGGA	GGACGGCCGT	ACACAGAAAA	ATGACNGCCA	TGGCCCTTAG
GANGACGCC	TACACAGAAA	AATGACGCNA	TGGCCCTNAG	GGAGGACGGC	CATACCCANA
AAAATTGACG	GCNATNGCCC	TTAGGANGAC	GGCCGTNCCC	ANAAANANTG	ACNGCGGTNG
CCCTTAAGGA	AGATGAAAAT	CTGCCACCAA	AACNATTGGG	AATGCNCAGG	AAAANAACNA
ANATNGACCO	CACGTGGGGG	ATGGANCTTA	CNGCNATTAA	NATTGTTACC	ATTATCNACC
NAAGGACNNG	TTGCCGNCAA	•			
(2) INFORM	MATION FOR S	EQ ID NO:47	:		
(i) s	EOHENCE CUA	RACTERISTICS	•	*	
(1) 5	(A) LENGTH.	600 base pa	i:		·
	(B) TYPE: pi	cleic acid	urs		
		ONESS: doubl			
	(D) TOPOLOGY				
(ii) M	OLECULE TYPE	E: DNA (geno	omic)		
(vi) o	RIGINAL SOUP	PCR •			
		d: Leishmani	a chagasi		٠.

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

CGTCCGAGAA ACCCGTACAT GTATGCTGCT GGTAGAAGGC GCAGAGCTGG TCCCTCTGAT

GCACAAGCAT GAGGTCGTAC ATTGCCTGGT TCGTCATTTT CCAGAGCACA ACGAGCAGCG

ATGCCCCCCA	AGGGCGAGGA	ATCACGCGGT	GAGGCGATGG	CGATGCTCTC	GGAGCTCCGC	300
TTTGGCTTCA	TCACGGCACC	GGAGGTGAAA	AGCCTGATTG	AGAGTGCCAC	CAAGGGCAGC	360
GAGGAGCTGA	ATGCGGTGCA	GCGCGCTAAC	TTGCGGGAGA	TGAGGCGTGC	GTGGAAGAGC	420
GCCACCGCCT	TGCCGGCTGA	GTTTGTGGGC	CGCAAGATGC	GCCTCACGAC	ACACGCGCAC	480
AGCGTGTGGC	GCGACAGCCG	CAAAGCAAAT	GACTTCGCCA	AGTTCCTACC	GGTGCTCAGG	540
GACCTGGTGG	CGCTCGCCCG	TGAGGAGGGC	TCATACCTCG	CCGCCGGCAC	CTCCCTCTCC	600
CCGTATGAGG	CGCTCATGAA	CGAGTACGAG	CCAGGAATCA	CGACACAAAA	GCTGGATGAG	660
GTGTACGCAA	ATGTAAAGTC	GTGGCTGCCG	CAGCTGCTAA	AGGACATTGT	GCAGAAGCAG	720
TCCGGCGAGT	CGGTGATTGC	GTTCTCGCAT	AAGTTCCCGC	AGGACAAGCA	GGAAGCACTG	780
TGCAAGGAAT	TCATGAAGAT	CTGGCACTTC	GACACCGATG	CCGGTCGCCT	CGACGTCAGC	840
CCCCACCCTT	TCACGGGAAT	GACGAAGGAG	GACTGCCGAC	TCACAACAAA	CTACATCGAA	900
GACACGTTTG	TTCAGAGCTT	GTATGGCGTC	ATCCACGAGA	GTGGGCATGG	CAAGTACGAG	960
CAGAACTGTG	GCCCACGCGA	GCACATCACG	CAGCCGGTGT	GCAACGCCCG	CTCTCTTGGC	1020
CTGCATGAGA	GCCAGAGCCT	CTTTGCGGAG	TTTCAGATCG	GCCACGCGAC	GCCCTTCATC	1080
	CAACTCGCCT	TCCTGAGTTC	TTCGAGGCGC	AGCCAGCGTT	CTCGCAGGAC	1140
AACATGCGCA	AGTCGCTGCA	GCAGGTGAAG	CCGGGCTACA	TTCGCGTCGA	TGCCGATGAG	1200
GTGTGCTACC	CTCTGCACGT	GATCCTGCGC	TACGAGATCG	AGCGCGACTT	GATGGAGGC	1260
AAAATGGAGG	TGGAAGACGT	GCCGCGCGCG	TGGAACGCAA	AGATGCAGGA	GTACTTGGGT	1320
CTCTCAACGG	AGGGCCGTGA	CGACGTTGGG	TGCCTGCAGG	ACGTGCATTG	GTCCATGGTG	1380
CGCTCGGCTA	CTCTCCGACG	TACTCGCTCG	GCGCCATGTA	TGCGGCGCAG	ATCATGGCGA	1440
GCATCCGAAA	GGAGCTGGGA	GACGACAAGG	TGGATGAGTG	CCTGCGCACC	GGTGAGCTCG	1500
GCCCCTCCT	' GGAAAAGCAG	CAGGAGAAGA	TCTGGGATCA	TGGGTGCCTG	TACGAGACGG	1560
ACGACCTCAT	GACGCGTGCG	ACGGGCGAGA	CGCTGAACCC	CGAGTACCTG	CGCCGCCACC	1620
TGGAGGCGCG	CTACATAAAC	GCCTGAGTCG	CGAGCGGTTG	ACACACGCGC	TCGCTAGCAC	1680
ATGACGCGTC	CTTTATTATTC	TTTGTTGTGC	ATTCGGAATT	CCGCGGAATT	CGATATCAAG	1740
CTTATCGA						1748

(ii)	MOLECULE	TYPE:	DNA	(genomic)	)
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### (vi) ORIGINAL SOURCE:

(A) ORGANISM: Leishmania chagasi

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

CGGCCGCCTC	AGCGAGGAGG	AGATCGAGCG	CATGGTGCGC	GAGGCTGCCG	AGTTCGAGGA	6
TGAGGACCGC	AAGGTGCGCG	AACGTGTCGA	AGCGAAGAAC	TCGCTAGAGA	GCATCGCGTA	120
CTCGCTTCGC	AACCAGATCA	ACGACAAGGA	CAAGCTTGGT	GACAAGCTCG	CCGCGGACGA	180
CAAGAAGGCG	ATCGAGGAGG	CTGTGAAGGA	TGCCCTCGAC	TTTGTCCACG	AGAACCCCAA	240
TGCAGACCGT	GAGGAGTTCG	AGGCTGCTCG	CACGAAGCTG	CAGAGTGTGA	CGAACCCCAT	300
CATTCAAAAG	GTGTACCAGG	GCGCCGCCGG	CTCTGGTGCA	GAAGAGGCGG	ACGCGATGGA	360
TGACTTGTTA	GTCGGCCGCG	TGAAAAGAAA	AACAGGGAAA	GCGGGAACAT	NCCACAANAA	420
CCNAAGAAGA	AAGGGGGTNG	CGACACCGCT	CGAACACCGA	CGGCNCACAT	NCNTCATGGG	480
CATGCTCAGC	TTTCCTCTCC	CCAACAAACC	AGAAGGTTTT	CTCCAAACNC	CGTCTCNGCN	540
CCCAAAATAC	GGAAANGTTA	ANCGAAAAAN	CCCCTTCCÁC	CAATTGNNGT	ТСТТТСТТТ	600

# (2) INFORMATION FOR SEQ ID NO:45:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1748 base pairs
  - (B) TYPE: nucleic acid .
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania chagasi

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

					,	
CTAGTGGATC	CCCCGGGCTG	CAGGAATTCA	CGGAATACGT	ACCTCCTCCC	CCTTCTTGGT	60
AGAAGAACAA	CAACAACGTT	CAAGACGACG	CCGCGCCTTC	TTGTACCGCA	TTTGCTTCTG	120
AGCACGTTCA	ATCCGTGCCT	TGCAAACATG	GAGGCGTACA	AGAAGCTGGA	AACGATCTTT	180
ACGAAGGTCT	ACCGCCTGGA	CCACTTCCTC	GGTCTGGGCA	ACTGGGACAT	GAACACAAAC	240

Ala Arg Xaa Ile Phe Val Lys Thr Leu Thr Gly Xaa Thr Ile Ala Leu 1 5 10 15

Glu Val Glu Pro Ser Asp Thr Ile Glu Asn Val Lys Ala Lys Ile Gln 20 25 30

Asp Lys Glu Gly Ile Pro Pro Asp Gln Gln Arg Leu Ile Phe Ala Gly
35 40 45

Lys Gln Leu Glu Xaa Gly Arg Thr Leu Ser Asp Tyr Asn Ile Gln Lys 50 55 60

Glu Ser Thr Leu His Leu Val Leu Arg Leu Arg Gly Gly Met Xaa Ile 65 70 75 80

Phe Val Lys Thr Leu Thr Gly Xaa Thr Ile Ala Leu Glu Val Glu Pro 85 90 95

Asn Asp

#### (2) INFORMATION FOR SEQ ID NO:43:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 39 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

Leu Gln Gln Arg Leu Asp Thr Ala Thr Gln Gln Arg Ala Glu Leu Glu 1 5 10 15

Ala Arg Val Ala Arg Leu Ala Ala Asp Arg Asp Glu Ala Arg Gln Gln 20 25 30

Leu Ala Ala Asn Ala Glu Glu 35

#### (2) INFORMATION FOR SEQ ID NO:44:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 600 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear

### (A) ORGANISM: Leishmania major

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:
- Leu Gln Gln Arg Leu Asp Thr Ala Thr Gln Gln Arg Ala Glu Leu Glu

  1 10 15
- Ala Arg Val Ala Arg Leu Ala Ala Asp Arg Asp Glu Ala Arg Gln Gln 20 25 30
- Leu Ala Ala Asn Ala Glu Glu Leu Gln Gln Arg Leu Asp Thr Ala Thr 35 40 45
- Gln Gln Arg Ala Glu Leu Glu Ala Arg Val Ala Arg Leu Ala Ala Asp 50 55 60
- Gly Asp Glu Ala Arg Gln Gln Leu Ala Ala Asn Ala Glu Glu Leu Gln 65 70 75 80
- Gln Arg Leu Asp Thr Ala Thr Gln Gln Arg Ala Glu Leu Glu Ala Gln 85 90 95
- Val Ala Arg Leu Ala Ala Asn Ala Glu Glu Leu Gln Gln Arg Leu Asp 100 105 110
- Thr Ala Thr Gln Gln Arg Ala Glu Leu Glu Ala Arg Val Ala Arg Leu
  115 120 125
- Ala Ala Asp Arg Asp Glu Ala Arg Gln Gln Leu Ala Ala Asn Ala Glu 130 135 140
- Glu Leu Gln Gln Arg Leu Asp Thr Ala Thr Gln Gln Arg Ala Glu Leu 145 150 155 160
- Glu Ala Gln Val Ala Arg Leu Ala Ala 165

## (2) INFORMATION FOR SEQ ID NO:42:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 98 amino acids
  - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

Gln Gly Gln Xaa Phe Val Gly Asn Ser Leu Val Val Arg Leu Ile Asp 20 25 30

Xaa Leu Xaa Gln Xaa Pro Ala Gly Tyr Pro Val Tyr Xaa Asn Arg Gly 35 40 45

Ala Xaa Gly Ile Xaa Xaa Leu Leu Ser Thr Ala Ala Gly Val Xaa Arg 50 55 60

Ala 65

- (2) INFORMATION FOR SEQ ID NO:40:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 78 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania major
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

Asp Asp His Ala Pro Lys Glu Asp Gly His Ala Pro Lys Asn Asp Asp 1 5 10 15

His Ala Pro Lys Glu Asp Gly His Ala Pro Lys Asn Asp Asp His Ala 20 25 30

Pro Lys Glu Asp Gly His Ala Pro Lys Asn Asp Gly Asp Val Gln Xaa 35 40 45

Lys Ser Glu Asp Gly Asp Asn Val Gly Glu Gly Gly Lys Gly Asn Glu 50 55 60

Asp Gly Asn Asp Asp Gln Pro Lys Glu His Ala Ala Gly Asn 65 70 75

- (2) INFORMATION FOR SEQ ID NO:41:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 169 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:

Asn Asn Ala Arg Gln Ala Glu Asp Pro Ala Arg Leu Ile 65 70 75

- (2) INFORMATION FOR SEQ ID NO:38:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 68 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania major
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:
  - His Glu Pro Gln Trp Ser Ser Met Lys Ile Leu Gln Tyr Leu Thr Leu 1 5 10 15
  - Asp Gly Thr Gln Val Ser Gly Thr Leu Pro Pro Gln Trp Ser Ala Met 20 25 30
  - Ala Ser Val Arg Ile Leu Asn Leu Xaa Gly Thr Glu Val Ser Gly Thr 35 40 45
  - Leu Pro Pro Glu Trp Ile Ser Met Xaa Arg Leu Gln Thr Leu Asn Leu
    50 55 60

Arg Arg Thr Lys

- (2) INFORMATION FOR SEQ ID NO:39:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 65 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania major
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:
  - Ala Arg Glu Ala Gln Leu Ala His Arg Ile Cys Asp Tyr Leu Pro Glu

    5 10 15

20

.

30

Leu Val Val Ala His Glu Gly Asp Gly Gln Ser Ser Ala Ala His Ala 35 40 45

Ala Ser Ala Ala Asp Ala Val Asn Val Leu Leu Thr Arg Glu Arg Gln 50 55 60

Ile Val Ala Glu Asp Glu Arg Asp Ala Leu His Ile Asn Ala Thr Arg
65 70 75 80

Pro Gln Val Arg Cys Asn Xaa His Ala Ala Val Ser Ile Thr Glu Cys 85 90 95

Arg Ile Ile Ser Ser Arg Ser Ala Trp Gly Met Ser Pro Cys Met Ala 100 105 110

Asp Thr Ala Lys Phe Ser Arg Val Ile Phe Leu Ala Ser Cys Ser Thr 115 120 125

Phe Leu Arg Val Leu Xaa Lys Thr Thr Ala Trp Ala Ile Val Lys Leu 130 135 140

Ser Tyr 145

### (2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 77 amino acids
  - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

Ala Gln Gly Phe Arg Val Ile Phe Thr His Gly Gly Ala Gln Met Gly
1 10 15

Glu Val Asn Thr Arg Thr Glu Leu Leu Asp His Ile Asn Gln Ile Val 20 25 30

Val Ser Thr His Ala Val Xaa Thr Gly Ala His Gly Lys Thr Val Cys
35 40 45

His Ala Val Tyr Gly Ile Asn His Pro Leu His Ile Phe Asn Gly Gly 50 55 60

## (A) ORGANISM: Leishmania major

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

GGCACGANAG	ATCTTCGTGA	AGACGCTGAC	CGGCAANACG	ATCGCGCTGG	AGGTGGAGCC	60
GAGCGACACG	ATCGAGAACG	TGAAGGCCAA	GATCCAGGAC	AAGGAGGGCA	TCCCGCCGGA	120
CCAGCAGCGC	CTGATCTTCG	CCGGCAAGCA	GCTGGAGGAN	GGCCGCACGC	TCTCGGACTA	180
CAACATCCAG	AAGGAGTCCA	CGCTGCACCT	GGTGCTGCGC	CTGCGCGGCG	GCATGCANAT	240
CTTCGTGAAA	ACGCTNACCG	GCAANACAAT	CGCGCTGGAA	GTGGAGCCGA	ACGACCNATC	300
GAAAACGTGA	AGGCCNANAT	CCANGACAAG	GAAGGCNTCC	CGCCGGANCA	GCACGCCTGA	360
TCTTCCNCCG	GCAACCACTT	GANGAAGGGC	NCACGCTCTC	NGACTACNAC	ATCCANAAAG	420
GATTCCNCCC	TGCACCTTGT	TGCTTGCNCC	TTGCTCGGGG	GGCATGCCNA	ATCTTCCTTN	480
AAAACCTCAA	CCGGCAANAA	CAATCCCCCN	CNGAAGTTGG	AACCCAACCA	NCCCATTCNA	540
AAACTTTAAA	GGCCNNNATT	CCNGAACAAN	GAAGGGCTTC	CCCCCGGAC	CNNCAANCNC	600
CCTGATTNTT	CCCCCGGNNN	NCANTTTGGA	ANGAAGGGCC	CCNCCCTCCN	CCGAATTNCN	660
ACNTCCCNAA	ANGGATTCCC	CCCCTNCCCT	TGNTTTTTGC	GCCNNNNNNC	GGCNNCNTNC	720
CNAAATTCCG	NCCNAAGGNC	CCCANTANAN	CNACTTTCCC	NTTCCCCCCC	NNNNTTTTGC	780
NTAAANTTTT	TNCCCCCNNA	AANNTCCCNT	TTNCNANTTN	ΔN	•	022

## (2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 146 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

Gly Thr Ser Pro Cys Leu His Leu Leu Ala Asp Ile Arg Gly Glu Phe 1 5 10 15

Phe Asn Leu Arg Arg Val Glu Leu Leu Asn Val Ala Gln Gln Ala Gln

TGCACGGGGT	TGCTGTGGCT	GCACCTCCTG	ACCACTGCCA	GCTTTCTTGG	CTTGCCTCCC	420
CTCTGCGCCT	CCGCTCGTGC	CGCTCGTGCC	GAATTCGATA	TCAAGCTTAT	CGATACCGTC	480
NACCTCGAAG	GGGGCCCGG	TTACCCATTC	GCCCTATANT	GAGTCNTATT	ACAATTCCTG	540
GCGTCGTTTT	ACACGTCGTG	ACTGGGAAAA	ACCCTGGCGT	TCCCCACTTA	TCGCCTTGCA	600

### (2) INFORMATION FOR SEQ ID NO:34:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 516 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

AGCTGCAGCA	GCGCCTAGAC	ACCGCCACGC	AGCAGCGCGC	CGAGCTGGAG	GCACGGGTGG	60
CACGGCTGGC	CGCGGACCGC	GACGAGGCGC	GCCAGCAGCT	GGCCGCGAAC	GCCGAGGAGC	120
TGCAGCAGCG	CCTAGACACC	GCCACGCAGC	AGCGCGCCGA	GCTGGAGGCA	CGGGTGGCAC	180
GGCTGGCCGC	GGACGGCGAC	GAGGCCCGCC	AGCAGCTGGC	CGCGAACGCC	GAGGAGCTGC	240
AGCAGCGCCT	AGACACCGCC	ACGCAGCAGC	GCGCCGAGCT	GGAGGCACAG	GTGGCACGGC	300
TGGCCGCGAA	CGCCGAGGAG	CTGCAGCAGC	GCCTAGACAC	CGCCACGCAG	CAGCGCGCCG	360
AGCTGGAGGC	ACGGGTGGCA	CGGCTGGCCG	CGGACCGCGA	CGAGGCGCGC	CAGCAGCTGG	420
CCGCGAACGC	CGAGGAGCTG	CAGCAGCGCC	TAGACACCGC	CACGCAGCAG	CGCGCCGAGC	480
TGGARGCACA	GGTGGCACGG	CTGGCCGCGA	AMGCCG			516

### (2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 822 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:

(xi)	SEQUENCE	DESCRIPTION:	SEO	ID	NO:32:
------	----------	--------------	-----	----	--------

NGCACGAGAA	GCGCAACTGG	CGCATCGCAT	CTGTGACTAT	CTGCCTGAAC	AGGGGCAATN	60
GTTTGTTGGT	AÁCAGCCTGG	TGGTACGTCT	GATTGATNCG	CTTNCGCAAN	TTCCGGCAGG	120
TTACCCGGTG	TACANCAACC	GTGGGGCCAN	CGGTATCNAC	NGGCTGCTTT	CGACCGCCGC	180
CGGNGTTCAN	CGGGCAANCG	GCAAACCGAC	GCTGGCGATT	GTGGGCGATC	TCTCCGCACT	240
TTACGATCTC	AACGCNCTGG	CGTTATTGCG	TCAGGTTTCT	GCGCCGCTGG	TATTAATTGT	300
GGTGAACAAC	AACGGCNGGG	CAAAATTTTC	TCGCTGTTGC	CAACGCCCCC	AAAGCNAGCG	360
TGAAGCGTTT	CTATCTGATG	CCGCAAAACG	TCCATTTTGA	AACACGCCGC	CNCCCATGTT	420
TCGANCTGAA	AATATCATCG	TCCGCAAAAC	TGGCANGAAA	CTTNGÄAAAC	CGCATTTTGC	480
CGACNCCCTG	GCNCACGCCC	AACCCACCCA	CCGGTTGATT	GAAAATGGTG	GGTTAACGAA	540
NCCNNATGGG	TGCCCCAAAN	CNCNNCCANC	CAAATTTCTG	GGCCCAGGTT	AAANCCCTTT	600

# (2) INFORMATION FOR SEQ ID NO:33:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 600 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

ACGATGACCA TGCCCCGAAG GAGGATGGCC ATGC	GCCGAA GAACGATGAC CATGCCCCGA 60
AGGAGGATGG CCATGCGCCG AAGAACGATG ACCA	TGCCCC GAAGGAGGAT GGCCATGCGC 120
CGAAGAACGA CGGGGATGTG CAGAANAAGA GCGA	AGATGG AGACAACGTG GGAGAGGGAG 180
GCAAGGGCAA TGAGGATGGT AACGATGATC AGCC	GAAGGA GCACGCTGCC GGCAACTAGT 240
GGGCTGCGTC CGGGCTTGTG TGCGANCCGT GCTC	TGCACC CCGCCGCTCG TGCATCCTCG 300
CATGTGGACT GCGTGTGTCT CTCCCGCTTT GTCT	CTCTCC CCCACACAGT GGCTGATGCC 360

TTCAAATTNA	TTCCCNACCT	NCCNTNNCCA	AANNTANCNA	ATAATCANNC	CCCTNTCANN	540
ANNTCCCANC	TTACCCTCCN	NTNGNNGGGN	NNNCCNATTN	CCCCAANCCC	NCNCTAAATA	600

### (2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 600 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

GGCACGAGCC	TCAGTGGAGC	TCAATGAAGA	TATTGCAGTA	TCTTACTCTG	GATGGCACTC	60
AGGTCTCCGG	CACGCTGCCG	CCCCAGTGGA	GCGCGATGGC	ATCGGTGCGA	ATTCTTAACC	120
TGNAGGGTAC	TGAGGTCTCT	GGTACGCTGC	CGCCTGAGTG	GATATCNATG	ANCAGGCTGC	180
AAACTCTGAA	TCTGCGGCGC	ACGAAANTAT	CCGGCACTCT	GCCGCCCGAA	TGGANTTCTA	240
TGAACAGCCT	GGAGTACTTT	CACCTTTATC	TTACTCAGGT	CTCCGGCACG	CTGCCGCCCG	300
AGTGGAGTGG	GATGTCNAAG	GCCGCATACT	TCTGGCTGGA	ATACTGCGAC	CTGTCCGGCA	360
NTCTGCCGCC	CNAGTGGTCG	TCNATGCCAA	AGCTGCGCGG	TATCTCACTG	ANCGGCAACA	420
AATTCTTGCG	NGTGTNTNCC	NGACTCNTGG	GATTCAGAAA	GGTGGTCCTT	GTTGTTGGGC	480
ATCNAAGGAN	CAAACCCCAA	NGGGCCCNCN	AATTGCTTGG	GCNTGCTTAA	GGANTTGCAC	540
NAACCAACNC	CNCCAAAAAC	CCCCCCACC	NCNAAANNAC	NANCCCCCAC	TTAANNCCCN	600

### (2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 600 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

(xi)	SEQUENCE	DESCRIPTION:	SEO	ID	NO:29:

GGCACGAGCC	CTTGCCTACA	TTTGCTCGCC	GATATTCGCG	GGGAGTTCTT	CAATTTGCGT	6
CGCGTAGAAC	TGCTCAATGT	CGCGCAACAA	GCGCAGCTCG	TCGTGGCGCA	CGAAGGTGAT	12
GGCCAGTCCA	GTGCGGCCCA	TGCGGCCAGT	GCGGCCGATG	CGGTGAATGT	ACTGCTCACG	180
CGCGAGCGGC	AAATCGTAGC	TGAGGACGAG	CGAGACGCGC	TCCACATCAA	TGCCACGCGC	240
CCACAGGTCC	CTTGTAATGA	NCACGCGGCT	GTGTCCATTA	CGGAATGCCG	CATAATCTCG	300
TCGCGCTCCG	CCTGGGGCAT	GTCGCCGTGC	ATGGCGGACA	CAGCGAAATT	CTCGCGCGTC	360
ATCTTCTTGG	CAAGCTGCTC	CACCTTTTTG	CGGGTGTTGC	ANAAAACCAC	NGCGTGGGCG	420
ATCGTTAAGC	TGTCGTACAA	ACTCCATCAA	GAAATCGAAT	TTGTTTTTCT	CTTCGTCNAC	480
NGANACAAAN	TACTGTTTAA	CGCTNTCCAC	GGTGATCTCA			520

# (2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 600 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

GGCACAAGGT	TTTCGGGTTA	TCTTCACGCA	TGGTGGAGCG	CAGATGGGTG	AAGTAAATAC	60
GCGGACCGAA	CTGCTTGATC	ATATCAACCA	GATCGTTGTC	AGCACGCACG	CCGTANGAAC	120
CGGTGCACAT	GGTAAAACCG	TNTGCCATGC	TGTTTACGGT	ATCAACCATC	CACTGCATAT	180
CTTCAATGGT	GGAAACAATG	CGCGGCAGGC	CGAGGATCCG	GCGCGGCTCA	TCATNNAGNT	240
NATNAACCAN	TCGCACGTCT	ANTTCTGCAC	ТАААСТАСАА	NTATCGGTNA	CATATNATAA	300
GGCCNATTTT	CGGTCCAGGA	NTATGTNCTN	TCAAAATGCC	NCGTTANNCA	CTCTTAAATG	360
TCTCANGNGN	AAANTNGTTC	TAAAGGGTGT	CCAAAANNTN	NTTACCNTTC.	CCCNCTTACT	420
TCAANANCTC	CTCNAATTCC	CNGGCCCTTN	GACNANNATT	TNCTATTAAA	ANATANAANN	480

Gly Glu Val Cys Pro Ala Asn Trp Lys Lys Gly Ala Pro Thr Met Lys 180 185 190

Pro Glu Pro Lys Ala Ser Val Glu Gly Tyr Phe Ser Lys Gln 195 200 205

- (2) INFORMATION FOR SEQ ID NO:27:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 51 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: other nucleic acid
  - (A) DESCRIPTION: /desc = "50 PCR primer (MAPS-1-50His) to simultaneously amplify MAPS-1 cDNA for both L. major and L. tropica while adding 6 His residues to amino terminal end of encoded protein."
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

CAATTACATA TGCATCACCA TCACCATCAC ATGTCCTGCG GTAACGCCAA G

51

- (2) INFORMATION FOR SEQ ID NO:28:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 31 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: other nucleic acid
  - (A) DESCRIPTION: /desc = "3' PCR primer (MAPS-1-3ÕR1) to simultaneously amplify MAPS-1 cDNA for both L. major and L. tropica.Ó
    - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

CATGGAATTC TTACTGCTTG CTGAAGTATC C

- (2) INFORMATION FOR SEQ ID NO:29:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 520 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania major

AAC	CA	GG	CGA	G GT	G TG	C CC	C GC	G AA	TG	G AAG	G AA	G GG	C GC	c cc	C ACG	
Lys 175	His	s Gly	y Gla	u Vai	1 Cy:	s Pro	o Ala	a Ası	ı Trj	185	Ly	s Gl	y Al	a Pro	o Thr 190	
ATC Met	Lys	CCC Pro	G GAA	A CCC 1 Pro 195	) Lys	G GCC	TC:	T GTO	GAC Glu 200	ı Gly	TAC Tyi	C TTO	C AGG	C AA( r Lys 20!	G CAG S Gln	
TAA	GAA1	TCC	ATG	•,										•		
(2)	INF	ORMA	MOITA	FOF	SEC	) ID	NO:2	26 :								
			( <i>P</i> (E	JENCE () LE () TY () TC	NGTH PE: POLC	: 20 amin GY:	6 am 10 ac line	ino id ar		ls						
				CULE												
	(	X1)	SEQU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	26:	-				•
Met 1	His	His	His	His 5	His	His	Met	Ser	Cys 10		Asn	Ala	Lys	Ile 15	Asn	
Ser	Pro	Ala	Pro 20	Pro	Phe	Glu	Glu	Met 25	Ala	Leu	Met	Pro	Asn 30		Ser	
Phe	Lys	Lys 35	Ile	Ser	Leu	Ser	Ala 40	Tyr	Lys	Gly	Lys	Trp	Val	Val	Leu	
Phe	Phe 50	Tyr	Pro	Leu	Asp	Phe 55	Thr	Phe	Val	Cys	Pro 60	Thr	Glu	Ile	Ile	
Ala 65	Phe	Ser	Asp	Asn	Val 70	Ser	Arg	Phe	Asn	Glu 75	Leu	Asn	Cys	Glu	Val 80	
Leu	Ala	Cys	Ser	Met 85	Asp	Ser	Glu	Tyr	Ala 90	His	Leu	Gln	Trp	Thr 95	Leu	
Ġln	Asp	Arg	Lys 100	Lys	Gly	Gly	Leu	Gly 105	Ala	Met	Ala	Ile	Pro	Met	Leu	-
Ala	Asp	Lys 115	Thr	Lys	Ser	Ile	Ala 120	Arg	Ser	Tyr	Gly <sub>.</sub>	Val 125	Leu	Glu	Glu	
Ser	Gln 130	Gly	Val	Ala	Tyr	Arg 135	Gly	Leu	Phe	Ile	Ile 140	Asp	Pro	Arg	Gly	
let l45	Val	Arg	Gln	Ile	Thr 150	Val	Asn	Asp	Met	Pro 155	Val	Gly	Arg	Asn	Val 160	
3lu	Glu	Ala	Leu	Arg 165	Leu	Leu	Glu	Ala	Leu 170	Gln	Phe	Val	Gļu	Lys 175	His	

(ii) MOLECULE	TYPE:	CDNA
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### (vi) ORIGINAL SOURCE:

(A) ORGANISM: Leishmania tropica

### (ix) FEATURE:

(A) NAME/KEY: CDS(B) LOCATION: 7..624

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

(xi) SEQUENCE DESCR	IPTION: SEQ ID NO:25	* ***
TTACAT ATG CAT CAC CAC Met His His His I	CAC CAC CAC ATG TCC His His His His Met Ser	
ATC AAC TCT CCC GCG CCG Ile Asn Ser Pro Ala Pro 15 20		
GGC AGC TTC AAG AAG ATC Gly Ser Phe Lys Lys Ile 35		
GTG CTC TTC TTC TAC CCG Val Leu Phe Phe Tyr Pro 50		
ATC ATC GCG TTC TCC GAC Ile Ile Ala Phe Ser Asp 65		
GAG GTC CTC GCG TGC TCG Glu Val Leu Ala Cys Ser 80		
ACG CTG CAG GAC CGC AAG Thr Leu Gln Asp Arg Lys 95 100	Lys Gly Gly Leu Gly	
ATG CTG GCC GAC AAG ACT Met Leu Ala Asp Lys Thr 115		
GAG GAG AGC CAG GGC GTG Glu Glu Ser Gln Gly Val 130		
CGT GGC ATG GTG CGT CAG Arg Gly Met Val Arg Gln 145		
AAC GTG GAG GAG GCT CTG Asn Val Glu Glu Ala Leu 160		·

- (A) LENGTH: 199 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Met Ser Cys Gly Asn Ala Lys Ile Asn Ser Pro Ala Pro Ser Phe Glu

1 5 10 15

Glu Val Ala Leu Met Pro Asn Gly Ser Phe Lys Lys Ile Ser Leu Ser 20 25 30

Ser Tyr Lys Gly Lys Trp Val Val Leu Phe Phe Tyr Pro Leu Asp Phe 35 40 45

Ser Phe Val Cys Pro Thr Glu Val Ile Ala Phe Ser Asp Ser Val Ser
50 55 60

Arg Phe Asn Glu Leu Asn Cys Glu Val Leu Ala Cys Ser Ile Asp Ser 65 70 75 80

Glu Tyr Ala His Leu Gln Trp Thr Leu Gln Asp Arg Lys Lys Gly Gly
85 90 95

Leu Gly Thr Met Ala Ile Pro Met Leu Ala Asp Lys Thr Lys Ser Ile 100 105 110

Ala Arg Ser Tyr Gly Val Leu Glu Glu Ser Gln Gly Val Ala Tyr Arg 115 120 125

Gly Leu Phe Ile Ile Asp Pro His Gly Met Leu Arg Gln Ile Thr Val 130 135 140

Asn Asp Met Pro Val Gly Arg Ser Val Glu Glu Val Leu Arg Leu Leu 145 150 155 160

Glu Ala Phe Gln Phe Val Glu Lys His Gly Glu Val Cys Pro Ala Asn 165 170 175

Trp Lys Lys Gly Ala Pro Thr Met Lys Pro Glu Pro Asn Ala Ser Val 180 185 190

Glu Gly Tyr Phe Ser Lys Gln 195

- (2) INFORMATION FOR SEQ ID NO:25:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 637 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear

			_		CTC Leu			-									149
					GAC Asp												197
			•		GTG Val												245
					GAC Asp												293
					GGC Gly 95												341
					AGC Ser												389
					TAC Tyr												437
					ACC Thr												485
					CTG Leu												533
					GCG Ala 175												581
					TCT												623
TAA	ACCT	GTG 2	AGCG'	TCGC	AG G	AGTC	AGTG'	r GA	CCTC	ACCC	GCC'	rctg(	CCA (	GTGG	GTGCG	A	683
GAG	GGCG'	TGA (	GGGA'	TTGT	GG G	AAGG	CTGT	r gg	TATA	GATG	CAG	ACAG	CGA '	TGAA'	TGCAA	* . }	743
TCC	CACA	CAC '	TGGC	CCTC	CT C	AGCC	CTCT	C CA	CACA	GACA	CAC	GCAC	GCA '	TGTG			797

## (2) INFORMATION FOR SEQ ID NO:24:

## (i) SEQUENCE CHARACTERISTICS:

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									104						
Arg	Leu	Ser	Asp 180	Ala	Cys ·	Ser	Val	Pro 185	Asn	Cys	Lys	Lys	Cys 190	Glu	Thr
Gly	Thr	Ser 195	Arg	Leu	Cys	Ala	Glu 200	Cys	Asp	Thr	Gly	Tyr 205	Ser	Leu	Ser
Ala	Asp 210	Ala	Thr	Ser	Cys	Ser 215	Ser	Pro	Thr	Thr	Gln 220	Pro	Cys	Glu	Val
Glu 225	His.	Cys	Asn	Thr	Cys 230	Val	Asn	Gly	Asp	Ser 235	Thr	Arg	Cys	Ala	Tyr 240
Cys	Asn	Thr	Gly	Tyr 245	Tyr	Val	Ser	Asp	Gly 250	Lys	Cys	Lys	Ala	Met 255	Gln
Gly	Cys	Tyr	Val 260	Ser	Asn	Cys	Ala	Gln 265	Cys	Met	Leu	Leu	Asp 270	Ser	Thr
Lys	Cys	Ser 275		Cys	Val	Lys	Gly 280	Tyr	Leu	Leu	Thr	Ser 285	Ser	Tyr	Ser
Cys	Val 290	Ser	Gln	Lys	Val	Ile 295	Asn	Ser	Ala	Ala	Ala 300	Pro	Tyr	Ser	Leu

Trp Val Ala Ala Ala Val Leu Leu Thr Ser Phe Ala Met His Leu Ala

### (2) INFORMATION FOR SEQ ID NO:23:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 797 base pairs
  - (B) TYPE: nucleic acid

310

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major
- (ix) FEATURE:

305

- (A) NAME/KEY: CDS
- (B) LOCATION: 27..623
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

CTGTACTTTA TTGCCACCAG CCAGCC ATG TCC TGC GGT AAC GCC AAG ATC AAC Met Ser Cys Gly Asn Ala Lys Ile Asn

TCT CCC GCG CCG TCC TTC GAG GAG GTG GCG CTC ATG CCC AAC GGC AGC Ser Pro Ala Pro Ser Phe Glu Glu Val Ala Leu Met Pro Asn Gly Ser 10 15 20 25

101

CCAC	AGTO	CCC F	CAGO	CGCAC	SA CA	ACACO	GTGT	TAI	AACGO	GCGC	ÄGG	CATCO	CCT .	CCCT	ATCACT
TCAT	TTCT	CC 1	DAAA	CCAC	CT C	ACCAI	GTC	G CAC	CACCO	CCC	TCC	CCAT	rcg	GCCG	CCTTC
CGGG	CGC	AGC 1	GTGC	CGGAZ	AT GO	GTGT	rgtgo	TCC	FACCI	CGT	TCCI	:GGC	AGC	TCAC	rcgcat
GTGI	ACAC	GCC P	CTC	CAACO	CA CO	JAAAE	GCTCT	CTI	CTG	CGCA	CATA	LAAA	AA	AAAA	AAAAA
AAAA	ACTO	GA C	GGGG	GGCC	CC GC	STACO	CAA	Ą		,				;	٠
(2)	INFO	RMAT	NOI	FOR	SEO	ID N	10:22	2:					•		•
,								rics:	:	ů.					
		, _	(A)	LEN	GTH:		ami	ino a	cids	5					
						3Y: ]									
	(i	.i) N	OLEC	CULE	TYPE	E: pı	rotei	in	•		•				
	()	ci) S	SEQUE	ENCE	DESC	CRIPT	CION	: SEC	) ID	NO:2	22:				
Val 1	Leu	Pro	Asp	Met 5	Thr	Cys	Ser	Leu	Thr 10	Gly	Leu	Gln	Cys	Thr 15	Asp
Pro	Asn	Cys	Lys 20	Thr	Cys	Thr	Thr	Tyr 25	Gly	Gln	Cys	Thr	Asp 30	Cys	Asn
Asp	Gly	Tyr 35	Gly	Leu	Thr	Ser	Ser 40	Ser	Val	Cys	Val	Arg 45	Суз	Ser	Val
Ala	Gly 50	Cys	Lys	Ser	Cys	Pro 55	Val	Asp	Ala	Asn	Val 60	Cys	Lys	Val	Cys
Leu 65	Gly	Gly	Ser	Glu	Pro 70	Ile	Asn	Asn	Met	Cys 75	Pro	Cys	Thr	Asp	Pro 80
Asn	Cys	Ala	Ser	Cys 85	Pro	Ser	Asp	Ala	Gly 90	Thr	Cys	Thr	Gln	Cys 95	Ala
Asn	Gly	Tyr	Gly 100	Leu	Val	Asp	Gly	Ala 105	Cys	Val	Arg	Cys	Gln 110	Glu	Pro
Asn	Cys	Phe 115	Ser	Cys	Asp	Ser	Asp 120	Ala	Asn	Lys	Cys	Thr 125	Gln	Cys	Ala
Pro	Asn 130	Tyr	Tyr	Leu	Thr	Pro 135	Leu	Leu	Thr	Cys	Ser 140	Pro	Val	Ala	Cys
Asn 145	Ile	Glu	His	Cys	Met 150	Gln	Суз	Asp	Pro	Gln 155	Thr	Pro	Ser	Arg	Cys 160
Gln	Glu	Cys	Val	Ser 165	Pro	Tyr	Val	Val	Asp 170	Ser	Tyr	Asp	Gly	Leu 175	Cys

Pro	Val	Ala	Cys	Asn 145		Glu	His	Cys	Met 150		Ċys	Asp	Pro	Gln 155	Thr	•	
CCG Pro	TCG Ser	CGC	TGC Cys 160	CAG Gln	GAG Glu	TGC Cys	GTG Val	TCC Ser 165	Pro	TAC Tyr	GTG Val	GTT Val	GAC Asp 170	Ser	TAC		529
GAC Asp	GGC Gly	CTC Leu 175	Cys	AGG Arg	CTC Leu	TCC Ser	GAT Asp 180	GCC	TGC Cys	TCC Ser	GTG Val	CCC Pro 185	AAC Asn	TGC Cys	AAG Lys		577
AAG Lys	TGC Cys 190	GAG Glu	ACC Thr	GGT Gly	ACC Thr	TCC Ser 195	AGG Arg	CTC Leu	TGC Cys	GCC Ala	GAG Glu 200	TGC Cys	GAC Asp	ACC	GGC		625
TAC Tyr 205	Ser	CTC Leu	TCC	GCC Ala	GAC Asp 210	GCG Ala	ACG Thr	AGC Ser	TGC Cys	AGC Ser 215	AGT Ser	CCA Pro	ACC Thr	ACG Thr	CAG Gln 220	.*	673
CCG Pro	TGC Cys	GAG Glu	GTG Val	GAG Glu 225	CAC His	TGC Cys	AAC Asn	ACA Thr	TGT Cys 230	GTG Val	AAC Asn	GGC Gly	GAT Asp	AGC Ser 235	ACC Thr		721
CGC Arg	TGT Cys	GCC Ala	TAC Tyr 240	TGC Cys	AAC Asn	ACC Thr	GGC Gly	TAC Tyr 245	TAC Tyr	GTC Val	TCC Ser	GAT Asp	GGC Gly 250	AAG Lys	TGC Cys		769
AAG Lys	GCC Ala	ATG Met 255	CAG Gln	GGC Gly	TGC Cys	TAC Tyr	GTG Val 260	TCG Ser	AAC Asn	TGC Cys	GCG Ala	CAG Gln 265	TGC Cys	ATG Met	CTG Leu		817
Leu	GAC Asp 270	AGC Ser	ACC Thr	AAG Lys	TGC Cys	TCC Ser 275	ACG Thr	TGC Cys	GTG Val	AAA Lys	GGG Gly 280	TAC Tyr	CTG Leu	CTC Leu	ACG Thr		865
TCG Ser 285	TCC Ser	TAC Tyr	AGT Ser	Cys	GTC Val 290	TCG Ser	CAG Gln	AAA Lys	Val	ATC Ile 295	AAC Asn	AGT Ser	GCG Ala	Ala	GCG Ala 300		913
CCC Pro	TAC Tyr	TCT Ser	Leu	TGG Trp 305	GTG Val	GCC Ala	GCC Ala	GCC Ala	GTG Val 310	CTC Leu	CTC Leu	ACC Thr	Ser	TTT Phe 315	GCC Ala		961
ATG Met	CAC His	Leu	GCA Ala 320	TAGT	GCGC	AG C	GGCA	TGCG	A AC	AACC	CCAC	TCT	CATT	CTC		10	013 ·
CAAC	ATGT	GC A	TACA	CACA	C AC	ACAG	ACAG	CGG	GGCA	GCA	cccc	CTCC	CC A	CACA	CACAC	10	073
ACGC.	ACTT	cc c	CCTT	GTCT'	r Gt	TCTT	CTTT	CCT	CGTT	CGC .	ATTT	СТТТ	CT C	TCGT	GCGCT	11	133
3GCG	CCGG	CC T	CCTG	CACG'	r CG	CTCC	CCTC	CCC	CTAA	CCT	CTAT	TCTC'	TC T	CTCT	CTCTC	13	193
CTC	GCCG	GC A	TCAT	TGCT	r cr	TACC	CTTT	тст	GATC	CTT (	GCTC	GCGTC	3G G	ന്ദ്രേച്ച	<sup>ሮ</sup> ል ሮጥር፤	1-	152

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									101					*	
	(i)	(A (B (C	L) LE 3) TY 2) SI	EE CHENGTHE PE: TRANI OPOLO	H: 15 nucl	23 b .eic ESS:	ase acid	pair l	cs.		•				
	(ii)	MOL	ECUL	E TY	PE:	CDNA		* 4							
	(vi)			AL SC RGANI			shma	ınia	majo	r					
	(ix)	( A		: ME/I CATI			973								
	(xi)	SEÇ	UENC	CE DE	ESCRI	PTIC	N: S	SEQ ]	D NO	:21:			. •		
GAAT	TCGG	CA C							ACA T						49
	TGC Cys														97
	GAC Asp 30														145
	TGC Cys														193
	AAA Lys														241
	ACC Thr														289
	CAG Gln														337
	CAG Gln 110							14							385
	CAA Gln					Tyr									433

CCG GTG GCC TGC AAC ATC GAG CAC TGC ATG CAG TGC GAC CCA CAG ACG

481

											• .					
TG:	rgta(	GCCA	TGG	CTTC	AGG	AGAG	AAAA	CA A	AATA	CAAG	G AA	AGGC	'AATA	TGI	'AACTI	ATG
GG	GTTC(	CCTT	TTT	TACT	ATG	CAAA	GTTT	TT A	TAAC'	TCCT	G AT	CGGC	AAAA	ACA	ACAAC	AA
CCC	GCCA'	TACA	CCA	AGAG	CAA .	ATGC'	TTTC	TT C	TGCG	GACT	G TG	CTTC	TGTT	TTT	TTTTA	TG
AAC	GAG	IGAC	TCG	CGCG.	ATG	AAAA	GTGT	GT G	CGTG	3GAG	A TG'	TATT	TCCT	TTT	TTTGT	`TC
ATA	AGTGO	GCGA	CAG	CTCA	CTG '	TTGA(	CGAT	GA C	<b>LAAA</b>	(AAA	A AA	AAAA	AAAA	CTC	GAG	
(2)	INE	ORM	OITA	V FOI	R SE	Q ID	NO:2	20:								
:		(i)	( <i>I</i>	1) LI 3) TY	ENGTI (PE :	ARACT H: 15 amir OGY:	o ac	nino cid		ls						
	. (	ii)	MOLE	CULE	TYP	E: p	rote	in	•							•
	. (	xi)	SEQU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	20:					
Met 1	Ser	Ser	Glu	Arg	Thr	Phe	Île	Ala	Val		Pro	Asp	Gly	Val	Gln	
Arg	Gly	Leu	Val 20	Gly	Glu	Ile	Ile	Ala 25	Arg	Phe	Glu	Arg	Lys 30		Tyr	-
Lys	Leu	Val 35	Ala	Leu	Lys	Ile	Leu 40	Gln	Pro	Thr	Thr	Glu 45		Ala	Gln	•
Gly	His 50	Tyr	Lys	Asp	Leu	Cys 55	Ser	Lys	Pro	Phe	Phe 60	Pro	Ala	Leu	Val	
Lys 65	Tyr	Phe	Ser	Ser	Gly 70	Pro	Ile	Val	Cys	Met 75	Val	Trp	Glu	Gly	Lys 80	
Asn	Val	Val	Lys	Ser 85	Gly	Arg	Val	Leu	Leu 90	Gly	Ala	Thr	Asn	Pro 95	Ala	
Asp	Ser	Gln	Pro 100	Gly	Thr	Ile	Arg	Gly 105	Asp	Phe	Ala	Val	Asp 110	Val	Gly	
Arg	Asn	Val 115	Cys	His	Gly	Ser	Asp 120	Ser	Val	Glu	Ser	Ala 125	Glu	Arg	Glu	
Ilė	Ala 130	Phe	Trp	Phe	Lys	Ala 135	Asp	Glu	Ile	Ala	Ser 140	Trp	Thr	Ser	His	
Ser 145	Val	Ser	Gln	Ile	Tyr 150	Glu		•				• •				

(2) INFORMATION FOR SEQ ID NO:21:

# (ix) FEATURE:

(A) NAME/KEY: CDS(B) LOCATION: 71..523

GAA'			QUENC					-				ATTC	ACT (	CTTCC	ATTTC	60
		ACC 1	ATG I	rcc 1	cc c	ag c	CGC A	ACC I	TTT A	ATT C	GCC C	TC	AAG (	ccg (	BAC	109
			1				5					10			-	
			CGC Arg													157
			AAG Lys													205
			GGT Gly		•								_			253
			AAG Lys 65													301
			AAC Asn										Gly	_		349
			GAC Asp													397
	Val		CGC Arg													445
			ATC Ile												Trp	493
			TCC Ser 145	Val		-			Glu		CGGT	GAT	TGCG	GACA	CG	543
CTT	TGAG	GAC	GTAG	CTGT	AC C	CCCA	ATGA	A TT	CTTC	TCTG	AAA	ACCA	CAT	CATA	AGCCTC	60
TTA	AGAG	GTT	ATTT	TTCT	TG A	TCGA	TGCC	C <sub>_</sub> GG	TGGT	GACC	AGC	ACCA	TTC	CTTT	ATCGGA	66
ጥጥረ	י אַ ריידיר	מיממי	כידיכיכי	тасс	GD D	ጥሮልጥ	ርጥልር	т сс	GGTG	AGAG	TGG	GCTC	TGG	AGGA	GACTGT	72

Lys	His	Gly	Leu	Glu	Val	Ile	Tyr	Met	Ile	Glu	Pro	Ile	Asp	Glu	Tyr
		515					520					525			-

- Cys Val Gln Gln Leu Lys Glu Phe Glu Gly Lys Thr Leu Val Ser Val 530 535 540
- Thr Lys Glu Gly Leu Glu Leu Pro Glu Asp Glu Glu Glu Lys Lys Lys 545 550 555 560
- Gln Glu Glu Lys Lys Thr Lys Phe Glu Asn Leu Cys Lys Ile Met Lys
  565 570 575
- Asp Ile Leu Glu Lys Lys Val Glu Lys Val Val Val Ser Asn Arg Leu
  580 585 590
- Val Thr Ser Pro Cys Cys Leu Val Thr Ser Thr Tyr Gly Trp Thr Ala 595 600 605
- Asn Met Glu Arg Ile Met Lys Ala Gln Ala Leu Arg Asp Asn Ser Thr 610 615 620
- Met Gly Tyr Met Ala Ala Lys Lys His Leu Glu Ile Asn Pro Asp His 625 630 635 640
- Ser Ile Ile Glu Thr Leu Arg Gln Lys Ala Glu Ala Asp Lys Asn Asp 645 650 655
- Lys Ser Val Lys Asp Leu Val Ile Leu Leu Tyr Glu Thr Ala Leu Leu 660 665 670
- Ser Ser Gly Phe Ser Leu Glu Asp Pro Gln Thr His Ala Asn Arg Ile 675 680 685
- Tyr Arg Met Ile Lys Leu Gly Leu Gly Ile Asp Glu Asp Asp Pro Thr 690 695 700
- Ala Asp Asp Thr Ser Ala Ala Val Thr Glu Glu Met Pro Pro Leu Glu 705 710 715 720
- Gly Asp Asp Asp Thr Ser Arg Met Glu Glu Val Asp
  725 730

### (2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1019 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

Glu 225	Arg	Asp	Lys	Glu	Val 230	Ser	Asp	Asp	Glu	Ala 235	Glu	Glu	Lys	Glu	Asp 240
Lys	Glu	Glu	Glu	Lys 245	Glu	Lys	Glu	Glu	Lys 250	Glu	Ser	Glu	Asp	Lys 255	Pro
Glu	Ile	Glu	Asp 260	Val	Gly	Ser	_		Glu	Asp	Glu	Lys	Lys 270	Asp	Gly
Asp	Lys	Lys 275	Lys	Lys	Lys	Lys	Ile 280	Lys	Glu	Lys	Tyr	Ile 285	Asp	Lys	Glu
Glu	Leu 290	Asn	Lys	Thr	Lys	Pro 295	Ile	Trp	Thr	Arg	Asn 300	Pro	Asp	Asp	Ile
Thr 305	Asn	Glu	Glu	Tyr	Gly 310	Glu	Phe	Tyr	Lys	Ser	Leu	Thr	Asn	Asp	Trp 320
Glu	Asp	His	Leu	Ala 325	Val	Lys	His	Phe	Ser 330	Val	Glu	Gly	Gln	Leu 335	Glu
Phe	Arg	Ala	Leu 340	Leu	Phe	Val	Pro	Arg 345	Arg	Ala	Pro	Phe	Asp 350	Leu	Phe
Glu	Asn	Arg 355	Lys	Lys	Lys	Asn	Asn 360	Ile	Lys	Leu	Tyr	Val 365	Arg	Arg	Val
Phe	Ile 370	Met	Asp	Asn	Cys	Glu 375	Glu	Leu	Ile	Pro	Glu 380	Tyr	Leu	Asn	Phe
Ile 385	Arg	Gly	Val	Val	Asp 390	Ser	Glu	Asp	Leu	Pro 395	Leu	Asn	Ile	Ser	Arg 400
				405		:			410				Lys	415	
Val	Lys	Lys	Cys 420	Leu	Glu	Leu	Phe	Thr 425	Glu	Leu	Ala	Glu	Asp 430	Lys	Glu
Asn	Tyr	Lys 435	Lys	Phe	Tyr	Glu	Gln 440	Phe	Ser	Lys	Asn	Ile 445	Lys	Leu	Gly
	His 450	Glu	Asp	Ser	Gln	Asn 455	Arg	Lys	Lys	Leu	Ser 460	Glu	Leu	Leu	Arg
Tyr 465	_	Thr	Ser	Ala	Ser 470	Gly	Asp	Glu	Met	Val 475	Ser	Leu	Lys	Asp	Tyr 480
			,	485				_	490				Ile	495	
Glu	Thr	Lys	Asp 500	Gln	Val	Ala	Asn	Ser 505		Phe	Val	Glu	Arg 510	Leu	Arg

## (2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 732 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:
- Met Pro Glu Glu Thr Gln Thr Gln Asp Gln Pro Met Glu Glu Glu Glu 1 5 10 15
- Val Glu Thr Phe Ala Phe Gln Ala Glu Ile Ala Gln Leu Met Ser Leu 20 25 30
- Ile Ile Asn Thr Phe Tyr Ser Asn Lys Glu Ile Phe Leu Arg Glu Leu 35 40 45
- Ile Ser Asn Ser Ser Asp Ala Leu Asp Lys Ile Arg Tyr Glu Ser Leu 50 55 60
- Thr Asp Pro Ser Lys Leu Asp Ser Gly Lys Glu Leu His Ile Asn Leu 65 70 75 80
- Ile Pro Asn Lys Gln Asp Arg Ala Leu Thr Ile Val Asp Thr Gly Ile 85 90 95
- Gly Met Thr Lys Ala Asp Leu Ile Asn Asn Leu Gly Thr Ile Ala Lys
  100 105 110
- Ser Gly Thr Lys Ala Phe Met Glu Ala Leu Gln Ala Gly Ala Asp Ile 115 120 125
- Ser Met Ile Gly Gln Phe Gly Val Gly Phe Tyr Ser Ala Tyr Leu Val 130 135 140
- Ala Glu Lys Val Thr Val Ile Thr Lys His Asn Asp Asp Glu Gln Tyr 145 150 155 160
- Ala Trp Glu Ser Ser Ala Gly Gly Ser Phe Thr Val Arg Thr Asp Thr
  165 170 175
- Gly Glu Pro Met Gly Arg Gly Thr Lys Val Ile Leu His Leu Lys Glu 180 185 190
- Asp Gln Thr Glu Tyr Leu Glu Glu Arg Arg Ile Lys Glu Ile Val Lys 195 200 205
- Lys His Ser Gln Phe Ile Gly Tyr Pro Ile Thr Leu Phe Val Glu Lys 210 215 220

								) )							
Gly	Lys	Asn	Val 420	Lys	Leu	Gly	Ile	His 425	Glu	Asp	Ser	Ala	Asn 430	Arg	Lys
Lys	Leu	Met 435	Glu	Leu	Leu	Arg	Phe 440	His	Ser	Ser	Glu	Ser 445	Gly	Glu	Asp
Met	Thr 450	Thr	Leu	Lys	Asp	Tyr 455	Val	Thr	Arg	Met	Lys 460	Glu	Gly	Gln	Lys
Cys 465	Ile	Tyr	Tyr	Val	Thr 470	Gly	Asp	Ser	Lys	Lys 475	Lys	Leu	Glu	Thr	Ser 480
Pro	Phe	Ile	Glu	Gln 485	Ala	Arg	Arg	Arg	Gly 490		Glu	Val	Leu	Phe 495	Met
Thr	Glu	Pro	Ile 500	Asp	Glu	Tyr	Val	Met 505	Gln	Gln	Val	Lys	Asp 510	Phe	Glu
Asp	Lys	Lys 515	Phe	Ala	Cys	Leu	Thr 520	Lys	Glu	Gly	Val	His 525	Phe	Glu	Glu
Thr	Glu 530	Glu	Glu	Lys	Lys	Gln 535	Arg	Glu	Glu	Glu	Lys 540	Thr	Ala	Tyr	Glu
Arg <b>5</b> 45	Leu	Cys	Lys	Ala	Met 550	Lys	Asp	Val	Leu	Gly 555	Asp	Lys	Val	Glu	Lys 560
Val	Val	Val	Ser	Glu 565	Arg	Leu	Ala	Thr	Ser 570	Pro	Cys	Ile	Leu	Val 575	Thr
Ser	Glu-	Phé	Gly 580	Trp	Seř	Ala	His	Met 585	Glu	Gln	Ile	Met	Arg 590	Asn	Gln
Ala	Leu	Arg 595	Asp	Ser	Ser	Met	Ser 600	Ala	Tyr	Met	Met	Ser 605	Lys	Lys	Thr
Met	Glu 610	Ile	Asn	Pro	Ala	His 615	Pro	Ile	Val	Lys	Glu 620	Leu	Lys	Arg	Arg
Val 625	Glu	Ala	Asp	Glu	Asn 630	Asp	Lys	Ala	Val	Lys 635		Leu	Val	Tyr	Leu 640
Leu	Phe	Asp	Thr	Ala 645		Leu	Thr	Ser	Gly 650		Thr	Leu	Asp	Asp 655	
Thr	Ser	Tyr	Ala 660		Arg	Ile	His	Arg 665		Ile	Lys	Leu	Gly 670	Leu	Ser
Leu	Asp	Asp 675		Asp	Asn	Gly	Asn 680		Glu	Ala	Glu	Pro 685		Ala	Ala
Val	Pro 690	Ala	Glu	Pro	Val	Ala 695		Thr	Ser	Ser	Met		Gln	Val	Asp

Va]	l Ala 130	a Ası	Ar	g Vai	l Thi	r Va. 135	l Val	l Sei	r Ly:	s Ası	n Ası 140		o As	p Gl	u Ala
Ty1	Thr	Tr	Glı	u Sei	Sei 150	Ala	a Gly	/ Gly	/ Thi	r Phe 159		c Val	l Th:	r Pro	Thr 160
Pro	Asp	Cys	a Asp	165	ı Lys	Arg	g Gly	Thr	170		⊵ Va]	l Let	His	5 Let 175	Lys
Glu	Asp	Glr	180	ı Glu	ı Tyr	Leu	Glu	Glu 185	arg	J Arg	J Leu	Lys	190		ı Ile
Lys	Lys	His 195	Ser	Glu	Phe	Ile	Gly 200	Tyr	Asp	) Ile	Glu	Leu 205		: Val	Glu
	210					215					220				Ala
223				Glu	230					235					240
				245					250					255	Lys <sub>.</sub>
		•	260					265					270	•	
		.2 1,5		,			280				-	285			Tyr
	290			Asn		295	•				300				
303				Gln	310	*.				315				·	320
				Asp 325					330					335	
			340	Arg				345					350		
		355		Leu			360					365			
	370			Ile		375					380				
363				Lys	390	•				395					400
Glu	Ile	Ala	Glu	Asn 405	Lys	Glu	Asp	Tyr	Lys 410	Lys	Phe	Tyr	Glu	Gln 415	Phe

Asn Pro Lys His Pro Ile Ile Lys Glu Leu Arg Arg Arg Val Glu Ala 610 615 620

Asp Glu Asn Asp Lys Ala Val Lys Asp Leu Val Phe Leu Leu Phe Asp 625 630 635 640

Thr Ser Leu Leu Thr Ser Gly Phe Gln Leu Glu Asp Pro Thr Tyr Ala 645 650 655

Glu Arg Ile Asn Arg Met Ile Lys Leu Gly Leu Ser Leu Asp Glu Glu 660 665 670

Glu Glu Glu Ala Val Glu Ala Ala Val Ala Glu Thr Ala Pro Ala 675 680 685

Glu Val Thr Ala Gly Thr Ser Ser Met Glu Leu Val Asp 690 695 700

### (2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 704 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Met Thr Glu Thr Phe Ala Phe Gln Ala Glu Ile Asn Gln Leu Met Ser 1 5 10 15

Leu Ile Ile Asn Thr Phe Tyr Ser Asn Lys Glu Ile Phe Leu Arg Glu 20 25 30

Leu Ile Ser Asn Ala Ser Asp Ala Cys Asp Lys Ile Arg Tyr Gln Ser 35 40 45

Leu Thr Asn Gln Ala Val Leu Gly Asp Glu Ser His Leu Arg Ile Arg 50 55 60

Val Val Pro Asp Lys Ala Asn Lys Thr Leu Thr Val Glu Asp Thr Gly
65 70 75 80

Ile Gly Met Thr Lys Ala Glu Leu Val Asn Asn Leu Gly Thr Ile Ala 85 90 95

Arg Ser Gly Thr Lys Ala Phe Met Glu Ala Leu Glu Ala Gly Gly Asp 100 105 110

Met Ser Met Ile Gly Gln Phe Gly Val Gly Phe Tyr Ser Ala Tyr Leu 115 120 125

Phe	Asp	Met	Leu	Glu 325		Asn	Lys	Lys	Arg 33.0		Asn	Ile	Lys	Leu 335	-
Val	Arg	Arg	Val 340		Ile	Met	Asp	Asn 345	Cys	Glu	Asp	Leu	Cys 350		Ası
Trp	Leu	Gly 355	Phe	Val	Lys	Gly	Val 360		Asp	Ser	Glu	Asp 365	Leu	Pro	Let
Asn	11e 370	Ser	Arg	Glu	Asn	Leu 375	Gln	Gln	Asn	Lys	Ile 380	Leu	Lys	Val	Ile
Arg 385	Lys	Asn	Ile	Val	Lys 390	Lys	Cys	Leu	Glu	Met 395	Phe	Glu	Glu	Val	Ala 400
Glu	Asn	Lys	Glu	Asp 405	Tyr	Lys	Gln	Phe	Tyr 410	Glu	Gln	Phe	Gly	Lys 415	Asn
Ile	Lys	Leu	Gly 420	Ile	His	Glu	Asp	Thr 425	Ala	Asn	Arg	Lys	Lys 430	Leu	Met
Glu	Leu	Leu 435	Arg	Phe	Tyr	Ser	Thr 440	Glu	Ser	Gly	Glu	Val 445	Met	Thr	Thr
Leu	Lys 450	Asp	Tyr	Val	Thr	Arg 455	Met	Lys	Ala	Glu	Gln 460	Asn	Ser	Ile	Tyr
Tyr 465	Ile	Thr	Gly	Asp	Ser 470	Lys	Lys	Lys	Leu	Glu 475	Ser	Ser	Pro	Phe	Ile 480
		-		485					490				Thr	495	
Гуr	Asp	Glu	Tyr 500	Val	Met	Gln	Gln	Val 505	Lys	Asp	Phe	Glu	Asp 510	Lys	Lys
Phe	Ala	Cys 515	Leu	Thr	Lys	Glu	Gly 520	Val	His	Phe	Glu	Glu 525	Ser	Glu	Glu
Slu	Lys 530	Lys	Gln	Arg	Glu	Glu 535.		Lys	Ala	Thr	Cys 540	Glu	Lys	Leu	Cys
Lys 545	Thr	Met	Lys		Val 550	Leu	Gly	Asp	Lys	Val 555	Glu	Lys	Val	Thr	Val 560
Ser	Glu	Arg	Leu	Ser 565	Thr	Ser	Pro	Cys	Ile 570	Leu	Val	Thr	Ser	Glu 575	Phe
Sly	Trp	Ser	Ala 580	His	Met	Glu	Gln	Met 585	Met	Arg	Asn	Gln	Ala 590	Leu	Arg
Asp		Ser 595		Ala	Gln		Met 600	Met	Ser	Lys		Thr	Met	Glu	Leu

20 25 30 Val Ile Ser Asn Ala Ser Asp Ala Cys Asp Lys Ile Arg Tyr Gln Ser Leu Thr Asp Pro Ala Val Leu Gly Asp Ala Thr Arg Leu Cys Val Arg Val Val Pro Asp Lys Glu Asn Lys Thr Leu Thr Val Glu Asp Asn Gly 70 75 Ile Gly Met Thr Lys Ala Asp Leu Val Asn Asn Leu Gly Thr Ile Ala 85 90 Arg Ser Gly Thr Lys Ala Phe Met Glu Ala Leu Glu Ala Gly Ala Asp 100 Met Ser Met Ile Gly Gln Phe Gly Val Gly Phe Tyr Ser Ala Tyr Leu 115 Val Ala Asp Arg Val Thr Val Thr Ser Lys Asn Asn Ser Asp Glu Val 135 Tyr Val Trp Glu Ser Ser Ala Gly Gly Thr Phe Thr Ile Thr Ser Ala 150 155 Pro Glu Ser Asp Met Lys Leu Pro Ala Arg Ile Thr Leu His Leu Lys 170 165 Glu Asp Gln Leu Glu Tyr Leu Glu Ala Arg Arg Leu Lys Glu Leu Ile Lys Lys His Ser Glu Phe Ile Gly Tyr Asp Ile Glu Leu Met Val Glu 200 Lys Thr Thr Glu Lys Glu Val Thr Asp Glu Asp Glu Glu Glu Ala Lys 210 215 Lys Ala Asp Glu Asp Gly Glu Glu Pro Lys Val Glu Glu Val Thr Glu 225 Gly Glu Glu Asp Lys Lys Lys Thr Lys Lys Val Lys Glu Val Thr Lys Glu Tyr Glu Val Gln Asn Lys His Lys Pro Leu Trp Thr Arg Asp 260 265 270 Pro Lys Asp Val Thr Lys Glu Glu Tyr Ala Ala Phe Tyr Lys Ala Ile 275 280 Ser Asn Asp Trp Glu Asp Pro Pro Ala Thr Lys His Phe Ser Val Glu 295

Gly Gln Leu Glu Phe Arg Ala Ile Met Phe Val Pro Lys Arg Ala Pro

315

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Lys Val Phe Asp Glu 1 5

- (2) INFORMATION FOR SEQ ID NO:14:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 30 base pairs
    - (B) TYPE: nucleic acid
      - (C) STRANDEDNESS: single
      - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

GGATCCATGG TCAAGTCCCA CTACATCTGC

30

- (2) INFORMATION FOR SEQ ID NO:15:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 33 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GAATTCAGAC CGGATAGAAA TAAGCCAATG AAA

33

- (2) INFORMATION FOR SEQ ID NO:16:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 701 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Met Thr Glu Thr Phe Ala Phe Gln Ala Glu Ile Asn Gln Leu Met Ser

1 10 15

Leu Ile Ile Asn Thr Phe Tyr Ser Asn Lys Glu Ile Phe Leu Arg Asp

- (ix) FEATURE:
  - (A) NAME/KEY: Modified-site
  - (B) LOCATION: 6
- (D) OTHER INFORMATION: /note= "Where Xaa is either a Leu or Lys Residue"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Xaa Gln Xaa Pro Gln Xaa Val Phe Asp Glu Xaa Xaa 1 5 10

- (2) INFORMATION FOR SEQ ID NO:12:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 26 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ix) FEATURE:
    - (A) NAME/KEY: -
    - (B) LOCATION: 11
    - (D) OTHER INFORMATION: /note= "Where n is inosine"
  - (ix) FEATURE:
    - (A) NAME/KEY: -
    - (B) LOCATION: 17
    - (D) OTHER INFORMATION: /note= "Where n is inosine"
  - (ix) FEATURE:
    - (A) NAME/KEY: -
    - (B) LOCATION: 20
    - (D) OTHER INFORMATION: /note= "Where n is inosine"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

## GGAATTCCCC NCAGCTNGTN TTCGAC

- (2) INFORMATION FOR SEQ ID NO:13:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 5 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear

165

170

175

Leu Asp Glu Ala Asp Glu Met Leu Ser Gln Gly Phe Ala Asp Gln Ile 180 185 190

Tyr Glu Ile Phe Arg Phe Leu Pro Lys Asp Ile Gln Val Ala Leu Phe 195 200 205

Ser Ala Thr Met Pro Glu Glu Val Leu Glu Leu Thr Lys Lys Phe Met 210 215 220

Arg Asp Pro Val Arg Ile Leu Val Lys Arg Glu Ser Leu Thr Leu Glu 225 230 235 240

Gly Ile Lys Gln Phe Phe Ile Ala Val Glu Glu Glu His Lys Leu Asp 245 250 255

Thr Leu Met Asp Leu Tyr Glu Thr Val Ser Ile Ala Gln Ser Val Ile 260 265 270

Phe Ala Asn Thr Arg Arg Lys Val Asp Trp Ile Ala Glu Lys Leu Asn 275 280 285

Gln Ser Asn His Thr Val Ser Ser Met His Ala Glu Met Pro Lys Ser 290 295 300

Asp Arg Glu Arg Val Met Asn Thr Phe Arg Ser Gly Ser Ser Arg Val 305 310 315 320

Leu Val Thr Thr Asp Leu Val Ala Arg Gly Ile Asp Val His His Val 325 330 335

Asn Ile Val Ile Asn Phe Asp Leu Pro Thr Asn Lys Glu Asn Tyr Leu 340 345 350

His Arg Ile Gly Arg Gly Gly Arg Tyr Gly Val Lys Gly Val Ala Ile 355 360 365

Asn Phe Val Thr Glu Lys Asp Val Glu Leu Leu His Glu Ile Glu Gly 370 380

His Tyr His Thr Gln Ile Asp Glu Leu Pro Val Asp Phe Ala Ala Tyr 385 390 395 400

Leu Gly Glu

# (2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 12 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

AGAGAACGCA	CATCGTAACA	CAGCCACGCG	AACGATAGTA	AGGGCGTGCG	GCGGCGTTCC	1426
CCTCCTCCTG	CCAGCGGCCC	CCCTCCGCAG	CGCTTCTCTT	TTGAGAGGGG	GGCAGGGGGA	1486
GGCGCTGCGC	CTGGCTGGAT	GTGTGCTTGA	GCTTGCATTC	CGTCAAGCAA	GTGCTTTGTT	1546
TTAATTATGC	GCGCCGTTTT	GTTGCTCGTC	CCTTTCGTTG	GTGTTTTTC	GGCCGAAACG	1606
GCGTTTAAAG	CA `					1618

#### (2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 403 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Met Ser Gln Gln Asp Arg Val Ala Pro Gln Asp Gln Asp Ser Phe Leu

Asp Asp Gln Pro Gly Val Arg Pro Ile Pro Ser Phe Asp Asp Met Pro 25

Leu His Gln Asn Leu Leu Arg Gly Ile Tyr Ser Tyr Gly Phe Glu Lys 35

Pro Ser Ser Ile Gln Gln Arg Ala Ile Ala Pro Phe Thr Arg Gly Gly 50 . 55

Asp Ile Ile Ala Gln Ala Gln Ser Gly Thr Gly Lys Thr Gly Ala Phe

Ser Ile Gly Leu Leu Gln Arg Leu Asp Phe Arg His Asn Leu Ile Gln 90

Gly Leu Val Leu Ser Pro Thr Arg Glu Leu Ala Leu Gln Thr Ala Glu 100

Val Ile Ser Arg Ile Gly Glu Phe Leu Ser Asn Ser Ala Lys Phe Cys 115

Glu Thr Phe Val Gly Gly Thr Arg Val Gln Asp Asp Leu Arg Lys Leu 135

Gln Ala Gly Val Val Val Ala Val Gly Thr Pro Gly Arg Val Ser Asp 150 155 160

Val Ile Lys Arg Gly Ala Leu Arg Thr Glu Ser Leu Arg Val Leu Val

	GAG Glu	ATC Ile 195	Phe	CGC Arg	TTC Phe	CTG Leu	CCG Pro 200	AAG Lys	GAC Asp	Ile	CAG Gln	GTC Val 205	Ala	CTC Lev	TTC Phe	TCC Ser		741
	GCC Ala 210	Thr	ATG Met	CCG Pro	GAG Glu	GAG Glu 215	GTG Val	CTG Leu	GAG Glu	CTG Leu	ACA Thr 220	Lys	AAG Lys	TTC Phe	ATG Met	CGC Arg 225		789
	GAC Asp	CCC Pro	GTA Val	CGC Arg	ATT Ile 230	CTC Leu	GTG Val	AAG Lys	CGC	GAG Glu 235	AGC Ser	CTG Leu	ACG Thr	CTG Leu	GAG Glu 240	GGC Gly		837
*	ATC Ile	AAG Lys	CAG Gln	TTC Phe 245	TTC Phe	ATC Ile	GCC Ala	GTC Val	GAG Glu 250	GAG Glu	GAG Glu	CAC	AAG Lys	CTG Leu 255	GAC Asp	ACG Thr		885
	CTG Leu	ATG Met	GAC Asp 260	CTG Leu	TAC Tyr	GAG Glu	ACC Thr	GTG Val 265	TCC Ser	ATC Ile	GCG Ala	CAG Gln	TCC Ser 270	GTC Val	ATC Ile	TTC Phe		933
	GCC Ala	AAC Asn 275	ACC Thr	CGC Arg	CGC Arg	AAG Lys	GTG Val 280	GAC Asp	TGG Trp	ATC: Ile	GCC Ala	GAG Glu 285	AAG Lys	CTG Leu	AAT Asn	CAG Gln		981
	AGC Ser 290	AAC Asn	CAC His	ACC Thr	GTC Val	AGC Ser 295	AGC Ser	ATG Met	CAC His	GCC Ala	GAG Glu 300	ATG Met	CCC Pro	AAG Lys	AGC Ser	GAC Asp 305		1029
	CGC Arg	GAG Glu	CGC Arg	GTÇ Val	ATG Met 310	AAC Asn	ACC Thr	TTC Phe	CGC Arg	AGC Ser 315	GGC Gly	AGC Ser	TCC Ser	CGC Arg	GTG Val 320	CTC Leu		1077
	GTA Val	ACG Thr	ACC Thr	GAC Asp 325	CTC Leu	GTG Val	GCC Ala	CGC Arg	GGC Gly 330	ATC Ile	GAC Asp	GTG Val	CAC His	CAC His 335	GTG Val	AAC Asn	:	1125
						GAC Asp	Leu										:	1173
	CGC Arg	ATT Ile 355	GGC Gly	CGC Arg	GGC Gly	GGC Gly	CGC Arg 360	TAC Tyr	GGC Gly	GTA Val	Lys	GGT Gly 365	GTT Val	GCC Ala	ATC Ile	AAC Asn	:	1221
	TTC Phe 370	GTG Val	ACG Thr	GAG Glu	Lys	GAC Asp 375	GTG Val	GAG Glu	CTG Leu	Leu	CAC His 380	GAG Glu	ATC Ile	GAG Glu	GGG Gly	CAC His 385		1269
	TAC Tyr	CAC His	ACG Thr	Gln	ATC Ile 390	GAT Asp	GAG Glu	CTC Leu	Pro	GTG Val 395	GAC Asp	TTT Phe	GCC Ala	GCC Ala	TAC Tyr 400	CTC Leu	. 1	317
	GGC Gly	GAG Glu	TGA	GCGG	GCCC	CT G	cccc	CCTT	c cc	TGCC	cccc	TCT	CGCG	ACG			1	.366

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

(XI) Sugarred Subtrillion. Sug 15 No. 17.	
CCACTCTCTC GGTCGTCTGT CTCCCACGCG CGCACGCAGT TGATTTCCGC CTTCTTAAAC	60
GCTCTCTTTT TTTTTATTTT TCACCTGACC AACCGCACCA CGTCGGCCTC CATC ATG Met 1	117
TCG CAG CAA GAC CGA GTT GCC CCA CAG GAC CAG GAC TCG TTC CTC GAC Ser Gln Gln Asp Arg Val Ala Pro Gln Asp Gln Asp Ser Phe Leu Asp 5 10 15	165
GAC CAG CCC GGC GTC CGC CCG ATC CCG TCC TTC GAT GAC ATG CCG TTG  Asp Gln Pro Gly Val Arg Pro Ile Pro Ser Phe Asp Asp Met Pro Leu  20 25 30	213
CAC CAG AAC CTT CTG CGC GGC ATC TAC TCG TAC GGC TTC GAG AAA CCG His Gln Asn Leu Leu Arg Gly Ile Tyr Ser Tyr Gly Phe Glu Lys Pro 35 40 45	261
TCC AGC ATC CAG CAG CGC GCC ATC GCC CCC TTC ACG CGC GGC GAC Ser Ser Ile Gln Gln Arg Ala Ile Ala Pro Phe Thr Arg Gly Gly Asp 50 55 60 65	309
ATC ATC GCG CAG GCG CAG TCC GGT ACC GGC AAG ACG GGC GCC TTC TCC  Ile Ile Ala Gln Ala Gln Ser Gly Thr Gly Lys Thr Gly Ala Phe Ser  70 75 80	357
ATC GGC CTG CTG CAG CGC CTG GAC TTC CGC CAC AAC CTG ATC CAG GGC  Ile Gly Leu Leu Gln Arg Leu Asp Phe Arg His Asn Leu Ile Gln Gly  85 90 95	405
CTC GTG CTC TCC CCG ACC CGC GAG CTG GCC CTG CAG ACG GCG GAG GTG Leu Val Leu Ser Pro Thr Arg Glu Leu Ala Leu Gln Thr Ala Glu Val 100 105 110	453
ATC AGC CGC ATC GGC GAG TTC CTG TCG AAC AGC GCG AAG TTC TGT GAG  Ile Ser Arg Ile Gly Glu Phe Leu Ser Asn Ser Ala Lys Phe Cys Glu  115 120 125	501
ACC TTT GTG GGT GGC ACG CGC GTG CAG GAT GAC CTG CGC AAG CTG CAG Thr Phe Val Gly Gly Thr Arg Val Gln Asp Asp Leu Arg Lys Leu Gln 130 135 140 145	549
GCT GGC GTC GTC GCC GTG GGG ACG CCG GGC CGC GTG TCC GAC GTG Ala Gly Val Val Ala Val Gly Thr Pro Gly Arg Val Ser Asp Val 150 155 160	597
ATC AAG CGC GGC GCG CTG CGC ACC GAG TCC CTG CGC GTG CTC GTC Ile Lys Arg Gly Ala Leu Arg Thr Glu Ser Leu Arg Val Leu Val Leu  165 170 175	645
GAC GAG GCT GAT GAG ATG CTG TCT CAG GGC TTC GCG GAT CAG ATT TAC Asp Glu Ala Asp Glu Met Leu Ser Gln Gly Phe Ala Asp Gln Ile Tyr 180 185 190	693

WO 98/35045 PCT/US98/03002

84

Asp	Val	Glu	Lys	Ser	Ile	Leu	Ile	Val	Asp	Gly	Leu	Tyr	Arg	Asp	Gly
	370					375					380				

- Pro Ala Tyr Gln Thr Gly Ile Arg Leu Gly Asp Val Leu Leu Arg Ile 385 390 395 400
- Ala Gly Val Tyr Val Asp Ser Ile Ala Lys Ala Arg Gln Val Val Asp 405 410 415
- Ala Arg Cys Arg Cys Gly Cys Val Val Pro Val Thr Leu Ala Thr Lys
  420 425 430
- Met Asn Gln Gln Tyr Ser Val Ala Leu Tyr Ile Met Thr Val Asp Pro 435 440 445
- Gln His Asn Asp Lys Pro Phe Phe Phe Asp Val His Ile His His Arg 450 455 460
- Ile Glu Ser Ser His Met Gly Lys Lys Ala Gln Trp Met Glu Val Leu 465 470 475 480
- Glu Ser Pro Ser Val Ser Ser Ala Ala Thr Thr Pro Leu Val Pro Leu 485 490 495
- Leu Arg Glu Pro Thr Pro Arg Arg Gly Ser Glu Leu Gln Ser Ser Ala 500 505 510
- Arg Ser Ala Phe Val Ala Thr Ser Tyr Phe Ser Ser Ala Arg Arg Ser 515 520 525
- Val Ser Ser Glu Ser Glu Arg Pro Arg Gly Ser Ser Ser Val Ala Met 530 535 540

. . . . .

Ala Glu Glu Ala Ile Ala Leu Ala Pro Gln Gly Tyr Thr Pro Pro Asn 545 550 555 560

Gln Val Arg Gly Arg Ser 565

- (2) INFORMATION FOR SEQ ID NO:9:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 1618 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: 115..1323

Ala	Glu	Leu	Glu	Ala 85	Ala	Glu	Glu	Ala	Ala 90	Arg	Leu	Glu	Ala	Met 95	His
Glu	Ala	Glu	Gln 100	Ala	Arg	Ser	Gln	Ala 105	Leu	Glu	Glu	Ala	Ala 110	Arg	Leu
Arg	Ala	Glu 115		Glu	Glu	Ala	Glu 120	Glu	Ala	Ala	Arg	Leu 125	Asp	Val	Met
His	Ala 130	Ala	Glù	Gln	Ala	Arg 135	Val	Gln	Ala	Leu	Glu 140	Glu	Ala	Ala	Arg
Leu 145	Arg	Ala	Glu	Leu	Glu 150	Glu	Ala	Glu	Glu	Ala 155	Ala	Arg	Leu	Glu	Ala 160
Met	His	Glu	Ala	Glu 165	Gln	Ala	Arg	Ser	Gln 170	Ala	Leu	Glu	Glu	Ala 175	Ala
Arg	Leu	Arg	Ala 180	Glu	Leu	Glu	Ala	Ala 185	Glu	Glu	Ala	Ala	Arg 190	Leu	Asp
Val	Met	His 195	Glu	Ala	Glu	Gln	Ala 200	Arg	Val	Gln	Ala	Leu 205	Glu	Glu	Ala
Ala	Arg 210	Leu	Asp	Val	Met	His 215	Glu	Ala	Glu	Gln	Ala 220	Arg	Val	Gln	Ala
Leu 225	Glu	Glu	Ala	Ala	Arg 230	Leu	Arg	Ala	Glu	Leu 235	Glu	Ala	Ala	Glu	Glu 240
Ala	Ala	Arg	Leu	Asp 245	Val	Met	His	Glu	Ala 250	Glu	Gln	Ala	Arg	Val 255	Gln
Ala	Leu	Glu	Glu 260	Ala	Ala	Arg	Leu	Arg 265	Ala	Glu	Leu	Glu	Ala 270	Ala	Glu
Glu	Ala	Ala 275	Arg	Leu	Asp	Val	Met 280		Glu	Gly	Glu	Gln 285	Ala	Arg	Val
Gln	Ala 290	Leu	Glu	Glu	Ala	Ala 295	Arg	Leu	Glu	Ala	Met 300	His	Glu	Ala	Glu
Gln 305	Ala	Arg	Ser	Gln	Ala 310	Leu	Glu	Glu	Ala	Ala 315	Arg	Leu	Cys	Ala	Glu 320
Leu	Glu	Ala	Glu	Glu 325	Glu	Glu	Lys	Asp	Glu 330	Arg	Pro	Ala	Thr	Ser 335	Ser
Tyr	Ser	Glu	Glu 340	Суз	Lys	Gly	Arg	Leu 345	Leu	Ser	Arg	Ala	Arg 350	Pro	Asp
Dw-	7	B ===	Des -	<b>T</b>	~	<b>3</b>		<b>5</b> 1	<b>-</b>	~1.	34 - 2:	0	<b>.</b>	<b>.</b>	~ 7

									82						
Ile 465		Ser	Ser	His	Met 470	Gly	Lys	Lys		Gln 475	Trp	Met	Glu	Val	Leu 480
				Val		TCG Ser									
;				485					490					495	
			Pro			CGT Arg		Gly					Ser		
CCT	TCC	ccc	500	CTT	CCC	200	m cm	505	<b>E</b> EC			222	510		
Arg	Ser	Ala 515	Phe	Val	Ala	ACG Thr	Ser 520	Tyr	Phe	TCG Ser	AGC Ser	GCG Ala 525	Arg	AGG Arg	TCG Ser
GTC	AGC	TCA	GAA	AGT	GAG	CGA		CGC	GGG	TCC	TCT		GTG	GCT	ATG
Val	Ser 530	Ser	Glu	Ser	Glu	Arg 535	Pro	Arg	Gly	Ser	Ser 540	Ser	.Val	Ala	Met
GCG Ala	GAG	GAG	GCG	ATC	GCG	CTG	GCG	CCG	CAA	GGG	TAT	ACC	CCA	CCC	AAC
545		Giu			550	Leù	Ala	PIO	GIN	555	ıyr	Tnr	Pro	Pro	Asn 560
CAA Gln	GTG Val	CGC Arg	GGC Glv	CGT Arg	AGT Ser	TGAC	GTC1	CT G	STGTO	AGTG	T GT	GTCG	CTCC	2	•
		-	•	565				•						•	
GTC'	rcct'	rcc 1	TTTT	CGTC	A TO	STGTI	'TTA'I	TCA	TTTC	TTT	TTC				
(2)	INF	DRMA'I	ON	FOR	SEQ	ID N	: 8 : Oi	1	•			•			
		(i) S	EQUE	NCE	CHAR	LACTE	RTS1								
			(A)	TEN	COMT.			CICS:							
			(B)	TYP	E: a	566 mino	ami aci	.no a .d		•		• - ,	-		
			(B) (D)	TYP	E: a	mino Y: 1	ami aci inea	no a .d ir							
			(B) (D)	TYP TOP ULE	E: a	mino Y: 1	ami aci inea otei	no a d ir n	cids						
	(2	ci) S	(B) (D) OLEC	TYP TOP TULE	E: a OLOG TYPE DESC	mino Y: 1 : pr	ami aci inea otei	no a d ir n SEQ	cids	NO : 8					
Gln 1	(2	ci) S	(B) (D) OLEC	TYP TOP TULE	E: a OLOG TYPE DESC	mino Y: 1	ami aci inea otei	no a d ir n SEQ	cids	NO : 8		Leu	Arg	Ala 15	Glu
1	(a Ala	ki) S Arg	(B) (D) OLEC EQUE Val	TYP TOP SULE SNCE Gln 5	E: a OLOG  TYPE  DESC	mino Y: 1 : pr	ami aci inea otei ION:	no a d ir n SEQ	o ID Ala	NO:8 Ala	Arg			15	
1 Leu	() Ala Glu	ci) S Arg Ala	(B) (D) OLEC EQUE Val Ala 20	TYP TOP CULE CNCE Gln 5	E: a OLOG TYPE DESC Ala Glu	mino Y: 1 : pr :RIPT	ami aci inea otei ION: Glu	no and ar second are second ar second ar second are second ar second are second	ID Ala 10	NO:8 Ala Asp	Arg Val	Met	His 30	15 Ala	Ala

Ala Glu Gln Ala Arg Val Gln Ala Leu Glu Glu Ala Ala Arg Leu Arg

									91							
				245					250		-			255		
					GCG Ala											816
					GAT Asp										GTC Val	864
					GCG Ala											912
					GCC Ala 310											960
					GAG Glu											1008
					AAA Lys											1056
					CCG Pro											1104
					ATT Ile											1152
	_				GGC Gly 390											1200
					GAT Asp											1248
					GGC Gly											 1296
					AGC Ser											1344
					CCC Pro											1392
ATC	GAG	AGC	TCG	CAC	ATG	GGG	AAG	AAG	GCG	CAG	TGG	ATG	GAA	GTT	CTT	1440

														CGC Arg	GCG Ala		144
		Glu												CAT His			192
														CTC Leu			240
														ATG Met 95			288
														CGT Arg	CTC Leu		336
														GTC Val			384
CAT His	GCG Ala 130	GCC Ala	GAG Glu	CAG Gln	GCC Ala	CGC Arg 135	GTC Val	CAG Gln	GCC Ala	CTC Leu	GAG Glu 140	GAG Glu	GCA Ala	GCG Ala	CGT Arg		432
														GAG Glu		·	480
			Ala											GCA Ala 175			528
														CTG Leu			576
														GAG Glu		· .	624
														CAG Gln			672
				Ala										GAG Glu	GAG Glu 240		720
														GTC Val			768

Lys	Leu	Cys	Lys	Thr	Met	Lys	Glu	Val	Leu	Gly	Asp	Lys	Val	Glu	Lys
•		-	500			_	,	505			,		510		

Val Thr Val Ser Glu Arg Leu Leu Thr Ser Pro Cys Ile Leu Val Thr
515 520 525

Ser Glu Phe Gly Trp Ser Ala His Met Glu Gln Ile Met Arg Asn Gln 530 540

Ala Leu Arg Asp Ser Ser Met Ala Gln Tyr Met Val Ser Lys Lys Thr 545 550 555 560

Met Glu Val Asn Pro Asp His Pro Ile Ile Lys Glu Leu Arg Arg Arg 565 570 575

Val Glu Ala Asp Glu Asn Asp Lys Ala Val Lys Asp Leu Val Phe Leu
580 585 590

Leu Phe Asp Thr Ser Leu Leu Thr Ser Gly Phe Gln Leu Asp Asp Pro 595 600 605

Thr Gly Tyr Ala Glu Arg Ile Asn Arg Met Ile Lys Leu Gly Leu Ser
610 615 620

Leu Asp Glu Glu Glu Glu Val Ala Glu Ala Pro Pro Ala Glu Ala 625 630 635 640

Ala Pro Ala Glu Val Thr Ala Gly Thr Ser Ser Met Glu Gln Val Asp
645 650 655

#### (2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1771 base pairs
    - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..1698
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CAG GCC CGC GTC CAG GCC CTC GAG GAG GCA GCG CGT CTC CGC GCG GAG
Gln Ala Arg Val Gln Ala Leu Glu Glu Ala Ala Arg Leu Arg Ala Glu

1 5 10 15

96

CTG GAG GCG GCC GAG GAG GCG GCC CGC CTG GAT GTC ATG CAT GCG GCC Leu Glu Ala Ala Glu Glu Ala Ala Arg Leu Asp Val Met His Ala Ala

3.0

	•	195					200				•	205			
Glu	Val 210	Lys	Lys	Thr	Tyr	Glu 215	Val	Lys	Asn	Lys	His 220	Lys	Pro	Leu	Trp
Thr 225	Arg	Asp	Thr	Lys	Asp 230	Val	Thr	Lys		Glu 235	Tyr	Ala	Ala	Phe	Tyr 240
Lys	Ala	Ile	Ser	Asn 245	Asp	Trp	Glu	Asp	Thr 250	Ala	Ala	Thr	Lys	His 255	Phe
Ser	Val	Glu	Gly 260	Gln	Leu	Glu	Phe	Arg 265	Ala	Ile	Ala	Phe	Val 270	Pro	Lys
Arg	Ala	Pro 275	Phe	Asp	Met	Phe	Glu 280	Pro	Asn	Lys	Lys	Arg 285	Asn	Asn	Ile
Lys	Leu 290	Tyr	Val	Arg	Arg	Val 295	Phe	Ile	Met	Asp	Asn 300	Cys	Glu	Asp	Leu
Cys 305	Pro	Asp	Trp	Leu	Gly 310	Phe	Val	Lys	Gly	Val 315	Val	Asp	Ser	Glu	Asp 320
Leu	Pro	Leu	Asn	Ile 325	Ser	Arg	Glu	Asn	Leu 330	Gln	Gln	Asn	Lys	Ile 335	Leu
Lys	Val	Ile	Arg 340	Lys	Asn	Ile	Val	Lys 345	Lys	Cys	Leu	Glu	Leu 350	Phe	Glu
Glu	Ile	Ala 355	Glu	Asn	Lys	Glu	Asp 360	Tyr	Lys	Gln	Phe	Tyr 365	Glu	Gln	Phe
Gly	Lys 370	Asn	Ile	Lys	Leu	Gly 375	Ile	His	Glu	Asp	Thr 380	Ala	Asn	Arg	Lys
Lys 385	Leu	Met	Glu	Leu	Leu 390	Arg	Phe	Tyr	Ser	Thr 395	Glu	Ser	Gly	Glu	Glu 400
Met	Thr	Thr	Leu	Lys 405	qaA	Tyr	Val	Thr	Arg 410	Met	ГАè	Pro	Glu	Gln 415	Lys
Ser	Ile	Tyr	Tyr 420	Ile	Thr	Gly	Asp	Ser 425	Lys	Lys	Lys	Leu	Glu 430	Ser	Ser
Pro	Phe	Ile 435	Glu	Lys	Ala	Arg	Arg 440	Cys ,	Gly	Leu	Glu	Val 445	Leu	Phe	Met
Thr	Glu 450	Pro	Ile	Asp	Glu	Tyr 455	Val	Met	Gln	Gln	Val 460	Lys	Asp	Phe	Glu
Asp 465	Lys	Lys	Phe	Ala	Cys 470	Leu	Thr	Lys	Glu	Gly 475	Val	His	Phe	Glu	Glu 480
Ser	Glu	Glu	Glu	Lys 485	Lys	Gln	Arg	Glu	Glu 490	Lys	Lys	Ala	Ala	Cys 495	Glu

815 820 825 830

GAC TGAGCCGGTA A Asp 2040

### (2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 656 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Ser Leu Thr Asp Pro Ala Val Leu Gly Glu Glu Thr His Leu Arg Val 1 5 10 15

Arg Val Val Pro Asp Lys Ala Asn Lys Thr Leu Thr Val Glu Asp Asn 20 25 30

Gly Ile Gly Met Thr Lys Ala Asp Leu Val Asn Asn Leu Gly Thr Ile 35 40 45

Ala Arg Ser Gly Thr Lys Ala Phe Met Glu Ala Leu Glu Ala Gly Gly 50 55 60

Asp Met Ser Met Ile Gly Gln Phe Gly Val Gly Phe Tyr Ser Ala Tyr 65 70 75 80

Leu Val Ala Asp Arg Val Thr Val Val Ser Lys Asn Asn Ser Asp Glu 85 90 95

Ala Tyr Trp Glu Ser Ser Ala Gly Gly Thr Phe Thr Ile Thr Ser Val

Gln Glu Ser Asp Met Lys Arg Gly Thr Ser Thr Thr Leu His Leu Lys 115 120 125

Glu Asp Gln Glu Tyr Leu Glu Glu Arg Arg Val Lys Glu Leu Ile 130 135 140

Lys Lys His Ser Glu Phe Ile Gly Tyr Asp Ile Glu Leu Met Val Glu 145 150 155 160

Lys Thr Ala Glu Lys Glu Val Thr Asp Glu Asp Glu Glu Glu Asp Glu 165 170 175

Ser Lys Lys Ser Cys Gly Asp Glu Gly Glu Pro Lys Val Glu Glu 180 185 190

Val Thr Glu Gly Gly Glu Asp Lys Lys Lys Lys Thr Lys Lys Val Lys

												GAG Glu					1402
												GTG Val 635					1450
												GTG Val					1498
												AAG Lys					1546
Glu	Lys	Leu	Cys	Lys 675	Thr	Met	Lys	Glu	Val 680	Leu	Gly	GAC Asp	Lys	Val 685	Glu		1594
Lys	Val	Thr	Val 690	Ser	Glu	Arg	Leu	Leu 695	Thr	Ser	Pro	TGC Cys	Ile 700	Leu	Val	i.	1642
Thr	Ser	Glu 705	Phe	Gly	Trp	Ser	Ala 710	His	Met	Glu	Gln	ATC Ile 715	Met	Arg	Asn		1690
Gln	Ala 720	Leu	Arg	Asp	Ser	Ser 725	Met	Ala	Gln	Tyr	Met 730	GTG Val	Ser	Lys	Lys		1738
Thr 735	Met	Glu	Val	Asn	Pro 740	Asp	His	Pro	Ile	Ile 745	Lys	GAG Glu	Leu	Arg	Arg 750		1786
Arg	Val	Glu	Ala	Asp 755	Glu	Asn	Asp	Lys	Ala 760	Val	Lys	GAC Asp	Leu	Val 765	Phe		1834
Leu	Leu	Phe	Asp 770	Thr	Ser	Leu	Leu	Thr 775	Ser	Gly	Phe	CAG Gln	Leu 780	Asp	Asp		1882
Pro	Thr	Gly 785	Tyr	Ala	Glu	Arg	Ile 790	Asn	Arg	Met	Ile	AAG Lys 795	Leu	Gly	Leu		1930
Ser	Leu 800	Asp	Glu	Glu	Glu	Glu 805	Glu	Val	Ala	Glu	Ala 810	CCG Pro	Pro	Ala	Glu		1978
												ATG Met					2026

									13							
AAG Lys	GAG Glu	GTG Val 385	AAG Lys	AAG Lys	ACG Thr	TAC Tyr	GAG Glu 390	GTC Val	AAG Lys	AAC Asn	AAG Lys	CAC His 395	AAG Lys	CCG Pro	CTC Leu	730
	ACG Thr 400															778
	AAG Lys															826
	TCG Ser															874
	CGC															922
	AAG Lys															970
CTG Leu	TGC Cys 480	CCG Pro	GAC Asp	TGG Trp	CTC Leu	GGC Gly 485	TTC Phe	GTG Val	AAG Lys	GGC Gly	GTC Val 490	GTG Val	GAC Asp	AGC Ser	GAG Glu	1018
GAC Asp 495	CTG Leu	CCG	CTG Leu	AAC Asn	ATC Ile 500	TCG Ser	CGC Arg	GAG Glu	AAC Asn	CTG Leu 505	CAG Gln	CAG Gln	AAC Asn	AAG Lys	ATC Ile 510	 1066
CTG Leu	AAG Lys	GTG Val	ATC Ile	CGC Arg 515	AAG Lys	AAC Asn	ATC	GTG Val	AAG Lys 520	AAG Lys	TGC Cys	CTG Leu	GAG Glu	CTG Leu 525	TTC Phe	1114
GAA Glu	GAG	ATA Ile	GCG Ala 530	Glu	AAC Asn	AAG Lys	GAG Glu	GAC Asp 535	Tyr	AAG Lys	CAG Gln	TTC Phe	TAC Tyr 540	GAG Glu	CAG Gln	1162
TT( Phe	GGC Gly	AAG Lys	Asn	ATC	AAC Lys	CTG Leu	GGC Gly 550	, Ile	CAC His	GAG Glu	GAC Asp	ACG Thr	Ala	AAC Asn	CGC Arg	1210
AA( Lys	3 AAC 3 Lys 560	Lev	ATG Met	GAG Glu	TTC Lev	CTG Lev 565	Arg	TTC Phe	TAC Tyr	AGC Ser	ACC Thr	: Glu	TCG Ser	GGG Gly	GAG Glu	1258
GA: G1: 57:	u Met	G ACC	ACA Thr	CTC	5 AAC 1 Lys 580	a Ası	TAC Ty	C GTO	ACC Thr	G CGC Arg 589	g Met	AAC Lys	CCG Pro	GAC Glu	G CAG I Gln 590	1306
AA Ly	G TC	G ATO	C TAC	TAC Ty:	: Il	C AC	r GGG	y Asi	AGC Sei	r Ly	G AAC s Lys	AAC s Lys	CTC	GAC Glu 609	G TCG 1 Ser	 1354

170/3

CGC	GGTG	GCG	GCCG	CTCT.	AG A	ACTÀ	GTGG.	A TC	cccc	GGGC	TGC	AGGA	ATT	CGGC	ACGAGA		60
				sp P					ly G					TG C eu A 1		1	.06
										Thr				GAG Glu 205		1	.54
														GGC Gly			02
														GCC Ala		2	50
														TCC Ser		2	98
	Leu													TCG Ser		3	46
														ACG Thr 285		3	94
														CAÇ His		4	42
														GAG Glu		4	90
														ATG Met		5	38
														GAG Glu		5	86
			Lys											GTG Val 365		6:	34
															GTG Val	6	82.

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Val Lys Ser His Tyr Ile Cys Ala Gly Arg Leu Val Arg Ile Leu
1 5 10 15

Arg Gly Pro Arg Gln Asp Arg Val Gly Val Ile Val Asp Ile Val Asp 20 25 30

Ala Asn Arg Val Leu Val Glu Asn Pro Glu Asp Ala Lys Met Trp Arg
35 40 45

His Val Gln Asn Leu Lys Asn Val Glu Pro Leu Lys Tyr Cys Val Ser 50 55 60

Val Ser Arg Asn Cys Ser Ala Lys Ala Leu Lys Asp Ala Leu Ala Ser 65 70 75 80

Ser Lys Ala Leu Glu Lys Tyr Ala Lys Thr Arg Thr Ala Ala Arg Val 85 90 95

Glu Ala Lys Lys Ala Cys Ala Ala Ser Thr Asp Phe Glu Arg Tyr Gln
100 105 110

Leu Arg Val Ala Arg Arg Ser Arg Ala His Trp Ala Arg Lys Val Phe
115 120 125

Asp Glu Lys Asp Ala Lys Thr Pro Val Ser Trp His Lys Val Ala Leu 130 135 140

Lys Lys Met Gln Lys Lys Ala Ala Lys Met Asp Ser Thr Glu Gly Ala 145 150 155 160

Lys Arg Arg Met Gln Lys Ala Ile Ala Ala Arg Lys Ala Lys Lys 165 170 175

- (2) INFORMATION FOR SEQ ID NO:5:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 2040 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: 62..2029
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

			GTG Val												GGT Gly		100
O17	9	Deu	,u1	560	*16	пец	мg	GIY	565	Arg	GIII	Asp	Arg	570	GIÀ		
GTG	ATC	GTC	GAC	ATT	GTC	GAC	GCG	AAC	CGC	GTG	CTG	GTG	GAG	AAC	CCG		148
va1.	116	Val	Asp 575	He	Val	Asp	Ala	Asn 580	Arg	Val	Leu	Val	Glu 585	Asn	Pro		
GAG	GAC	GCG	AAG	ATG	TGG	CGC	CAC	GTG	CAG	AAC	CTG	AAG	AAC	GTG	GAG		196
Glu	Asp	Ala 590	Lys	Met	Trp	Arg	His 595	Val	Gln	Asn	Leu	Lys 600	Asn	Val	Glu		
CCG	CTG	AAG	TAC	TGC	GTG	AGC	GTC	AGC	CGC	AAC	TGC	AGC	GCG	AAG	GCG		244
Pro	Leu 605	Lys	Tyr	Cys	Val	Ser 610	Val	Ser	Arg	Asn	Сув 615	Ser	Ala	Lys	Ala		
CTG	AAG	GAT	GCG	CTG	GCC	TCG.	TCG	AAG	GCG	CTG	GAG	AAG	TAC	GCG	AAG	;	292
Leu 620	Lys	Asp	Ala	Leu	Ala 625	Ser	Ser	Lys	Ala	Leu 630	Glu	Lys	Tyr	Ala	Lys 635		
ACG	CGC	ACT	GCT	GCG	CGC	GTG	GAG	GCG	AAG	AAG	GCG	TGC	GCC	GCG	TCG	:	340
Thr	Arg	Thr	Ala	Ala 640	Arg	Val	Glu	Ala	Lys 645	Lys	Ala	Cys	Ala	Ala 650	Ser		
ACG	GAC	TTC	GAG	CGC	TAC	CAG	CTG	CGC	GTT	GCG	CGC	CGT	TCT	CGC	GCG.	:	388
Thr	Asp	Phe	Glu 655	Arg	Tyr	Gln	Leu	Arg 660	Val	Ala	Arg	Arg	Ser 665	Arg	Ala		
			CGC													4	136
nis	iip	670	Arg	гÀг	vai	Phe	Asp 675	G1u	Lys	Asp	Ala	Lys 680	Thr	Pro	Val		
			AAG													. 4	184
ser	685	HIS	Lys	Val	Ala	Leu 690	Lys	Lys	Met	Gln	Lys 695	Lys	Ala	Ala	Lys		
			ACC													9	32
Met 700	Asp	Ser	Thr		Gly 705	Ala	Lys	Arg	Arg	Met 710	Gln	Lys	Ala	Ile	Ala 715		
			GCG Ala			TAAG	GCCA	ATA C	CCTC	ACTT	C GC	TTGT	TTCG	;		5	80
TGAT	TTTT	CG 1	rggga	GTCG	G TG	GCCC	TACC	AGC	GGTC	TTT	CATT	GGCT	TA T	TTCT	'ATCCG		40
GTCI	GAAA	GA C	GTAC	AAAA:	A AA	AAAA	AAAA	AAA	AAA					•		. 6	76

# (2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 175 amino acids
  - (B) TYPE: amino acid

375

380

Ile Lys Arg Asn Pro Ala Glu His Thr Ser Tyr Ser Asn Arg Ala Ala 385 390 395 400

Ala Tyr Ile Lys Leu Gly Ala Phe Asn Asp Ala Leu Lys Asp Ala Glu 405 410 415

Lys Cys Ile Glu Leu Lys Pro Asp Phe Val Lys Gly Tyr Ala Arg Lys 420 425 430

Gly His Ala Tyr Phe Trp Thr Lys Gln Tyr Asn Arg Ala Leu Gln Ala
435
440
445

Tyr Asp Glu Gly Leu Lys Val Asp Pro Ser Asn Ala Asp Cys Lys Asp
450
455
460

Cly Arg Tyr Arg Thr Ile Met Lys Ile Gln Glu Met Ala Ser Gly Gln 465 470 475 480

Ser Ala Asp Gly Asp Glu Ala Ala Arg Arg Ala Met Asp Asp Pro Glu 485 490 495

Ile Ala Ala Ile Met Gln Asp Ser Tyr Met Gln Leu Val Leu Lys Glu
500 505 510

Met Gln Asn Asp Pro Thr Arg Ile Gln Glu Tyr Met Lys Asp Ser Gly 515 520 525

Ile Ser Ser Lys Ile Asn Lys Leu Ile Ser Ala Gly Ile Ile Arg Phe
530 535 540

Gly Gln 545

#### (2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 676 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 26..550
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

AATTCGGCAC GAGGCATTGT GCATA ATG GTC AAG TCC CAC TAC ATC TGC GCG

Met Val Lys Ser His Tyr Ile Cys Ala

550 555

- Leu His Gly Met Arg Arg Tyr Asp Asp Ala Ile Ala Ala Tyr Glu Lys
  85 90 95
- Gly Leu Lys Val Asp Pro Ser Asn Ser Gly Cys Ala Gln Gly Val Lys 100 105 110
- Asp Val Gln Val Ala Lys Ala Arg Glu Ala Arg Asp Pro Ile Ala Arg 115 120 125
- Val Phe Thr Pro Glu Ala Phe Arg Lys Ile Gln Glu Asn Pro Lys Leu 130 135 140
- Ser Leu Leu Met Leu Gln Pro Asp Tyr Val Lys Met Val Asp Thr Val 145 150 155 160
- Ile Arg Asp Pro Ser Gln Gly Arg Leu Tyr Met Glu Asp Gln Arg Phe
  165 170 175
- Ala Leu Thr Leu Met Tyr Leu Ser Gly Met Lys Ile Pro Asn Asp Gly
  180 185 190
- Asp Gly Glu Glu Glu Glu Arg Pro Ser Ala Lys Ala Ala Glu Thr Ala 195 200 205
- Lys Pro Lys Glu Glu Lys Pro Leu Thr Asp Asn Glu Lys Glu Ala Leu 210 215 220
- Ala Leu Lys Glu Glu Gly Asn Lys Leu Tyr Leu Ser Lys Lys Phe Glu 225 230 235 240
- Glu Ala Leu Thr Lys Tyr Gln Glu Ala Gln Val Lys Asp Pro Asn Asn 245 250 255
- Thr Leu Tyr Ile Leu Asn Val Ser Ala Val Tyr Phe Glu Gln Gly Asp 260 265 270
- Tyr Asp Lys Cys Ile Ala Glu Cys Glu His Gly Ile Glu His Gly Arg 275 280 285
- Glu Asn His Cys Asp Tyr Thr Ile Ile Ala Lys Leu Met Thr Arg Asn 290 295 300
- Ala Leu Cys Leu Gln Arg Gln Arg Lys Tyr Glu Ala Ala Ile Asp Leu 305 310 315 320
- Tyr Lys Arg Ala Leu Val Glu Trp Arg Asn Pro Asp Thr Leu Lys Lys 325 330 335
- Leu Thr Glu Cys Glu Lys Glu His Gln Lys Ala Val Glu Glu Ala Tyr 340 345 350
- Ile Asp Pro Glu Ile Ala Lys Gln Lys Lys Asp Glu Gly Asn Gln Tyr 355 360 365
- Phe Lys Glu Asp Lys Phe Pro Glu Ala Val Ala Ala Tyr Thr Glu Ala

TGCAACGATT	TCGTTTTATT	TACGTCTGTG	TAGCTCCTCT	ATTCAACGGT	GCGATGACGC	2408
TAACGAAGCT	GGCCTGTATT	CGGCTAAGGC	GAAGGCAAAA	GACTAGGAGG	GGGGGGGAA	2468
GGAGACGGCG	TGACCATCAC	TGCGAAGAAA	CAAGCCGAAG	AAAAGGCCCC	GAACGCCTGC	2528
ATTTCCGCGC	GCCCTCGCCC	GCCTTCCTTC	CTTCCTTCGC	TCTCTCTCTC	TCTCTCTCTC	2588
GCTATCTTCT	CAACGGAGAC	ATGAAAGGCG	TTTGTTAGGA	AAAGAGGGGG	GGGGGAAGAG	2648
TGGGACGACG	CGCTGCGTCT	TTTGGGCACT	GGTCACGTGC	GTCACCCTCT	TTTTTTATCT	2708
CTATTGGCAC	TGTCTTGTTT	CTTTTCCCTT	TCCTATCATA	CGCGTCTCGC	AAACGACTCC	2768
GCGCTGAGCA	GCCATGTGCT	GCGGCGTGGA	GGAAGTACAC	AGACATCACG	GATGCATATG	2828
TGCGCGTCCG	TGTACGCGCT	TGTATGGGGC	TTCTAACAGC	GCCTGTGTGT	GTTTGTGTGT	2888
GTGTGTGTGT	GTGTGTCTGT	GTATTTCGAG	CGTCTGTATG	CTATTCTATT	AAGCACCGAA	2948
GAAGAGACAC	ACACGACAGC	GAAGGAGATG	GTGTCGGCTT	TTCGGCTAAT	CACTCCCTTC	3008
CATAGCTTCT	CTGAAGGAGG	CTCTCTTCCA	GAGGAATAGA	CTGCAGATGG	GGTCCACGTT	3068
TATCTGAGGA	GTCAACGGAA	АААААААА	AAAAAAAA	AAAAAAAA	AAAAAAAA	3128
CTCGAG		•				3134

## (2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 546 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Asp Ala Thr Glu Leu Lys Asn Lys Gly Asn Glu Glu Phe Ser Ala 1 5 10 15

Gly Arg Tyr Val Glu Ala Val Asn Tyr Phe Ser Lys Ala Ile Gln Leu 20 25 30

Asp Glu Gln Asn Ser Val Leu Tyr Ser Asn Arg Ser Ala Cys Phe Ala 35 40 45

Ala Met Gln Lys Tyr Lys Asp Ala Leu Asp Asp Ala Asp Lys Cys Ile 50 55 60

Ser Ile Lys Pro Asn Trp Ala Lys Gly Tyr Val Arg Arg Gly Ala Ala 65 70 75 80

									68							
Phe	Lys 370	Glu	Asp	Lys	Phe	Pro 375	Glu	Ala	Val	Ala	Ala 380	Tyr	Thr	Glu	Ala	٠
		CGC Arg														1620
		ATC Ile														1668
		ATT Ile														1716
		GCT Ala 435														1764
		GAG Glu														1812
		TAT Tyr														1860
		GAT Asp														1908
		GCA Ala														1956
ATG Met	CAG Gln	AAC Asn 515	GAT Asp	CCC Pro	ACG Thr	CGC Arg	ATT Ile 520	CAG Gln	GAG Glu	TAC Tyr	ATG Met	AAG Lys 525	GAC Asp	TCC Ser	GGG Gly	2004
		TCG Ser														2052
GGT Gly 545	CAG Gln	TAGA	CTTC	TA C	GCTC	CCTC	A TO	TTTT	CCGT	GTC	TTTC	CGT	CGGC	GGGT	`AT	2108
CGTA	AAGC	AC A	ATAA	AGCA	re ce	ATTC	ACAI	GCF	CGAG	TAA	AGTG	CTGC	GC C	TCTC	AAACA	2168
CGAC	GTCG	AG G	CTGT	GGTG	C AC	ATGC	GCGT	CCI	GCAT	GAA	GGTA	.GTGA	AG A	GGAA	AGTAA	2228
GGGA	TGTI	GT I	TGTG	GGCC	T TO	GTGG	CTGC	GCA	CACA	CCT	CTTA	TCTC	CT I	CGCT	TGGTA	2288
CCTT	CTCC	CT T	TTTC	GTCT	T CA	rccc	CCTT	TCI	CTTC	TCA	CGCT	CTCC	CT G	GCGC	GGTGG	2348

										0,								
1	45					150			•		155	•				160		
											ATG Met						948	
											AAG Lys						996	
											AAG Lys						1044	
			Lys								AAC Asn						1092	
A											CTC Leu 235						1140	
											GTG Val						1188	
											TAC Tyr						1236	
											GGT Gly						1284	
				Cys	Asp	Tyr		Ile	Ile	Ala	AAG Lys	Leu	Met				1332	
7											GAG Glu 315						1380	
											CCT Pro						 1428	
											GCG Ala						 1476	
				Glu										Asn		TAC Tyr	1524	
,	TTC	AAG	GAG	GAT	AAG	TTC	ccc	GAG	GCC	GTG	GCA	GCG	TAC	ACG	GAG	GCC	1572	

# (xi) SEQUENCE DESCRIPTION: SkEQ ID NO:1:

CAAGTGTCGA AGGACAGTGT TCNCCGTGTG AGATCGCCGG CTGTGCGTGT GAAGGCGGTG	60
CCATCGGANA AACAACACCG GTGGANCCGC AGGAAACCAT CTTTCTCCGC AGGTCTCTTT	120
TTGTTGTCGA TTGAGAGTGC NCCAAACCCT GCTGGTGCCC TTCTCACATA TCATGTTTTT	180
CGTTGTGCGC TCGCTTTGCC TTTCCTCTCC TTTCCCTCTC TTCCGTGGTG CCGTGTATAC	240
TTCTGGCACC CGCTACGTCA CTTCGCTGGT TTGAACAGAA CCACTGTGAA CACCCACGGG	300
CGATCGCACA CATACACATC CCTCACTCAC ACACACAGCT ACATCTATCC TACATAAAGC	360
TGAAAAAAA GTCTACGAAC AATTTTGTTT TTACAGTGCG TTGCCGCACA TTTCTCCGTA	420
ATG GAC GCA ACT GAG CTG AAG AAC AAG GGG AAC GAA GAG TTC TCC GCC Met Asp Ala Thr Glu Leu Lys Asn Lys Gly Asn Glu Glu Phe Ser Ala  1 5 10 15	468
GGC CGC TAT GTG GAG GCG GTG AAC TAC TTC TCA AAG GCG ATC CAG TTG Gly Arg Tyr Val Glu Ala Val Asn Tyr Phe Ser Lys Ala Ile Gln Leu 20 25 30	516
GAT GAG CAG AAC AGT GTC CTC TAC AGC AAC CGC TCC GCC TGT TTT GCA Asp Glu Gln Asn Ser Val Leu Tyr Ser Asn Arg Ser Ala Cys Phe Ala 35 40 45	564
GCC ATG CAG AAA TAC AAG GAC GCG CTG GAC GAC GCC GAC AAG TGC ATC Ala Met Gln Lys Tyr Lys Asp Ala Leu Asp Asp Ala Asp Lys Cys Ile 50 55 60	612
TCG ATC AAG CCG AAT TGG GCC AAG GGC TAC GTG CGC CGA GGA GCA GCT Ser Ile Lys Pro Asn Trp Ala Lys Gly Tyr Val Arg Arg Gly Ala Ala	660
65 70 75 80	
CTC CAT GGC ATG CGC CGC TAC GAC GAT GCC ATT GCC GCG TAT GAA AAG Leu His Gly Met Arg Arg Tyr Asp Asp Ala Ile Ala Ala Tyr Glu Lys 85 90 95	708
GGG CTC AAG GTG GAC CCT TCC AAC AGC GGC TGC GCG CAG GGC GTG AAG Gly Leu Lys Val Asp Pro Ser Asn Ser Gly Cys Ala Gln Gly Val Lys 100 105 110	756
GAC GTG CAG GTA GCC AAG GCC CGC GAA GCA CGT GAC CCC ATC GCT CGC Asp Val Gln Val Ala Lys Ala Arg Glu Ala Arg Asp Pro Ile Ala Arg 115 120 125	804
GTC TTC ACC CCG GAG GCG TTC CGC AAG ATC CAA GAG AAT CCC AAG CTG Val Phe Thr Pro Glu Ala Phe Arg Lys Ile Gln Glu Asn Pro Lys Leu 130 135 140	852
TCT CTA CTT ATG CTG CAG CCG GAC TAC GTG AAG ATG GTA GAC ACC GTC	900

Ser Leu Leu Met Leu Gln Pro Asp Tyr Val Lys Met Val Asp Thr Val

#### SEQUENCE LISTING

- (1) GENERAL INFORMATION:
  - (i) APPLICANT: Corixa Corporation
- (ii) TITLE OF INVENTION: LEISHMANIA ANTIGENS FOR USEIN THE THERAPY AND DIAGNOSIS OF LEISHMANIASIS
  - (iii) NUMBER OF SEQUENCES: 87
  - (iv) CORRESPONDENCE ADDRESS:
    - (A) ADDRESSEE: SEED and BERRY LLP
    - (B) STREET: 6300 Columbia Center, 701 Fifth Avenue
    - (C) CITY: Seattle
    - (D) STATE: Washington
    - (E) COUNTRY: USA
    - (F) ZIP: 98104-7092
    - (v) COMPUTER READABLE FORM:
      - (A) MEDIUM TYPE: Floppy disk
      - (B) COMPUTER: IBM PC compatible
      - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
      - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
  - (vi) CURRENT APPLICATION DATA:
    - (A) APPLICATION NUMBER: US
    - (B) FILING DATE: 12-FEB-1998
    - (C) CLASSIFICATION:
  - (viii) ATTORNEY/AGENT INFORMATION:
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    - (B) REGISTRATION NUMBER: 31,392
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  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 3134 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: 421..2058

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# EXAMPLE 15 SYNTHESIS OF POLYPEPTIDES

Polypeptides may be synthesized on a Perkin Elmer/Applied Biosystems Division 430A peptide synthesizer using FMOC chemistry with HPTU (O-Benzotriazole-N,N,N',N'-tetramethyluronium hexafluorophosphate) activation. A Gly-Cys-Gly sequence may be attached to the amino terminus of the peptide to provide a method of conjugation, binding to an immobilized surface, or labeling of the peptide. Cleavage of the peptides from the solid support may be carried out using the following cleavage mixture: trifluoroacetic acid:ethanedithiol:thioanisole:water:phenol (40:1:2:2:3). After cleaving for 2 hours, the peptides may be precipitated in cold methyl-t-butyl-ether. The peptide pellets may then be dissolved in water containing 0.1% trifluoroacetic acid (TFA) and lyophilized prior to purification by C18 reverse phase HPLC. A gradient of 0%-60% acetonitrile (containing 0.1% TFA) in water (containing 0.1% TFA) may be used to elute the peptides. Following lyophilization of the pure fractions, the peptides may be characterized using electrospray or other types of mass spectrometry and by amino acid analysis.

From the foregoing, it will be appreciated that, although specific embodiments of the invention have been described herein for the purpose of illustration, various modifications may be made without deviating from the spirit and scope of the invention.

### EXAMPLE 15

# ISOLATION OF DNA ENCODING FOR *L. MAJOR* ANTIGENS BY CD4+ T CELL EXPRESSION CLONING

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This example illustrates the isolation of T cell antigens of L major using a direct T cell screening approach.

Leishmania-specific CD4+ T cell lines were derived from the PBMC of an individual who tested positive in a leishmania skin test but had no clinical history of disease. These T cell lines were used to screen a *L. major* amastigote cDNA expression library prepared as described in Example 1. Immunoreactive clones were isolated and sequenced as described above. The determined cDNA sequences for the 8 isolated clones referred to as 1G6-34, 1E6-44, 4A5-63, 1B11-39, 2A10-37, 4G2-83, 4H6-41, 8G3-100 are provided in SEQ ID NO: 72-79, respectively, with the corresponding predicted amino acid sequences being provided in SEQ ID NO: 80-87, respectively. The cDNA sequences provided for 1E6-44, 2A10-37, 4G2-83, 4H6-41 and 8G3-100 are believe to represent partial clones. All of these clones were shown to stimulate T cell proliferation.

20 above revealed no known homologies to the antigen 4A5-63. 1G6-34 was found to have some homology to histone H2B previously identified in *L. enrietti*. Antigens 1E6-44, 1B11-39 and 8G3-100 showed some homology to sequences previously identified in other eukaryotes, in particular *Saccharomyces cerevisae*. 2A10-37 and 4H6-41 were found to be homologous to the two previously identified proteins alpha tubulin from *L. donovani* and beta tubulin from *L. major*, respectively, and 4G2-83 was found to be homologous to elongation initiation factor 2 previously identified in *T. cruzi*.

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# EXAMPLE 14

ISOLATION AND CHARACTERIZATION OF DNA ENCODING FOR SOLUBLE ANTIGENS FROM
AN L. CHAGASI GENOMIC DNA LIBRARY

This example illustrates the preparation of five soluble *Leishmania* antigen genes from an *L. chagasi* genomic DNA library.

An L. chagasi genomic DNA expression library was prepared from L. chagasi promastigotes using the unidirectional Lambda ZAP (uni-ZAP) kit (Stratagene) according to the manufacturer's protocol. This library was screened with a high titer rabbit sera raised against L. major soluble antigens, as described above in Example 9. Five positive clones were identified. The phagemid were excised and DNA from each of the Five clones was sequenced using a Perkin Elmer/Applied Biosystems Division automated sequencer Model 373A. The DNA sequences for these antigens, referred to as LcgSP1, LcgSP3, LcgSP4, LcgSP8, and LcgSP10 are provided in SEQ ID NO:44-48, respectively, with the corresponding amino acid sequences being provided in SEQ ID NO:49-53, respectively.

Comparison of these sequences with known sequences in the gene bank as described above, revealed no known homologies to LcgSP3, LcgSP4, LcgSP8 and LcgSP10. LcgSP1 was found to be homologous to the known antigen HSP70.

Figures 30A and B illustrate the proliferative response of murine lymph nodes to recombinant LcgSP8, LcgSP10 and LcgSP3. Lymph nodes were taken BALB/c mice 17 days after infection with *L. major*. Infection occurred by footpad injection of 2 x 10<sup>6</sup> parasites/footpad. The cells were stimulated with recombinant antigen and proliferation was measured at 72 hours using <sup>3</sup>H-thymidine. Figure 30A shows the CPM, a direct measurement of mitotic activity in response to the antigens, and Figure 30B shows the stimulation index, which measures the proliferative response relative to the negative control.

In order to obtain a higher specificity for the detection of antibodies in sera from visceral leishmaniasis patients, a homologue of LmgSP9 was isolated from *L. chagasi*, one of the causative agents of visceral leishmaniasis. A total of 80,000 pfu of an amplified *L. chagasi* genomic library were screened with the entire coding region of LmgSP9 (amplified from *L. major* genomic DNA). Seven hybridizing clones were purified to homogeneity. The determined DNA sequences for two of these clones, referred to as Lc Gene A and LcGene B, are provided in SEQ ID NO: 59 and 60, respectively, with the corresponding predicted amino acid sequences being provided in SEQ ID NO: 61 and 62, respectively. The open reading frame for Lc Gene A was found to show some homology to Gene A/C, previously isolated from *L. major* (McKlean et al., *Mol. Bio. Parasitol.*, 85:221-231, 1997). The open reading frame for Lc Gene B showed some homology to Gene B of *L. major*, discussed above, and was found to contain eleven repeats of a 14 amino acid repeat unit (SEQ ID NO: 63), with each repeat being further divided into two 7 amino acid units, provided in SEQ ID NO: 64 and 65.

The diagnostic potentials of Lc Gene A and Lc Gene B were evaluated by ELISA as described above using sera from visceral leishmaniasis patients from Sudan and Brazil, and from uninfected controls. Absorbance values were compared to those obtained using LmgSP9. Much higher absorbance values were obtained with Lc Gene A and Lc Gene B than with LmgSP9, with Lc Gene B appearing to be more effective that Lc Gene A in detecting antibodies in certain cases. These results indicate that Lc Gene B is highly effective in the diagnosis of visceral leishmaniasis.

In order to assess the diagnostic potential of the repeats found within Lc Gene B, a series of 6 peptides were synthesized (SEQ ID NO: 66-71; referred to as Pep 1-6), differing in an R or H residue. An ELISA was carried out using the full-length LcGene B protein and the six peptides. The absorbance values obtained with Pep 3 were higher than those obtained with the other 5 peptides, however they were not as high as those obtained with the full length protein.

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Table 2

REACTIVITY OF LMGSP9 WITH SERA FROM LEISHMANIA PATIENTS

Patient No.	L. chagasi lysate	K39	LmgSP9
Sudanese samples:	·		7
B19	1.067	0.306	0.554
B25	1.884	3.435	0.974
B43	1.19	3.225	0.86
B47	2.405	2.892	0.375
<b>B5</b> 0	0.834	0.748	0.432
B58	0.921	0.235	0.92
B63	1.291	0.303	0.764
B70-	0.317	0.089	3.056
VL4	1.384	3.035	2.965
VL11	0.382	0.144	0.142
VL12	0.277	0.068	0.098
VL13	0.284	0.12	0.194
Brazilian samples:			
105	3.508	3.53	0.374
106	2.979	3.373	2.292
107	2.535	3.444	0.46
109	1.661	3.415	3.319
111	3.595	3.537	0.781
112	2.052	3.469	0.63
113	3.352	3.429	0.963
114	2.316	3.437	1.058
115	2.073	3.502	1.186
116	3.331	3.461	0.96
Normal Donors:	·		
129	0.157	0.104	0.08
130	0.195	0.076	0.095
131	0.254	0.134	0.086
132	0.102	0.035	0.043

succinyl-6-hydroxy-2,4-cyclohexadiene-1-carboxylic acid synthase). LmgSP9 and LmgSP19 were found to be homologous to a *L. major* hydrophilic surface protein referred to as Gene B (Flinn, H.M. et al. *Mol. Biochem. Parasit.* 65:259-270, 1994), and to ubiquitin, respectively. To the best of the inventors' knowledge, none of these antigens have been previously shown to elicit T or B cell responses.

The reactivity of recombinant LmgSP9 with sera from patients with visceral leishmaniasis, (from both Sudan and Brazil) and from normal donors was evaluated by ELISA as described above. The absorbance values were compared with those obtained using the known *Leishmania* antigen K39 described above, with *L. chagasi* lysate being employed as a positive control. Representative results of these assays are provided below in Table 2, wherein all the patients from Brazil and those from the Sudan designated as "VL" were inflicted with visceral leishmaniasis. The results demonstrated that LmgSP9 specifically detects antibody in most individuals with visceral leishmaniasis, regardless of geographical location. In several cases, the absorbance values of the antibody reactivity to LmgSP9 were comparable to that observed with K39. In addition, LmgSP9 detected several cases of leishmaniasis that were not detected using K39. These results indicate that LmgSP9 can be used to complement the reactivity of K39.

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alone. These results demonstrate that the *Leishmania* antigens of the present invention are effective in conferring protection against *Leishmania* infection.

# EXAMPLE 13

# Isolation of DNA Encoding for Soluble Antigens from an L. Major Genomic DNA Library

This example illustrates the isolation of seven soluble *Leishmania* antigen genes from an *L. major* genomic DNA library.

An L. major genomic DNA expression library was prepared from L. major promastigotes using the unidirectional Lambda ZAP (uni-ZAP) kit (Stratagene) according to the manufacturer's protocol. This library was screened with a high titer rabbit sera raised against L. major soluble antigens, as described above in Example 9. Seven positive clones were identified. The phagemid were excised and DNA from each of the seven clones was sequenced using a Perkin Elmer/Applied Biosystems Division automated sequencer Model 373A. The DNA sequences for these antigens, referred to as LmgSP1, LmgSP3, LmgSP5, LmgSP8, LmgSP9, LmgSP13, LmgSP19, are provided in SEQ ID NO:29-35, respectively, with the corresponding amino acid sequences being provided in SEQ ID NO: 36-42, respectively. LmgSP13 was found to contain a 39 amino acid repeat sequence shown in SEQ ID NO:43.

Subsequent studies resulted in the isolation of a full-length sequence for LmgSP9. The full-length DNA sequence is provided in SEQ ID NO: 54, with the corresponding predicted amino acid sequence being provided in SEQ ID NO: 55. The amino acid sequence was found to contain six 14 amino acid repeat units (SEQ ID NO: 56), with each unit being further divided into two 7 amino acid units, provided in SEQ ID NO: 57 and 58.

Comparison of the DNA and amino acid sequences for the isolated antigens as described above, revealed no significant homologies to LmgSP1, LmgSP3, and LmgSP13. LmgSP5 was found to be related to the known PSA2 family. LmgSP8 was found to bear some homology to a sequence previously identified in *E. coli* (2-

dilutions of sera from BALB/c mice previously administered either (i) saline solution; (ii) the adjuvant B. pertussis; (iii) soluble Leishmania antigens plus B. pertussis; (iv) live L. major promastigotes; or (v) soluble Leishmania antigens plus B. pertussis followed by live L. major promastigotes (as described below in Example 12). Considerably higher absorbances were seen with sera from mice infected with live L. major promastigotes and with mice infected with live L. major promastigotes following immunization with soluble Leishmania antigens plus B. pertussis, than with sera from the other three groups of mice, indicating that anti-MAPS-1A antibody titers increase following Leishmania infection.

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### EXAMPLE 12

USE OF LEISHMANIA ANTIGENS FOR VACCINATION AGAINST LEISHMANIA INFECTION

This example illustrates the effectiveness of *Leishmania* antigens in conferring protection against disease in the experimental murine leishmaniasis model system. For a discussion of the murine leishmaniasis model system see, for example, Reiner et al. Annu. Rev. Immunol., 13:151-77, 1995.

The effectiveness of (i) crude soluble *Leishmania* antigens, (ii) MAPS-1A, and (iii) a mixture of Ldp23, LbeIF4A and M15, as vaccines against *Leishmania* infection was determined as follows. BALB/c mice (5 per group) were immunized intra-peritoneally three times at biweekly intervals with either (i) 30 µg crude soluble *Leishmania* antigens, (ii)20 µg MAPS-1A or (iii) a mixture containing 10 µg each of LeIF, Ldp23 and M15, together with 100 µg of the adjuvant *C. parvum*. Two control groups were immunized with either saline or *C. parvum* alone. Two weeks after the last immunization, the mice were challenged with 2 x 10<sup>5</sup> late-log phase promastigotes of *L. major*. Infection was monitored weekly by measurement of footpad swelling. The amount of footpad swelling seen in mice immunized with either crude soluble *Leishmania* antigens, a mixture of Ldp23, LbeiF4A and M15 (Figure 28), or MAPS-1A (Figure 29) was significantly less than that seen in mice immunized with *C. parvum* 

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infection were highly responsive to MAPS-1A, whereas cells isolated 10 days post-infection were unresponsive.

## EXAMPLE 11

# IMMUNOREACTIVITY OF SOLUBLE *LEISHMANIA* ANTIGENS WITH SERA FROM *LEISHMANIA*-INFECTED PATIENTS

The reactivity of MAPS-1A with sera from uninfected individuals, from human leishmaniasis patients with cutaneous infection, from human patients with acute visceral leishmaniasis, and from *L. major*-infected BALB/c mice was determined as follows.

Assays were performed in 96-well plates coated with 200 ng antigen diluted to 50 μL in carbonate coating buffer, pH 9.6. The wells were coated overnight at 4 °C (or 2 hours at 37 °C). The plate contents were then removed and the wells were blocked for 2 hours with 200 μL of PBS/1% BSA. After the blocking step, the wells were washed five times with PBS/0.1% Tween 20<sup>TM</sup>. 50 μL sera, diluted 1:100 in PBS/0.1% Tween 20<sup>TM</sup>/0.1% BSA, was then added to each well and incubated for 30 minutes at room temperature. The plates were then washed again five times with PBS/0.1% Tween 20<sup>TM</sup>.

The enyzme conjugate (horseradish peroxidase - Protein A, Zymed, San Francisco, CA) was then diluted 1:10,000 in PBS/0.1% Tween 20<sup>TM</sup>/0.1% BSA, and 50 μL of the diluted conjugate was added to each well and incubated for 30 minutes at room temperature. Following incubation, the wells were washed five times with PBS/0.1% Tween 20<sup>TM</sup>. 100 μL of tetramethylbenzidine peroxidase (TMB) substrate (Kirkegaard and Perry Laboratories, Gaithersburg, MD) was added, undiluted, and incubated for about 15 minutes. The reaction was stopped with the addition of 100 μL of 1 N H<sub>2</sub>SO<sub>4</sub> to each well, and the plates were read at 450 nm.

As shown in Figure 26, approximately 50% of the samples from human leishmaniasis patients showed reactivities with recombinant MAPS-1A substantially above background. Figure 27 shows the reactivity of MAPS-1A with increasing

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Recombinant MAPS-1A protein having an amino-terminal HIS-Tag was prepared using a high level *E. coli* expression system and recombinant protein was purified by affinity chromatography as described in Example 1. Southern blot analysis of genomic DNA from *L. major* digested with a panel of restriction enzymes, seven other *Leishmania* species digested with PstI, and two other infectious-disease pathogens (*T. cruzi* and *T. brucei*), using the full length insert of MAPS-1A, demonstrated that MAPS-1A is present in all eight *Leishmania* species tested (Figure 23). Northern blot analysis of *L. major* promastigote and amastigote RNAs indicated that MAPS-1A is constitutively expressed.

Using oligonucleotide primers (SEQ ID NOs:27 and 28) based on the MAPS-1A cDNA sequence provided in SEQ ID NO: 23, the corresponding gene was isolated from *L. tropica* by means of PCR (using 30 cycles of the following temperature step sequence: 94 °C, 1 minute; 50 °C, 1 minute; 72 °C, 1 minute) The determined cDNA sequence for the *L. tropica* MAPS-1A protein is provided in SEQ ID NO: 25, with the corresponding amino acid sequence being provided in SEQ ID NO: 26.

The ability of recombinant MAPS-1A to stimulate cell proliferation was investigated as follows. PBMC from 3 *L. braziliensis*-infected patients having active mucosal leishmaniasis, from 4 patients post kala-azar infection (previously infected with *L. chagasi* and/or *L. donovani*) and from 3 uninfected-individuals were prepared as described above in Example 7. The ability of MAPS-1A to stimulate proliferation of these PBMC was determined as described in Example 8 above. As shown in Figure 24, significant levels of MAPS-1A specific PBMC proliferation were seen in 2 of the 7 *Leishmania* patients.

The ability of MAPS-1A to stimulate proliferation in mice lymph node cultures was determined as described in Example 8. Figure 25 shows the amount of proliferation stimulated by MAPS-1A (at 25 µg/ml, 5 µg/ml and 1 µg/ml) as compared to that stimulated by the positive control ConA and by crude L. major promastigote supernatant proteins, 20 days post-infection with L. major. Cells isolated 20 days post-

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Leishmania antigens or antigenic epitopes. Antigenic epitopes and polypeptides comprising such epitopes may be prepared by synthetic means, as described generally above and in detail in Example 15.

In certain aspects of the present invention, described in detail below, the polypeptides, antigenic epitopes and/or soluble *Leishmania* antigens may be incorporated into pharmaceutical compositions or vaccines. For clarity, the term "polypeptide" will be used when describing specific embodiments of the inventive therapeutic compositions and diagnostic methods. However, it will be clear to one of skill in the art that the antigenic epitopes of the present invention may also be employed in such compositions and methods.

Pharmaceutical compositions comprise one or more polypeptides, each of which may contain one or more of the above sequences (or variants thereof), and a physiologically acceptable carrier. Vaccines comprise one or more of the above polypeptides and a non-specific immune response enhancer, such as an adjuvant (e.g., LbeIF4A, interleukin-12 or other cytokines) or a liposome (into which the polypeptide is incorporated). Vaccines may additionally contain a delivery vehicle, such as a biodegradable microsphere (disclosed, for example, in U.S. Patent Nos. 4,897,268 and 5,075,109). Pharmaceutical compositions and vaccines within the scope of the present invention may also contain other *Leishmania* antigens, either incorporated into a combination polypeptide or present within one or more separate polypeptides.

Alternatively, a pharmaceutical composition or vaccine may contain DNA encoding one or more of the polypeptides as described above, such that the polypeptide is generated *in situ*. In such pharmaceutical compositions and vaccines, the DNA may be present within any of a variety of delivery systems known to those of ordinary skill in the art, including nucleic acid expression systems, bacteria and viral expression systems. Appropriate nucleic acid expression systems contain the necessary DNA sequences for expression in the patient (such as a suitable promoter and terminating signal). Bacterial delivery systems involve the administration of a bacterium (such as *Bacillus-Calmette-Guerrin*) that expresses an immunogenic portion of the polypeptide on its cell surface. In a preferred embodiment, the DNA may be

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introduced using a viral expression system (e.g., vaccinia or other pox virus, retrovirus, or adenovirus), which may involve the use of a non-pathogenic (defective), replication competent virus. Techniques for incorporating DNA into such expression systems are well known to those of ordinary skill in the art. The DNA may also be "naked," as described, for example, in Ulmer et al., Science 259:1745-1749 (1993) and reviewed by Cohen, Science 259:1691-1692 (1993). The uptake of naked DNA may be increased by coating the DNA onto biodegradable beads, which are efficiently transported into the cells.

While any suitable carrier known to those of ordinary skill in the art may be employed in the pharmaceutical compositions of this invention, the type of carrier will vary depending on the mode of administration. For parenteral administration, such as subcutaneous injection, the carrier preferably comprises water, saline, alcohol, a fat, a wax or a buffer. For oral administration, any of the above carriers or a solid carrier, such as mannitol, lactose, starch, magnesium stearate, sodium saccharine, talcum, cellulose, glucose, sucrose, and magnesium carbonate, may be employed. Biodegradable microspheres (e.g., polylactic galactide) may also be employed as carriers for the pharmaceutical compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Patent Nos. 4,897,268 and 5,075,109.

Any of a variety of adjuvants may be employed in the vaccines of this invention to nonspecifically enhance the immune response. Most adjuvants contain a substance designed to protect the antigen from rapid catabolism, such as aluminum hydroxide or mineral oil, and a nonspecific stimulator of immune responses, such as lipid A, *Bordella pertussis* or *Mycobacterium tuberculosis*. Suitable adjuvants are commercially available as, for example, Freund's Incomplete Adjuvant and Complete Adjuvant (Difco Laboratories, Detroit, MI), Merck Adjuvant 65 (Merck and Company, Inc., Rahway, NJ), alum, biodegradable microspheres, monophosphoryl lipid A and quil A. Preferred adjuvants include LbeIF4A, IL-12 and other cytokines such as IFN-γ or granulocyte-macrophage colony stimulating factor (GM-CSF). By virtue of its ability to induce an exclusive Th1 immune response, the use of LbeIF4A, and variants thereof, as an adjuvant in the vaccines of the present invention is particularly preferred.

In one preferred embodiment, compositions of the present invention include multiple polypeptides selected so as to provide enhanced protection against a variety of Leishmania species. Such polypeptides may be selected based on the species of origin of the native antigen or based on a high degree of conservation of amino acid sequence among different species of Leishmania. A combination of individual polypeptides may be particularly effective as a prophylactic and/or therapeutic vaccine because (1) stimulation of proliferation and/or cytokine production by individual polypeptides may be additive, (2) stimulation of proliferation and/or cytokine production by individual polypeptides may be synergistic, (3) individual polypeptides may stimulate cytokine profiles in such a way as to be complementary to each other and/or (4) individual polypeptides may be complementary to one another when certain of them are expressed more abundantly on the individual species or strain of Leishmania responsible for infection. A preferred combination contains polypeptides that comprise immunogenic portions of M15, Ldp23, Lbhsp83, Lt-1 and LbeIF4A. 15 Alternatively, or in addition, the combination may include one or more polypeptides comprising immunogenic portions of other Leishmania antigens disclosed herein, and/or soluble Leishmania antigens.

The above pharmaceutical compositions and vaccines may be used, for example, to induce protective immunity against *Leishmania* in a patient, such as a human or a dog, to prevent leishmaniasis. Appropriate doses and methods of administration for this purposes are described in detail below.

The pharmaceutical compositions and vaccines described herein may also be used to stimulate an immune response, which may be cellular and/or humoral, in a patient. For Leishmania-infected patients, the immune responses that may be generated include a preferential Th1 immune response (i.e., a response characterized by the production of the cytokines interleukin-1, interleukin-2, interleukin-12 and/or interferon- $\gamma$ , as well as tumor necrosis factor- $\alpha$ ). For uninfected patients, the immune response may be the production of interleukin-12 and/or interleukin-2, or the stimulation of gamma delta T-cells. In either category of patient, the response stimulated may include IL-12 production. Such responses may also be elicited in

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biological samples of PBMC or components thereof derived from *Leishmania*-infected or uninfected individuals. As noted above, assays for any of the above cytokines may generally be performed using methods known to those of ordinary skill in the art, such as an enzyme-linked immunosorbent assay (ELISA).

Suitable pharmaceutical compositions and vaccines for use in this aspect of the present invention are those that contain at least one polypeptide comprising an immunogenic portion of a *Leishmania* antigen disclosed herein (or a variant thereof). Preferably, the polypeptides employed in the pharmaceutical compositions and vaccines are complementary, as described above. Soluble *Leishmania* antigens, with or without additional polypeptides, may also be employed.

The pharmaceutical compositions and vaccines described herein may also be used to treat a patient afflicted with a disease responsive to IL-12 stimulation. The patient may be any warm-blooded animal, such as a human or a dog. Such diseases include infections (which may be, for example, bacterial, viral or protozoan) or diseases such as cancer. In one embodiment, the disease is leishmaniasis, and the patient may display clinical symptoms or may be asymptomatic. In general, the responsiveness of a particular disease to IL-12 stimulation may be determined by evaluating the effect of treatment with a pharmaceutical composition or vaccine of the present invention on clinical correlates of immunity. For example, if treatment results in a heightened Th1 response or the conversion of a Th2 to a Th1 profile, with accompanying clinical improvement in the treated patient, the disease is responsive to IL-12 stimulation. Polypeptide administration may be as described below, or may extend for a longer period of time, depending on the indication. Preferably, the polypeptides employed in the pharmaceutical compositions and vaccines are complementary, as described above. A particularly preferred combination contains polypeptides that comprise immunogenic portions of M15, Ldp23, Lbhsp83, Lt-1 and LbeIF4A, Lmsp1a, Lmsp9a, and MAPS-Soluble Leishmania antigens, with or without additional polypeptides, may also be employed.

Routes and frequency of administration, as well as dosage, for the above aspects of the present invention will vary from individual to individual and may parallel

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those currently being used in immunization against other infections, including protozoan, viral and bacterial infections. In general, the pharmaceutical compositions and vaccines may be administered by injection (e.g., intracutaneous, intramuscular, intravenous or subcutaneous), intranasally (e.g., by aspiration) or orally. Between 1 and 12 doses may be administered over a 1 year period. For therapeutic vaccination (i.e., treatment of an infected individual), 12 doses are preferably administered, at one month For prophylactic use, 3 doses are preferably administered, at 3 month intervals. intervals. In either case, booster vaccinations may be given periodically thereafter. Alternate protocols may be appropriate for individual patients. A suitable dose is an amount of polypeptide or DNA that, when administered as described above, is capable of raising an immune response in an immunized patient sufficient to protect the patient from leishmaniasis for at least 1-2 years. In general, the amount of polypeptide present in a dose (or produced in situ by the DNA in a dose) ranges from about 100 ng to about 1mg per kg of host, typically from about 10 μg to about 100 μg. Suitable dose sizes will vary with the size of the patient, but will typically range from about 0.1 mL to about 5 mL.

In another aspect, this invention provides methods for using one or more of the polypeptides described above to diagnose *Leishmania* infection in a patient using a skin test. As used herein, a "skin test" is any assay performed directly on a patient in which a delayed-type hypersensitivity (DTH) reaction (such as induration and accompanying redness) is measured following intradermal injection of one or more polypeptides as described above. Such injection may be achieved using any suitable device sufficient to contact the polypeptide or polypeptides with dermal cells of the patient, such as a tuberculin syringe or 1 mL syringe. Preferably, the reaction is measured at least 48 hours after injection, more preferably 72 hours after injection.

The DTH reaction is a cell-mediated immune response, which is greater in patients that have been exposed previously to a test antigen (i.e., an immunogenic portion of a polypeptide employed, or a variant thereof). The response may measured visually, using a ruler. In general, induration that is greater than about 0.5 cm in diameter, preferably greater than about 1.0 cm in diameter, is a positive response,

indicative of *Leishmania* infection, which may or may not be manifested as an active disease.

The polypeptides of this invention are preferably formulated, for use in a skin test, as pharmaceutical compositions containing at least one polypeptide and a physiologically acceptable carrier, as described above. Such compositions typically contain one or more of the above polypeptides in an amount ranging from about 1 µg to 100 µg, preferably from about 10 µg to 50 µg in a volume of 0.1 mL. Preferably, the carrier employed in such pharmaceutical compositions is a saline solution with appropriate preservatives, such as phenol and/or Tween  $80^{TM}$ .

The inventive polypeptides may also be employed in combination with one or more known *Leishmania* antigens in the diagnosis of leishmaniasis, using, for example, the skin test described above. Preferably, individual polypeptides are chosen in such a way as to be complementary to each other. Examples of known *Leishmania* antigens which may be usefully employed in conjunction with the inventive polypeptides include K39 (Burns et al., <u>Proc. Natl. Acad. Sci. USA</u>, 1993 <u>90</u>:775-779).

The following Examples are offered by way of illustration and not by way of limitation.

#### **EXAMPLES**

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#### EXAMPLE 1

### PREPARATION OF M15

This Example illustrates the preparation of a *Leishmania* antigen M15, having the sequence provided in SEQ ID NO:2.

An L. major (Friedlan strain) amastigate cDNA expression library prepared in the  $\lambda$ ZAP II vector (Stratagene, La Jolla, CA) was screened according to manufacturer's instructions using sera obtained from L. major infected BALB/c mice (8 weeks post inoculation). Approximately 40,000 plaques were screened and four clones expressing reactive antigens were purified to homogeneity by two subsequent rounds of low density screening. Bluescript phagemid inserts were excised from positive clones

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for further analysis. An *EcoRI/SstII* restriction fragment from the 5' end of one partial cDNA insert isolated during first round screening (pLma1-1) was subsequently used as a probe to rescreen for clones containing full length cDNA inserts. The probe was labeled to high specific activity ( 10° cpm/μg) with [ -<sup>32</sup>P]dCTP using the random primer method and was used to screen 10,000 plaques of the *L. major* expression library described above. Positive clones were compared by restriction enzyme digestion and the clone with the largest insert (pf11-1) was chosen for subsequent analysis.

DNA sequence analyses were performed on an Applied Biosystems automated sequencer using Taq polymerase and dye coupled ddNTP terminators or dyelabeled sequencing primers. The complete sequence of the 2685 bp insert was determined using a combination of primer-directed sequencing and by sequencing a series of overlapping Exonuclease III deletion subclones generated using the Erase-abase system (Promega, Madison, WI). The sequence of this insert is provided in SEQ ID NO:1, and the deduced amino acid sequence is provided in SEQ ID NO:2.

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The complete insert of clone pfl-1 was excised by digestion with BamHI/KpnI and was subcloned in frame into BamHI/KpnI digested pQE31 (QUIAGEN) to generate the construct pM151A. E. coli containing this construct inducibly expressed high levels of the L. major antigen encoded by pfl1-1 (designated as M15) with the addition of a 6-histidine tag at the amino terminus. Large volume cultures (500 ml) of E. coli host cells containing the pM151A construct were induced to express recombinant protein by the addition of 2mM IPTG at mid-log phase of growth. Growth was continued for 4 to 5 hours and bacteria were then pelleted and washed once with cold PBS. Bacteria were resuspended in 20 ml of lysis buffer (50 mM Na<sub>2</sub>HPO<sub>4</sub>, pH 8.0, 300 mM NaCl, 10 mM  $\beta$ -mercaptoethanol) containing 20 mg of lysozyme and were lysed by a 1 hour incubation at 4°C followed by brief sonication. Insoluble material was removed by centrifugation at 10,000xg for 10 minutes and although the recombinant protein was found to be evenly distributed between the soluble and insoluble fractions the insoluble material was discarded at this point. Recombinant protein containing the amino terminal histidine tag was affinity purified using Ni-NTA resin (QIAGEN) according to the manufacturer's recommendations. Briefly, 8 ml of Ni-NTA resin resuspended in lysis buffer was added to the soluble lysate fraction and binding was conducted with constant mixing for 1 hour at 4°C. The mixture was then loaded into a gravity flow column and the non-binding material was allowed to flow through. The Ni-NTA matrix was washed 3 times with 25 ml of wash buffer (50 mM Na<sub>2</sub>HPO<sub>4</sub>, pH 6.0, 300 mM NaCl, 10 mM β-mercaptoethanol) and bound material was eluted in 25 ml of elution buffer (50 mM Na<sub>2</sub>HPO<sub>4</sub>, pH 5.0, 300 mM NaCl, 10mM β-mercaptoethanol). The eluted material was then dialyzed against 3 changes of PBS, sterile filtered and stored at -20°C. The purified recombinant protein was shown by SDS-PAGE analysis to be free of any significant amount of E. coli protein. A small number of bands of lower molecular weight were assumed to be proteolytic products of the L. major antigen based on their reactivity by western blot analysis. A high titre polyclonal antisera against M15 was generated in rabbits by repeated subcutaneous injection of recombinant protein. Western blot analysis of lysates from L. major promastigotes and amastigotes using this antisera indicated that the protein is constitutively expressed throughout the parasite lifecycle.

# **EXAMPLE 2**

#### PREPARATION OF LDP23

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This Example illustrates the preparation of a *Leishmania* antigen Ldp23, having the sequence provided in SEQ ID NO:4.

# A. Purification of MHC Class II-associated Peptides from P388D1 Macrophages Infected with L. donovani

To ascertain that *in vitro* infection of macrophages would load their MHC class II molecules with parasite peptides, initial experiments were carried out to test the ability of *L. donovani*-infected macrophage cell line P388D1 to present parasite antigens to *L. donovani* specific T-cells. This macrophage cell line was chosen because it has the same H-2 haplotype as the BALB/c mouse, which is a strain of mouse

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moderately susceptible to *L. donovani* infection and selected to conduct the *in vivo* experiments. Using a proportion of 3-5 parasites per cell and an initial incubation at room temperature for 4-6 hours follows by 37°C for 24-48 hours, close to 90% of the macrophages were infected. The level of MHC class II molecule expression, as determined by FACS analysis, indicated that infection did not cause an effect on the levels of MHC class II expression when compared to non-infected control cells.

To test the ability of the *L. donovani*-infected P388D1 cells to present parasite antigens, macrophages were infected as indicated above and incubated at 26°C for 6 hours, and then as 37°C for either 24, 48 or 72 hours. At each of these time points the non-adherent cells and free parasites were washed out and the adherent cells were mechanically dislodged, washed and fixed with paraformaldehyde. These cells were then used as antigen presenting cells (APCs) for purified lymph node T-cells from BALB/c mice immunized with *L. donovani* promastigotes. To generate these anti-*L. donovani* specific T-cells, BALB/c mice (H-2<sup>d</sup>) of both sexes (The Jackson Laboratory, Bar Harbor, ME) were immunized at 8 to 14 weeks of age in the rear foot pad with 5-10 x 10<sup>6</sup> *L. donovani* promastigotes emulsified in complete Freünd's adjuvant (CFA) (Difco Laboratories, Madison, MI) as described in Rodrigues et al., *Parasite Immunol.* 14:49 (1992). The draining lymph nodes were excised 8 days after the immunization and T-cells were purified in an anti-mouse Ig column to remove the B cells, as described in Bunn-Moreno and Campos-Neto, J. *Immunol.* 127:427 (1981), followed by a passage through a Sephadex G10 column to remove the macrophages.

Stimulation index was calculated by dividing the cpm obtained for the cells cultured in the presence of infected P388D1 macrophages by the cpm obtained for the cells cultured in the presence of non-infected macrophages, but subjected to the same conditions as the infected macrophages. The results shown Figure 1 indicate that *L. donovani*-infected P388D1 macrophage process parasite antigens and that optimal presentation occurs after 48 hours of infection. No stimulation of the T-cells by the non-infected macrophages was observed.

To isolate the MHC class II associated L. donovani peptides, P388D1 macrophages were infected with L. donovani promastigotes for an initial incubation of 6

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hours at room temperature. The cultures were then transferred to 37°C for the remainder of the 48 hour incubation period. At a ratio of 3-5 parasites per macrophage nearly 90% of the macrophages were infected after 24 hours of incubation at 37°C.

The MHC class II molecules were then affinity-purified. Approximately 1.5 x 10<sup>10</sup> L. donovani-infected or an equal number of non-infected P388D1 macrophages were used for each purification. The cells were harvested, washed with PBS and incubated for 30 minutes in cold lysis buffer (PBS, 1% Nonidet P40, 25mM iodoacctamide, 0.04% sodium azide, 1mM aprotinin and 1mM PMSF). The insoluble material was removed by centrifugation at 40,000g for 1 hour and the supernatant was recycled overnight at 4°C over a 5ml anti-MHC class II molecules (H-2<sup>d</sup>) Sepharose column (Protein G Sepharose column to which the monoclonal antibody MK-D6 has been bound). Culture supernatants of MK-D6 hybridoma cells (American Type Culture Collection, Rockville, MD) were employed as the source for anti-MHC class II (H-2<sup>d</sup>) monoclonal antibody. The column was washed with 50ml of lysis buffer and then with 50ml of PBS containing 0.5% octyl glucopyranoside detergent. Bound molecules were eluted from the column with 1M acetic acid in 0.2% NaCl. The MHC/peptide molecules were separated from the IgG (MK-D6 monoclonal antibody) using a Centricon 100 filter unit (Amicon Division, W.R. Grace & Co., Beverly, MA). The peptides were then dissociated from the class II molecules by the addition of acetic acid to 2.5M, followed by separation using a Centricon 10 filter unit. The resulting peptide preparation, present in the low molecular weight sample, was then dried using a speed vac concentrator (Savant Instrument Inc., Farmingdale, NY).

The peptides were redissolved in 200µl of 0.05% TFA and separated by reverse-phase high performance liquid chromatography (RP-HPLC) using a 2.1mm x 25cm Vydac C-18 column at a flow rate of 0.15ml/min employing a 1 to 30% acetonitrile gradient (60 min) followed by a 30 to 60% gradient (30 min) and then a 60 to 80% gradient (90-110 min). Non-infected P388D1 cells were similarly processed to serve as background control for endogenous MHC class II associated peptides. Figure 2 shows a representative experiment; four distinct peaks which are present only in the

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material isolated from infected macrophages (panel B), and not in the material isolated from uninfected macrophages (panel A) are indicated.

Out of three independent peptide extractions, twenty five distinct HPLC peptide peaks were isolated from L. donovani-infected macrophages and were subjected to protein sequence analysis using automated Edman degradation on an Applied Biosystems 477 gas-phase protein sequencer. Protein sequence and amino acid analysis were performed by the W.M. Keck Foundation, Biotechnology Resource Laboratory, Yale University, New Haven, CT. In practically all determinations, no assignment could be made for the first position. Also, in most cases the definition of the amino acid residues of the 10-15 positions was based on the quantitative dominance of one residue over others. Using this approach, the sequences obtained for several peptides showed the presence of 3-6 different residues in many of the 10-15 sequence cycles analyzed for each determination, reflecting a mixture of peptides. In addition, sequences could not be obtained for some peaks because the peptides were blocked. Notwithstanding, three peptides sequences were determined. Amino-acid sequences were searched for identity with proteins in the GenBank database using the GENPETP, PIR and SWISSPROT programs. The sequence data base analysis revealed that one of the peptides was highly homologous to glyceraldehyde-3-phosphate dehydrogenase of various species. Another 🗻 peptide had homology with elongation factor of several species, including Leishmania.

The third sequence was not clearly related to any known proteins, and is shown below:

XQXPQ(L/K)VFDEXX (SEQ ID NO:11).

# B. Cloning and Sequencing of the Ldp23 Gene

In order to retrieve the *L. donovani* protein that was processed into a peptide associated with the MHC class II molecules of infected macrophages, the peptide sequence of uncertain origin was chosen to guide the strategy for cloning the corresponding parasite gene. A DNA fragment was initially amplified from *L. donovani* promastigote cDNA by PCR. The sense primer was a peptide derived oligonucleotide (5' > GGAATTCCCCInCAGCTInGTInTTCGAC < 3') (SEQ ID NO:12) containing an *Eco*RI restriction endonuclease site (underlined). The bases were

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selected following the preferential codon usage of *L. donovani*, as described in Langford et al., *Exp. Parasitol.* 74:360 (1992). Inosine was used for the residues of positions 4, 6 and 7 because of the low codon usage assurance for the corresponding amino acids. In addition, the carboxyl-terminal L-glutamic acid was not included for the design of the primer. The antisense primer was a poly-thymidine oligonucleotide (oligo dT, downstream primer) containing a *XhoI* restriction endonuclease site.

The gene fragment was amplified from a *L. donovani* promastigote cDNA preparation using the following reaction conditions: one cycle of 3 min at 94°C immediately followed by 35 cycles of 1 min at 94°C, 1 min at 45°C and 1 min at 72°C. The *L. donovani* cDNA was prepared from 5 x 10<sup>7</sup> washed promastigote forms harvested at the log growth phase (3 days culture). The cDNA was obtained using an Invitrogen cDNA cycle<sup>TM</sup> kit (Invitrogen Co., San Diego, CA). Oligonucleotide primers were synthesized by the DNA Synthesis Laboratory, Department of Pathology, Yale University School of Medicine.

The PCR products were analyzed by gel electrophoresis. Only one band of approximately 300 bp was obtained. This fragment was cloned and its sequence confirmed the sequence of the peptide-based primer including the glutamic acid codon, deliberately not included in the primer sequence.

The PCR amplified gene fragment was ligated into the pCR<sup>TM</sup> vector using the TA cloning system (Invitrogen Co., San Diego, CA). Transformants were selected in LB medium containing 100µg/ml ampicillin and the plasmid DNA was isolated using the Wizard<sup>TM</sup> Minipreps DNA purification kit (Promega Co., Madison, WI). Insert DNA was released with the restriction enzymes *Eco*RI and *Xho*I (New England Biolabs, Beverly, MA), purified from an agarose gel electrophoresis and labeled with <sup>32</sup>P using a random priming method (Megaprime Labeling Kit, Amersham Life Science, Buckinghamshire, England).

This DNA fragment was used as probe to screen a *L. donovani* promastigote cDNA library as described in Skeiky et al., *Infect. Immun.* 62:1643 (1994). An approximately 650 bp cDNA (Ldp23) was excised from the phagemid by *in vivo* excision using the Stratagene protocol. DNA sequencing was performed using the

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Sequenase version 2 system (DNA sequencing kit) in the presence or absence of 7deaza-GTP (United States Biochemical, Cleveland, OH). The sequence is provided as SEQ ID NO:3, and shows complete homology with the original 300 bp PCR fragment. A 525 bp open reading frame containing an ATG codon that follows the last 4 bases of the spliced leader sequence and 3 stop codons adjacent to the poly A tail was identified. This frame also codes the carboxyl terminal sequence (KVFDE) (SEQ ID NO:13) of the purified MHC class II associated peptide. The sequence analysis of the deduced protein sequence revealed one potential glycosylation site (Asn-Cys-Ser) at positions 68-70.

Sequence analysis was performed using the University of Wisconsin Genetics Computer Group Programs and the GenBank and EMBL data bases of protein and DNA sequences. The search for homology of the Ldp23 gene with known sequences revealed no significant homology.

#### C. Bacterial Expression and Purification of Recombinant Protein

The recombinant L. donovani peptide donor protein was produced in 15 E. coli transformed with the pGEX 2T expression vector in which the Ldp23 gene was subcloned in frame. PCR was used to subclone the cloned gene in frame into the expression vector pGEX 2T. Primers containing the appropriate restriction site enzymes, initiation and termination codons were: 20 GGATCCATGGTCAAGTCCCACTACATCTGC <3' (SEQ ID NO:14) for the upstream primer and 5' > GAATTCAGACCGGATAGAAATAAGCCAATGAAA <3' (SEQ ID NO:15) for the downstream primer (restriction sites of BamHI and EcoRI are underlined respectively). PCR conditions were as indicated above for the amplification of the original peptide related DNA fragment. The template used was pBluescript plasmid containing the cloned gene from the cDNA library.

Overexpression of the recombinant fusion protein was accomplished by growing the transformed E. coli (DH5a) and inducing the tac promoter with 1mM isopropyl-β-thiogalactopyranoside (IPTG) (Stratagene, La Jolla, CA). Cells were collected, centrifuged, and analyzed for the presence of the fusion protein by SDS-A glutathione-S-transferase fusion protein of 43-44 kD was produced, PAGE.

indicating a leishmanial protein of approximately 18 kD, as glutathione-S-transferase (GST) has a MW of 26 kD. However, the fusion protein was very insoluble and therefore could not be purified by affinity chromatography using a glutathione column. The use of low concentrations of detergents like SDS, sarcosyl, deoxycolate, and octylglucopyranoside during the extraction steps was efficient to solubilize the protein but unfortunately prevented its binding to the glutathione column. Other maneuvers, such as the growth of the *E. coli* and incubation and induction of the *tac* promoter with IPTG at 33°C, did not improve the protein solubility. However, the purification was achieved by preparative SDS-PAGE. The band was visualized with 0.1M KCl, cut and electroeluted from the gel followed by extensive dialysis against PBS and concentration on Centricon 10 filters.

Approximately 500µg of purified protein was obtained. The purified protein is shown in Figure 3. In panel A, E. coli (DH5α) transformed with the expression vector pGEX 2T containing the Ldp23 gene was grown in LB medium and the tac promoter was induced with IPTG for 3 hours. The cells were pelleted, resuspended in loading buffer and submitted to SDS-PAGE (10%) under reducing condition. The gel was stained with Coomassie blue. Lane 1 shows the uninduced E. coli and land 2 shows the induced E. coli. The arrow indicates the recombinant protein. Panel B shows the protein prepared as in panel A and submitted to a preparative SDS-PAGE. The band corresponding to the overexpressed recombinant fusion protein was identified by KCl, cut out, electroeluted from the gel strip, dialyzed against PBS and submitted to analytical SDS-PAGE (12%). Numbers on the left side indicate the molecular weights of the markers. Attempts to further purify the leishmanial protein by cleaving it out from the fusion protein GST with thrombin were unsuccessful.

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# D. Expression of Ldp23

To ascertain that the Ldp23 peptide is expressed in *Leishmania* organisms, a Northern blot analysis was performed using RNA prepared from different promastigote growth phases (logarithmic and stationary) and from the amastigote form of these parasites.

The RNA was prepared from 2 x 10<sup>7</sup> parasite cells using the Micro RNA isolation kit (Stratagene, La Jolla, CA) according to the company's recommended instructions. RNA was prepared from *L. donovani* promastigotes (logarithmic growth phase); from *L. major* promastigotes (logarithmic and stationary growth phases); from *L. amazonensis*, both promastigotes (logarithmic and stationary growth phases) and amastigotes purified from CBA/J infected mice; and from *L. pifanoi*, both promastigotes (logarithmic and stationary growth phases) and amastigotes (from axenic culture medium). *L. donovani* (1S strain), *L. amazonensis* (MHOM/BR/77/LTB0016), *L. major* (MHOM/IR/79/LRC-L251) and *L. pifanoi* (MHOM/VE/60/Ltrod) promastigotes were grown and maintained at 26°C in Schneider's medium containing 20% FCS and 50µg/ml gentamicin. The amastigote forms of *L. amazonensis* were obtained by differential centrifugation of a "pus-like" foot pad lesion of a CBA/J mouse infected for 6 months with this parasite. *L. pifanoi* amastigotes were obtained from axenic culture as previously reported by Pan et al., *J. Euk. Microbiol. 40*:213 (1993).

The hybridization was carried out at 45°C in the presence of 50% formamide, 5x Denhardt's solution, 0.1% SDS, 100µg/ml single stranded salmon sperm DNA and 5x SSPE using 0.45µm Nytran membrane filters (Schleicher & Schuell, Keene, NH). The probe was the <sup>32</sup>P labeled Ldp23 gene.

Figure 4 shows that one single RNA band of 680 bp was observed for all growth phases and forms of all tested *Leishmania*. Within Figure 4, the numbers 1, 2 and 3 refer to RNA obtained from promastigotes at the logarithmic growth phase, promastigotes at the stationary growth phase and amastigote forms, respectively, and the numbers on the left side indicate the molecular weights of the markers in base pairs. This result is consistent with the corresponding gene size (525 bp) and with the molecular weight of the expressed protein and points to the ubiquitous distribution and expression of this gene within the genus *Leishmania*.

E. <u>Induction of Anti-L. donovani</u> Antibody Response in Mice and Rabbits by Purified Recombinant Protein

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In order to evaluate the immunogenicity of the recombinant leishmanial protein, and to investigate its expression in the parasites, mice and rabbits were immunized with the GST-fusion protein in CFA. BALB/c mice were immunized in the rear foot pad with 5-10µg of protein emulsified in CFA. Protein concentration was determined using the Bio-Rad Protein Assay reagent (Bio-Rad Laboratories, Richmond, CA). The mice were boosted 7 days later with 5-10µg of protein emulsified in incomplete Fretind's adjuvant (IFA) inoculated into the peritoneal cavity. The mice were bled 7 days after the second immunization. New Zealand white rabbits (Millbrook Farm, Amherst, MA) were immunized according to the following protocol: one intramuscular (IM) injection of 25-30µg of purified recombinant protein emulsified in CFA into each thigh on day one; one IM injection of 25-30µg of purified protein emulsified in IFA into each shoulder on day 7; on day 15, 25-30µg of the purified protein in PBS was injected into the subcutaneous tissue. The rabbit was bled 7 days after the last immunization.

Sera were prepared and the anti-Leishmania antibody response was measured by Western blot analysis and by FACScan. In both cases L. donovani promastigotes were used as antigen. Approximately 2 x 106 L. donovani promastigotes were grown in Schneider's medium for 3 days (log phase), were washed with PBS, lysed with SDS-PAGE loading buffer and submitted to electrophoresis under reducing conditions using a 15% polyacrylamide gel. The proteins were transferred onto 0.45  $\mu$ Immobilon-P transfer membrane (Millipore Co., Bedford, MA) using a wet-type electroblotter (Mini Trans-Blot Electrophoretic Transfer Cell, Bio Rad Life Science Division, Richmond, CA) for 2 hours at 50 V. The membranes were blocked overnight at room temperature with PBS containing 3% normal goat serum (NGS), 0.2% Tween-20 and 0.05% sodium azide, followed by 3 washes with PBS. The blots were then incubated for 3-4 hours at 4°C with a 1/200 dilution of pre-immune rabbit serum (lane A, Figure 5) or with the same dilution of anti-fusion protein rabbit antiserum (lane B, The sera was previously absorbed 2x with non-viable desiccated Figure 5). Mycobacterium tuberculosis H-37 RA (Difco Laboratories, Detroit, MI) and were diluted in PBS containing 1% NGS and 5% powdered non-fat bovine milk (Carnation,

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Nestlé Food Company, Glendale, CA). The membranes were then washed with PBS, incubated for 1 hour at room temperature with goat anti-rabbit IgG antibody conjugated with alkaline phosphatase (Promega, Madison, WI), washed once with PBS and 2x with veronal buffer pH 9.4. The reaction was visualized using the substrate mixture 5-bromo-4-chloro-3-indoyl-phosphate and nitroblue tetrazolium (Kirkegaard & Perry Laboratories Inc., Gaithersburg, MD) according to the manufacturer's instructions.

Figure 5 shows that the rabbit anti-recombinant protein antiserum detects a single protein of 23 kDa (Ldp23) in the *Leishmania* crude extract antigen preparation. No bands were observed when an anti-GST antiserum was used (not shown). Moreover, the FACScan analysis (Figure 6) shows that the antibody induced by the recombinant Ldp23 reacts with intact live *L. donovani* promastigotes, thus pointing to a cell surface expression of this molecule on these organisms. The dotted line in Figure 6 shows the indirect immunofluorescence performed using pre-immune mouse serum and the solid line in Figure 6 shows the result obtained with mouse anti-GST-Ldp23 antiserum. Both sera were diluted at 1/100. Parasites were washed with staining buffer and incubated with FITC conjugated goat anti-mouse immunoglobulin antibody. Fluorescence intensity was analyzed by FACScan.

# F. Recognition of Recombinant Ldp23 by Leishmania-Specific Lymph Node Tcells

To test the responsiveness of T-cells to the Ldp23 protein, two sets of experiments were performed. In the first experiment, lymph node T-cells (10<sup>5</sup>/well) from BALB/c mice immunized with *L. donovani* promastigotes (as described above) were stimulated to proliferate with 2 x 10<sup>5</sup> Mitomycin C-treated normal mononuclear spleen cells (APC) and pulsed with the purified recombinant fusion protein. Proliferation of T-cells was measured at 72 hours of culture. Values are expressed in Figure 7 as cpm and represent the mean of [<sup>3</sup>H]TdR incorporation of triplicate cultures. Background cpm of cells (T cells + APC) cultured in the presence of medium alone was 1291. Figure 7 shows that *Leishmania* specific T-cells proliferate well and in a dose response manner to recombinant Ldp23. No response was observed when purified GST

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was added instead of the recombinant fusion protein nor when lymph node T-cells from mice immunized with CFA alone were stimulated to proliferate in the presence of the Leishmanial fusion protein (not shown).

The recognition of the recombinant Ldp23 protein by Leishmania-specific T-cells was also tested using two murine models of leishmaniasis, the L. major highly susceptible BALB/c mice and the L. amazonensis susceptible CBA/J mice as described in Champsi and McMahon-Pratt, Infect. Immun. 56:3272 (1988). These models were selected to investigate the cytokine pattern induced by Ldp23. In the mouse model of leishmaniasis, resistance is associated with Th 1 cytokines while susceptibility is linked to Th 2 responses.

Lymph node cells were obtained 3 weeks after the initiation of infection of BALB/c mice with *L. major* and the ability of these cells to recognize the recombinant Ldp23 was measured by proliferation and by the production of the cytokines IFN-γ and IL-4. 2 x 10<sup>6</sup> cells obtained from the draining popliteal lymph node of infected mice were cultured for 72 hours in the presence of recombinant Ldp23 or *Leishmania* lysate. The levels of IFN-γ and IL-4 in culture supernatants were measured by ELISA as previously described (Chatelain et al., *J. Immunol. 148*:1172 (1992), Curry et al., *J. Immunol. Meth. 104*:137 (1987), and Mossman and Fong, *J. Immunol. Meth. 116*:151 (1989)) using specific anti IFN-γ and IL-4 monoclonal antibodies (PharMingen, San Diego, CA).

Ldp23 did stimulate these cells to proliferate (not shown) and induced a typical Th 1 type of cytokine response as indicated by the production of high levels of IFN-γ (panel A of Figure 8) and no IL-4 (panel B of Figure 8). Stimulation of these cells with a *Leishmania* crude lysate yielded a mixed Th cytokine profile. Exactly the same pattern of cytokine production was obtained from the CBA/J mice infected with *L. amazonensis* (not shown). These results clearly indicate that Ldp23 is a powerful and selective activator of the Th 1 cytokines by mouse cells.

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## **EXAMPLE 3**

#### PREPARATION OF HSP83

This Example illustrates the preparation of a *Leishmania* antigen Hsp83, having the sequence provided in SEQ ID NO:6.

A genomic expression library was constructed with sheared DNA from *I.. braziliensis* (MHOM/BR/75/M2903) in bacteriophage λZAP II (Stratagene, La Jolla, CA). The expression library was screened with *Escherichia coli* preadsorbed serum from an *L. braziliensis*-infected individual with ML. Immunoreactive plaques were purified, and the pBSK(-) phagemid was excised by protocols suggested by the manufacturer. Nested deletions were performed with exonuclease III to generate overlapping deletions for single-stranded template preparations and sequencing. Single-stranded templates were isolated following infection with VCSM13 helper phage as recommended by the manufacturer (Stratagene, La Jolla, CA) and sequenced by the dideoxy chain terminator method or by the *Taq* dye terminator system using the Applied Biosystems automated sequencer model 373A.

Recombinant antigens produced by these clones were purified from 500 ml of isopropyl-β-D-thiogalactopyranoside (IPTG)-induced cultures as described in Skeiky et al., *J. Exp. Med. 176*:201-211 (1992). These antigens were then assayed for the ability to stimulate PBMC from *Leishmania*-infected individuals to proliferate and secrete cytokine. Peripheral blood was obtained from individuals living in an area (Corte de Pedra, Bahia, Brazil) where *L. braziliensis* is endemic and where epidemiological, clinical, and immunological studies have been performed for over a decade, and PBMC were isolated from whole blood by density centrifugation through Ficoll (Winthrop Laboratories, New York, N.Y.). For *in vitro* proliferation assays, 2 X 10<sup>5</sup> to 4 X 10<sup>5</sup> cells per well were cultured in complete medium (RPMI 1640 supplemented with gentamicin, 2-mercaptoethanol, L-glutamine, and 10% screened pooled A+ human serum; Trimar, Hollywood, Calif.) in 96-well flat-bottom plates with or without 10 μg of the indicated antigens per ml or 5 μg of phytohemagglutinin per ml (Sigma Immunochemicals, St. Louis, Mo.) for 5 days. The cells were then pulsed with

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1  $\mu$ Ci of [³H]thymidine for the final 18 h of culture. For determination of cytokine production 0.5 to 1 ml of PBMC was cultured at 1 X 10<sup>6</sup> to 2 X 10<sup>6</sup> cells per ml with or without the *Leishmania* antigens for 48 and 72 h.

The supernatants and cells were harvested and analyzed for secreted cytokine or cytokine mRNAs. Aliquots of the supernatants were assayed for gamma interferon (IFN-γ), tumor necrosis factor alpha (TNF-α), interleukin-4 (IL-4), and IL-10 as described in Skeiky et al., J. Exp. Med. 181:1527-1537 (1995). For cytokine mRNA PCR analysis, total RNA was isolated from PBMC and cDNA was synthesized by using poly(dT) (Pharmacia, Piscataway, NJ) and avian mycloblastosis virus reverse transcriptase. Following normalization to β-actin, diluted cDNA was amplified by PCR using Taq polymerase (Perkin-Elmer Cetus, Foster City, CA) with 0.2 µM concentrations of the respective 5' and 3' external primers in a reaction volume of 50 µl. The nucleotide sequences of the primary pairs and the PCR conditions used were as described in Skeiky et al., J. Exp. Med. 181:1527-1537 (1995). We verified that our PCR conditions were within the semiquantitative range by initially performing serial dilutions of the cDNAs and varying the number of cycles used for PCR. Plasmids containing the human sequences for IL-2, IFN-γ, IL-4, IL-10, and β-actin were digested, and the DNA inserts were purified after separation on 1% agarose gels. Radiolabeled <sup>32</sup>P probes were prepared by the random priming method. PCR products were analyzed by electrophoresis on 1.5% agarose gels, transferred to nylon membranes, and probed with the appropriate <sup>32</sup>P-labeled DNA insert.

A recombinant clone was identified in the above assays which, following sequence comparison of its predicted amino acid sequence with sequences of other proteins, was identified as a *Leishmania braziliensis* homolog of the eukaryotic 83 kD heat shock protein (Lbhsp83). The sequence of the clone is provided in SEQ ID NO:5 and the deduced protein sequence is provided in SEQ ID NO:6. On the basis of the homology, this clone, designated Lbhsp83a, appears to lack the first 47 residues of the full length 703 amino acid residues. Lbhsp83 has an overall homology of 94% (91% identity and 3% conservative substitution), 91% (84% identity and 7% conservative substitution) and 77% (61% identity and 16% conservative substitution) with *L*.

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amazonensis hsp83, T. cruzi hsp83 and human hsp89, respectively. A second clone (designated Lbhsp83b), which contained the 43 kD C-terminal portion of hsp83 (residues 331 to 703) was also isolated. Figure 19 presents a comparison of the Lbhsp83 sequence with L. amazonensis hsp83(Lahsp83), T. cruzi hsp83 (Tchsp83) and human hsp89 (Huhsp89).

The results of proliferation assays using Lbhsp83a are shown in Table 1. Cells from all mucosal leishmaniasis (ML) patients proliferated strongly in response to Lbhsp83a, with stimulation indices (SIs) ranging from 19 to 558 (as compared to 20 to 1,634 for parasite lysate). Proliferation of PBMC from cutaneous leishmaniasis (CL) patients was variable and except for levels in two patients (IV and VII), levels were significantly lower than those of ML patients. By comparison, the proliferative responses of individuals with self-healing CL to Lbhsp83a were similar to those of individuals with ML. However, the responses of all six self-healing individuals to Lbhsp83 were consistently higher than those to Lbhsp83b. This suggests that PBMC from self-healing CL patients preferentially recognize one or more T-cell epitopes located within the amino portion of Lbhsp83.

Table 1

In vitro Proliferation of PMBC from L. braziliensis-infected Individuals
in Response to Lbhsp83

. :	Mean [3H]thymidine incorporation [103 cpm (SD)], SI with:			
Group and Patient	Lysate	Lbhsp83a	Lbhsp83b	-
ML				
İ	41.3, (1.3), 294	32.5, (6.6), 221	46.7, (1.4), 318	
П	44.2, (0.5), 104	20, (3.7), 47	36.7, (0.76), 86	
- III	27.4, (1.5), 150	8.1, (1.7), 44	9.9, (0.32), 54	
IV	52.7, (3.3), 138	54.1, (6.2), 142	32.0, (1.3), 84	
V	140.6, (7.6), 308	151.8, (57), 333	150.4, (7.9), 331	
VI	15.8, (1.8), 20	21.3, (4.4), 28	14.4, (1.3), 19	
VII	300.1, (9.4), 1634	102.1, (7.6), 558	41.7, (4.9), 228	

	Mean ['H]thymid	line incorporation [103 cp	m (SD)], SI with:
Group and Patient	Lysate	Lbhsp83a	Lbhsp83b
CL		•	
I	0.26, (0.0), 1.5	0.57, (0.3), 3.3	0.43, (0.17), 3.3
II	55.63, (8.6), 218	0.42, (0.0), 1.6	0.8, (0.14), 3.2
III	0.39, (0.5), 4.0	3.4, (0.5), 9	2.6, (0.9), 6.6
IV	19.14, (1.3), 87	7.17, (0.6), 32	5.9, (0.9), 27
v	0.32, (0.2), 3.0	1.47, (0.5), 14	0.3, (0.1), 3.0
VI ·	0.77, (0.1), 4.7	1.44, (0.2), 9	1.3, (0.6), 8.0
VII	4.01, (1.0), 2.0	60.3, (8.5), 15	66.7, (3.9), 16.6
Self-healing CL			
1	19.7, (4.4), 94	61.3, (4.6), 293	5.0, (2.0), 24
* II · · ·	0.6, (0.1), 6.5	7.0, (2.0), 79	1.2, (0.8), 13
111	59.6, (7.1), 519	49.4, (3.1), 429	21.4, (3.7), 186
IV	0.2, (0.1), 1.6	13.1, (1.7), 108	0.6, (0.1), 5
V	27.1, (2.0), 225	6.3, (2.6), 52	3.0, (1.5), 25
VI	130.3, (14), 340	28.2, (2.9), 74	7.7, (3.8), 20
Control (uninfected)			
. 1	0.19, (0.0), 1.4	0.18, (0.0), 1.3	0.40, (0.16), 2.8
II	0.31, (0.1), 1.7	0.19, (0.0), 1.0	0.27, (0.0), 1.5
III	0.44, (0.2), 4.1	0.48, (0.1), 5.0	0.51, (0.2), 5.2
IV	0.4, (0.1), 3.2	0.52, (0.2), 5.1	0.50, (0.1), 5.0

A more detailed analysis of cytokine patterns of PBMC from ML patients was performed by reverse transcriptase PCR. Cytokine mRNAs were evaluated in cells prior to culturing (Figure 9, lanes O) or following culturing in the absence (lanes –) or presence of the indicated antigen for 48 and 72 h. Figure 4A shows the results for five of the six ML patients whose PBMC were analyzed. In about half of the ML patients, noncultured (resting) PBMC had detectable levels of mRNA for IFN-γ, IL-2, and IL-4 but not IL-10. CL patient PBMC, however, had IL-10 mRNA in the resting state in addition to mRNAs for the other cytokines tested (Figure 4B). Following *in vitro* culture without antigen, the levels of mRNA for IFN-γ, IL-2, and IL-4 in resting cells from ML patients decreased to background levels while IL-10 mRNA levels increased. In contrast, PBMC of most CL patients had stable or increased IL-10

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mRNA, while the mRNAs for IL-2, IFN-γ, and IL-4 were reduced to barely detectable levels in the absence of antigen stimulation.

In PBMC of three ML patients, stimulation with lysate resulted in increased expression of mRNA for IFN-γ, IL-2, and IL-4 but not IL-10. By comparison, both Lbhsp83 polypeptides elicited the production of mRNA for IFN-γ and IL-2 from all ML patient PBMC tested. In contrast, profiles of mRNA for IL-10 and IL-4 differed for the two hsp83 polypeptides. Lbhsp83a stimulated the production of IL-10 but not IL-4 mRNA (patients I, II, III, and IV), while Lbhsp83b stimulated the production of IL-4 but not IL-10 mRNA in all six patients.

All CL patients tested responded to both Lbhsp83 polypeptides as well as to the parasite lysate by upregulating the synthesis of mRNAs for IL-2 and IFN-γ, and in two of four patients (I and IV), the level of IL-4 mRNA also increased, indicating stimulation of both Th1 and Th2 cytokines. Interestingly and as in the case of ML patient uncultured PBMC which did not have detectable levels of IL-10 mRNA, Lbhsp83a and not Lbhsp83b stimulated PBMC from one CL patient (IV) to synthesize IL-10 mRNA. However, in the other three patients (I, II, and III) with resting levels of IL-10 mRNA, both rLbhsp83 polypeptides as well as the parasite lysate downregulated the expression of IL-10 mRNA.

PBMC supernatants were also assayed for the presence of secreted IFN-γ

7. TNF-α, IL-4, and IL-10. Cells from all ML and self-healing CL patients (seven and six patients, respectively) and from four of seven CL patients were analyzed for secreted IFN-γ following stimulation with both rLbhsp83 polypeptides, parasite lysate and Lbhsp70, an *L. braziliensis* protein homologous to the eukaryotic 70 kD heat shock protein (Figure 10A). In general, rLbhsp83a stimulated patient PBMC to secrete higher levels of IFN-γ than did rLbhsp83b (0.2 to 36 and 0.13 to 28 ng/ml, respectively). The presence of secreted IFN-γ correlated well with the corresponding mRNA detected by PCR

PBMC from four of five ML patients (I, II, V, and VII) had supernatant TNF-α levels (0.8 to 2.2 ng/ml) higher than those detected in cultures of PBMC from uninfected controls following stimulation with parasite lysate (Figure 10B). Similarly,

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the same PBMC were stimulated by rLbhsp83 to produce levels of TNF- $\alpha$  in supernatant ranging from 0.61 to 2.9 ng/ml. Compared with those of uninfected controls, PBMC from three (I, V, and VI), five (I, II, IV, V, and VI), and two (II and V) of six individuals analyzed produced higher levels of TNF- $\alpha$  in response to parasite lysate, rLbhsp83a, and rLbhsp83b, respectively. The levels of TNF- $\alpha$  produced by PBMC from CL patients in response to parasite lysate were comparable to those produced by uninfected controls. However, rLbhsp83 stimulated TNF- $\alpha$  production in the PBMC of two of these patients. rLbhsp83a stimulated higher levels of TNF- $\alpha$  production than did rLbhsp83b. In the absence of antigen stimulation, only PBMC from ML patients (five of six) produced detectable levels of supernatant TNF- $\alpha$  (60 to 190 pg/ml).

In agreement with the IL-10 mRNA, IL-10 was detected by ELISA in the antigen-stimulated PMBC culture supernatants from ML and CL patients. The levels (49 to 190 pg) were significantly higher (up to 10-fold) following stimulation with rLbhsp83a compared with those after parallel stimulation of the same cells with rLbhsp83b (Figure 11). Parasite lysate also stimulated PMBC from some of the patients to produce IL-10. Although rLbhsp83 stimulated PMBC from uninfected individuals to produce IL-10, with one exception, the levels were lower than those observed with patient PMBC. IL-4 was not detected in any of the supernatants analyzed. Therefore, the level of any secreted IL-4 is below the detection limit of the ELISA employed (50 pg/ml). Taken together, the results demonstrate that a predominant Th1-type cytokine profile is associated with PMBC from *L. braziliensis*-infected individuals following stimulation with rLbhsp83 polypeptides.

antibody production to Lbhsp83, we compared the antibody (immunoglobulin G) reactivities to Lbhsp83 in sera from the three patient groups (Figure 12). The ELISA reactivities of ML patient sera with rLbhsp83a were comparable to those observed with parasite lysate, and in general, there was a direct correlation between ML patient anti-Lbhsp83 antibody titer and T-cell proliferation. Of 23 serum samples from ML patients analyzed, 22 were positive (~96%) with absorbance values of 0.20 to >3.0. Eleven of

the ML patient serum samples had optical density values that were >1. In general, CL patients had significantly lower anti-Lbhsp83 antibody titers ( $\frac{x}{x}$  = 0.74; standard error of the mean [SEM] = 0.1) compared to those of ML patients. Therefore, ML and CL patient anti-rhsp83 antibody titers correlated with their respective T-cell proliferative responses. Anti-rLbhsp83 antibody titers were significantly higher in patients with ML ( $\frac{x}{x}$  = 1.5; SEM = 0.2) than in self-healing CL patients ( $\frac{x}{x}$  = 0.35; SEM = 0.056), although their T-cell proliferative responses were similar. In fact, anti-Lbhsp83 antibody titers in serum from self-healing CL patients were comparable to those from uninfected controls ( $\frac{x}{x}$  = 0.24; SEM = 0.028). By using 2 standard deviations greater than the mean absorbance value of uninfected control (0.484) as a criterion for positive reactivity to Lbhsp83, eight of nine of the self-healing patient serum samples tested were negative.

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### **EXAMPLE 4**

### PREPARATION OF CLONES ENCODING LT-210

This Example illustrates the preparation of clones encoding portions of the *Leishmania* antigen Lt-210, and which has the sequence provided in SEQ ID NO:8.

An expression library was constructed from *L. tropica* (MHOM/SA/91/WR1063C) genomic DNA. The DNA was isolated by solubilizing *L. tropica* promastigotes in 10mM Tris-HCl, pH 8.3, 50mM EDTA, 1% SDS and treating with 100µg/ml RNaseA and 100µg/ml proteinase K. The sample was then sequentially extracted with an equal volume of phenol, phenol: chloroform (1:1), and Chloroform. DNA was precipitated by adding 0.1 volume of 3M sodium acetate (pH 5.2) and 2.5 volume 95% ethanol. The precipitate was resuspended in 10µM Tris, 1mM EDTA. DNA was sheared by passage through a 30-gauge needle to a size range of 2-6 kilobase, and was repaired by incubation with DNA polI in the presence of 100 µM each dATP, dCTP, dGTP, and dTTP. *Eco*RI adapters were ligated to the DNA

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fragments. After removal of unligated adapters by passage over a G-25 Sephadex<sup>™</sup> column, the fragments were inserted in *Eco*RI cut Lambda ZapII (Stratagene, La Jolla, CA).

Approximately 43,000 pfu were plated and screened with sera isolated from viscerotropic leishmaniasis (VTL) patients. Sera from VTL patients were received from Drs. M. Grogl and A. Magill. The VTL patient group included eight individuals from whom parasites were isolated and cultured, seven of which had confirmed infection with *L. tropica*. Four other patients were culture negative, but were still considered to be infected based on either PCR analysis or a positive monoclonal antibody smear (Dr. Max Grogl, personal communication). Serum samples from the 11 infected patients were pooled and anti-*E. coli* reactivity removed by affinity chromatography (Sambrook et al., *supra*, p. 12.27-12.28). Lambda phage expressing reactive proteins were detected after antibody binding by protein A-horseradish peroxidase and ABTS substrate.

Three clones, Lt-1, Lt-2, and Lt-3, containing a portion of the Lt-210 gene were identified and purified. The clones ranged in size from 1.4 to 3.3 kb and encoded polypeptides of 75 kD, 70 kD, and 120 kD, respectively. These three clones contain partial sequences of the Lt-210 gene. Lt-1 and Lt-2 are overlapping clones and were chosen for further study.

The DNA sequences of Lt-1 and Lt-2 were determined. Exonuclease III digestion was used to create overlapping deletions of the clones (Heinikoff, *Gene 28*:351-359, 1984). Single strand template was prepared and the sequence determined with Applied Biosystems Automated Sequencer model 373A or by Sanger dideoxy sequencing. The sequence on both strands of the coding portion of Lt-1 clone was determined. The partial sequence of one strand of Lt-2 clone was determined.

SEQ ID NO:7 presents the DNA sequence of Lt-1, and SEQ ID NO:8 provides the predicted amino acid sequence of the open reading frame. The DNA sequence of the coding portion of the Lt-1 clone includes a repeated nucleotide sequence at the 5' portion of the clone containing eight copies of a 99 bp repeat, three copies of a 60 bp repeat unit, which is part of the larger 99 bp repeat, and 800 bp of

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non-repeat sequence. The deduced amino acid sequence of the 99 bp repeat contains limited degeneracies. The mass of the predicted recombinant protein is 67,060 Daltons. A database search of PIR with the predicted amino acid sequence of the open reading frame yielded no significant homology to previously submitted sequences. Predicted secondary structure of the repeat portion of the clone is entirely  $\alpha$ -helical.

Sequence analysis of Lt-2 revealed that the 3' portion of the clone consisted of a mixture of 60 and 99 bp repeats that were identical, excepting occasional degeneracies, to the 60 and 99 bp repeats observed in Lt-1. Collectively, the sequencing data suggest that Lt-1 and Lt-2 are different portions of the same gene, Lt-2 being upstream of Lt-1, with possibly a small overlap.

Hybridization analysis confirmed that rLt-2 and rLt-1 contain overlapping sequences. Genomic DNAs of various *Leishmania* species were restricted with a variety of enzymes, separated by agarose gel electrophoresis, and blotted on Nytran membrane filter (Schleicher & Schuell, Keene, NH). Inserts from rLt-1 and rLt-2 were labeled with <sup>32</sup>P-CTP by reverse transcriptase from random oligonucleotide primers and used as probes after separation from unincorporated nucleotides on a Sephadex G-50 column. Hybridizations using the rLt-1 or the rLt-2 probe are performed in 0.2M NaH<sub>2</sub>PO<sub>4</sub>/3.6 M NaCl at 65°C, whereas hybridization using the rLt-1 r probe is performed in 0.2 M NaH<sub>2</sub>PO<sub>4</sub>/3.6 M NaCl/0.2 M EDTA at 60°C overnight. Filters are washed in 0.075 M NaCl/0.0075 M sodium citrate pH 7.0 (0.15 M NaCl/0.0150 M sodium citrate for the Lt-lr probe), plus 0.5% SDS at the same temperature as hybridization.

Genomic DNA from a number of Leishmania species including L. tropica were analyzed by Southern blots as described above using the Lt-1, Lt-2, and Lt-1r inserts separately as probes. Collectively, various digests of L. tropica DNA indicate that this gene has a low copy number. A similar, overlapping pattern was observed using either the Lt-1 or Lt-2 insert as a probe, consistent with the premise that these two clones contain sequences near or overlapping one another. In addition, sequences hybridizing with these clones are present in other Leishmania species.

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L. tropica isolates have limited heterogeneity. Southern analyses of digested genomic DNA from four L. tropica parasite strains isolated from VTL patients and three L. tropica parasite strains isolated from CL cases (two human, one canine) were performed. The Lt-lr insert described below was labeled and used as a probe. The seven different L. tropica isolates yielded similar intensities and restriction patterns, with only a single restriction fragment length polymorphism among the isolates. These data, along with Southern analyses with additional enzymes, indicate limited heterogeneity in this region among the L. tropica isolates.

The recombinant proteins of Lt-1 and Lt-2 were expressed and purified. The nested deletion set of Lt-1 formed for sequencing included a clone referred to as Lt-lr, which contains one and one-third repeats. This polypeptide was also expressed and purified. In vivo excision of the pBluescript SK- phagemid from Lambda Zap II was performed according to the manufacturer's protocol. Phagemid virus particles were used to infect E. coli XL-1 Blue. Production of protein was induced by the addition of IPTG. Protein was recovered by first lysing pellets of induced bacteria in buffer (LB, 50 mM Tris-HCl, pH 8.0, 100 mM NaCl, 10 mM EDTA) using a combination of lysozyme (750µg/mL) and sonication. rLt-1, rLt-2, and rLt-1r, were recovered from the inclusion bodies after solubilization in 8M urea (rLt-1 and rLt-2) or 4M urea (rLt-1r). Proteins rLt-1 and rLt-2 were enriched and separated by precipitation with 25%-40% ammonium sulfate and rLt-lr was enriched by precipitation with 10%-25% ammonium sulfate. The proteins were further purified by preparative gel electrophoresis in 10% Recombinant proteins were eluted from the gels and dialyzed in SDS-PAGE. Concentration was measured by the Pierce phosphate-buffered saline (PBS). (Rockford, IL) BCA assay, and purity assessed by Coomassie blue staining after SDS-PAGE.

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#### **EXAMPLE 5**

#### PREPARATION OF LBEIF4A

This example illustrates the molecular cloning of a DNA sequence encoding the L. braziliensis ribosomal antigen LbeIF4A.

A genomic expression library was constructed with sheared DNA from L. braziliensis (MHOM/BR/75/M2903) in bacteriophage  $\lambda$ ZAPII (Stratagene, La Jolla, CA). The expression library was screened with E. coli-preadsorbed patient sera from an L. braziliensis-infected individual with mucosal leishmaniasis. Plaques containing immunoreactive recombinant antigens were purified, and the pBSK(-) phagemid excised using the manufacturer's protocols. Nested deletions were performed with Exonuclease III to generate overlapping deletions for single stranded template preparations and sequencing. Single stranded templates were isolated following infection with VCSM13 helper phage as recommended by the manufacturer (Stratagene, La Jolla, CA) and sequenced by the dideoxy chain terminator method or by the Taq dye terminator system using the Applied Biosystems Automated Sequencer Model 373A.

The immunoreactive recombinant antigens were then analyzed in patient T-cell assays for their ability to stimulate a proliferative and cytokine production, as described in Examples 7 and 8 below.

A recombinant clone was identified in the above assays which, following sequence comparison of its predicted amino acid sequence with sequences of other proteins, was identified as a *Leishmania braziliensis* homolog of the eukaryotic initiation factor 4A (eIF4A). The isolated clone (pLeIF.1) lacked the first 48 amino acid residues (144 nucleotides) of the full length protein sequence. The pLeIF.1 insert was subsequently used to isolate the full length genomic sequence.

SEQ ID NO:9 shows the entire nucleotide sequence of the full-length LbeIF4A polypeptide. The open reading frame (nucleotides 115 to 1323) encodes a 403 amino acid protein with a predicted molecular weight of 45.3 kD. A comparison of the predicted protein sequence of LbeIF4A with the homologous proteins from tobacco (TeIF4A), mouse (MeIF4A), and yeast (YeIF4A) shows extensive sequence homology,

with the first 20-30 amino acids being the most variable. The lengths (403, 413, 407, and 395 amino acids), molecular weights (45.3, 46.8, 46.4, and 44.7 kDa), and isoelectric points (5.9, 5.4, 5.5, and 4.9) of LbeIF4A, TeIF4A, MeIF4A and YeIF4A, respectively, are similar. LbeIF4A shows an overall homology of 75.5% (57% identity, 18.5% conservative substitution) with TeIF4A, 68.6% (50% identity, 18.6% conservative substitution) with MeIF4A and 67.2% (47.6% identity, 19.6% conservative substitution) with YeIF4A.

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#### EXAMPLE 6

## PREPARATION OF SOLUBLE LEISHMANIA ANTIGENS

This Example illustrates the preparation of soluble *Leishmania* antigens from an *L. major* culture supernatant. *L. major* promastigotes were grown to late log phase in complex medium with serum until they reached a density of 2-3 x 10<sup>7</sup> viable organisms per mL of medium. The organisms were thoroughly washed to remove medium components and resuspended at 2-3 x 10<sup>7</sup> viable organisms per mL of defined serum-free medium consisting of equal parts RPMI 1640 and medium 199, both from Gibco BRL, Gaithersburg, MD. After 8-12 hours, the supernatant was removed, concentrated 10 fold and dialyzed against phosphate-buffered saline for 24 hours. Protein concentration was then determined and the presence of at least eight different antigens confirmed by SDS-PAGE. This mixture is referred to herein as "soluble *Leishmania* antigens."

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#### EXAMPLE 7

# Comparison of Interleukin-4 and Interferon- $\gamma$ Production Stimulated by *Leishmania* Antigens

This Example illustrates the immunogenic properties of the antigens prepared according to Examples 1, 2, 5 and 6, as determined by their ability to stimulate IL-4 and IFN-γ in lymph node cultures from infected mice and in human PBMC preparations. Lymph node cultures for use in these studies were prepared from *L. major*-infected BALB/c mice 10 days after infection, as described in Example 2. PBMC were prepared using peripheral blood obtained from individuals with cured *L. donovani* infections who were immunologically responsive to *Leishmania*. Diagnosis of the patients was made by clinical findings associated with at least one of the following: isolation of parasite from lesions, a positive skin test with *Leishmania* lysate or a positive serological test. Uninfected individuals were identified based on a lack of clinical signs or symptoms, a lack of history of exposure or travel to endemic areas, and the absence of a serological or cellular response to *Leishmania* antigens. Peripheral blood was collected and PBMC isolated by density centrifugation through Ficoll<sup>TM</sup> (Winthrop Laboratories, New York).

Culture supernatants were assayed for the levels of secreted IL-4 and IFN-γ. IFN-γ was quantitated by a double sandwich ELISA using mouse anti-human IFN-γ mAb (Chemicon, Temucula, CA) and polyclonal rabbit anti-human IFN-γ serum. Human rIFN-γ (Genentech Inc., San Francisco, CA) was used to generate a standard curve. IL-4 was quantitated in supernatants by a double sandwich ELISA using a mouse anti-human IL-4 mAb (M1) and a polyclonal rabbit anti-human IL-4 sera (P3). Human IL-4 (Immunex Corp., Seattle, WA) was used to generate a standard curve ranging from 50 pg/ml to 1 ng/ml.

Figures 13A and 13B, illustrate the mean level of secreted IL-4 and IFNγ, respectively, 72 hours after addition of 10 µg/mL of each of the following antigens to a lymph node culture prepared as described above: soluble *Leishmania* antigen (i.e., an extract prepared from ruptured promastigotes which contains membrane and internal antigens (SLA)), Ldp23, LbeIF4A (LeIF), Lbhsp83, M15 and LmeIF (the *L. major* homolog of LbeIF4A). The levels of secreted IL-4 and IFN-γ in medium alone (*i.e.*, unstimulated) are also shown. While SLA elicits a predominantly Th2 response from lymph node cells of Leishmania-infected mice, Ldp23, LbeIF4A, Lbhsp83 and M15 elicited relatively little IL-4 and large amounts of IFN-γ, consistent with a Th1 response profile.

Figure 14 shows the level of secreted IFN-γ in culture filtrate from infected and uninfected human PBMC preparations 72 hours after addition of 10 μg/mL *L. major* lysate, M15 or L-Rack, an immunodominant leishmanial antigen in murine leishmaniasis. Similarly, Figure 15 illustrates the level of secreted IFN-γ in culture filtrate from infected and uninfected human PBMC preparations 72 hours after addition of 10μg/mL *L. major* lysate, soluble *Leishmania* antigens (prepared as described in Example 6) or L-Rack. These results indicate that M15 and soluble *Leishmania* antigens, but not L-Rack, are potent stimulators of IFN-γ production in patient PBMC, but not in PBMC obtained from uninfected individuals. Thus, M15 and soluble *Leishmania* antigens elicit a dominant Th1 cytokine profile in both mice and humans infected with *Leishmania*.

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#### **EXAMPLE 8**

# COMPARISON OF PROLIFERATION STIMULATED BY LEISHMANIA ANTIGENS

This Example illustrates the immunogenic properties of the antigens prepared according to Examples 1, 2, 5 and 6, as determined by their ability to stimulate proliferation in lymph node cultures from infected mice and in human PBMC preparations.

For *in vitro* proliferation assays,  $2 - 4 \times 10^5$  cells/well were cultured in complete medium (RPMI 1640 supplemented with gentamycin, 2-ME, L-glutamine, and 10% screened pooled A+ human serum; Trimar, Hollywood, CA) in 96-well flat bottom plates with or without 10  $\mu$ g/ml of the indicated antigens or 5  $\mu$ g/ml PHA

(Sigma Immunochemicals, St. Louis, MO) for five days. The cells were then pulsed with 1  $\mu$ Ci of [<sup>3</sup>H] thymidine for the final 18 hours of culture.

Figure 16 illustrates the proliferation observed after addition of 10  $\mu$  g/mL or 20  $\mu$ g/mL of each of the following antigens to a lymph node culture prepared as described in Example 7: SLA, Ldp23, LbeIF4A, Lbhsp83, and M15. The level of proliferation without the addition of antigen is also shown. Data are represented as mean cpm. These results demonstrate that a variety of leishmanial antigens are capable of stimulatory lymph node cell proliferation from *Leishmania*-infected mice.

Figures 17 and 18 illustrate the proliferation observed in human PBMC preparations from Leishmania-immune and uninfected individuals following the addition of 10 µg/mL M15 and soluble Leishmania antigens, respectively. These values are compared to the proliferation observed following the addition of culture medium, L. major lysate or L-Rack. The results show that M15 and soluble Leishmania antigens stimulate proliferation in Leishmania-immune PBMC, but not in PBMC obtained from uninfected individuals, demonstrating that M15 and soluble antigens (but not L-Rack) are recognized by PBMC from individuals immune to Leishmania due to a previous infection.

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#### **EXAMPLE 9**

#### PREPARATION OF LMSP1A AND LMSP9A

This Example illustrates the preparation of two soluble *Leishmania* antigens, Lmspla and Lmsp9a.

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# A. Purification of Lmspla and Lmsp9a from a mixture of soluble L. major antigens

A high titer rabbit sera was raised against *L. major* soluble antigens, prepared as described above in Example 6. Specifically, a New Zealand white rabbit was immunized subcutaneously at multiple sites with 180 µg of *L. major* soluble antigens in a suspension containing 100 µg muramyl dipeptide and 50 % incomplete

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Freund's adjuvant. Six weeks later the rabbit was given a subcutaneous boost of  $100 \mu g$  of the same soluble antigen preparation in incomplete Freund's adjuvant. This was followed by two intravenous boosts spaced two weeks apart, each with  $100 \mu g$  of the soluble antigen preparation. Sera was collected from the rabbit 11 days after the final boost.

Anti E. coli antibody reactivities were removed from the rabbit sera by pre-adsorbing on nitrocellulose filters containing lysed E. coli. Adsorbed sera were evaluated by Western blot analysis using 10 µg Leishmania promastigote lysate (lane 1) and 1 µg soluble L. major antigen mixture (lane 2). As shown in Figure 20, the rabbit sera was found to be reactive with seven dominant antigens of the soluble L. major antigen mixture with molecular weights ranging from 18 to >200 kDa. A four times longer exposure of the same blot revealed three additional immunoreactive species with molecular weights less than 18 kDa. The same sera reacted with approximately 10 antigens of the promastigote lysate, but with a pattern significantly different from that observed with the soluble L. major antigens (Figure 20). This is suggestive of potential post-translational modification of the same antigen before (intracellular localization) and after secretion/shedding. Such modifications may include cleavage of a leader sequence and/or the addition of carbohydrate molecules to the secreted/shed antigens.

The rabbit sera described above was subsequently used to screen an *L. major* cDNA expression library prepared from *L. major* promastigote RNA using the unidirectional Lambda ZAP (uni-ZAP) kit (Stratagene) according to the manufacturer's protocol. A total of 70,000 pfu of the amplified cDNA library was screened with the rabbit sera at a 1:250 dilution. Nineteen positive clones were confirmed in the tertiary screening. The phagemid were excised and DNA from each of the 19 clones was sequenced using a Perkin Elmer/Applied Biosystems Division automated sequencer Model 373A. All 19 clones were found to represent two distinct sequences, referred to as Lmsp1a and Lmsp9a. The determined cDNA sequences for Lmsp1a and Lmsp9a are provided in SEQ ID NO: 19 and 21, respectively, with the corresponding amino acid sequences being provided in SEQ ID NO: 20 and 22, respectively.

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# B. Characterization of Lmspla and Lmsp9a

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Fig. 21 shows the full-length cDNA (SEQ ID NO: 19) and predicted amino acid sequence (SEQ ID NO: 20) for the antigen Lmsp1a. The EcoRI/XhoI insert is 1019 bp long and contains the following features: a) the last 17 nt of the spliced leader sequence characteristic of all trypanosoma nuclearly encoded mRNA; b) 39 nt of 5' untranslated sequence; c) an open reading frame of 453 nt long coding for a 151 deduced amino acid sequence with a predicted molecular mass of 16.641 kDa; and d) 471 nt of 3' untranslated sequence terminating with a poly A tail. The predicted amino acid sequence contains three potential phosphorylation sites at amino acid residues 3, 85 and 102. In addition, Lmsp1a contains an RGD sequence at residue 104, a sequence that may play a role in parasite invasion of the macrophage. RGD sequences have been shown to mediate the binding of various adhesion proteins to their cell surface receptors. There is no obvious leader sequence (secretory signal) at the amino terminal portion suggesting that the protein might be shed or excreted. Lmsp1a appears to be one of the most abundant antigens found in the culture supernatant of live promastigote, since 17 of the 19 clones contain sequences of variable lengths identical to Lmsp1a.

Comparison of the amino acid sequence of Lmps1a with known sequences using the DNA STAR system (Version 87) revealed that Lmsp1a shares between 65% to 70% homology with the eukaryotic nucleoside diphosphate kinase protein, also referred to in the mouse and human as a tumor metastasis inhibitor gene.

Southern blot analysis of genomic DNA from *L. major* (Friedlander strain) digested with a panel of restriction enzymes (lanes 1 to 7) and six other *Leishmania* species of different geographic locations digested with PstI (lanes 8 to 13) using the full-length cDNA insert of Lmps1a, demonstrated that Lmsp1a is present in all the species characterized with a high degree of conservation (Fig. 22). This suggests evolutionary significance for the maintenance of Lmsp1a and the existence of homologous species among all the *Leishmania* species.

The remaining two cDNA clones isolated from the soluble *L. major* antigen mixture represent identical sequences (referred to as Lmsp9a; SEQ ID NO: 21), suggesting that the two copies resulted from amplification of the primary library.

Sequencing of the Lmsp9a cDNA revealed that the clone does not contain the full length 5' sequence since it is lacking both the spliced leader and 5' untranslated sequences. The 3' end of the cDNA contains a poly A stretch, as would be expected for a *Leishmania* mRNA. Of the predicted translated sequence (SEQ ID NO: 22), 34 of the 201 amino acids (17%) represent cysteine residues. Comparison of the predicted protein sequence with those of known proteins as described above, revealed some homology with other cysteine rich proteins such as the major surface trophozoite antigen of *Giardia lamblia* and furin proteases.

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### EXAMPLE 10

## PREPARATION AND CHARACTERIZATION OF MAPS-1A

This Example illustrates the preparation and characterization of the Leishmania antigen MAPS-1A (SEQ ID NO: 24).

A pool of sera was obtained from 5 BALB/c mice that had been given a primary immunization and two boosts with crude *L. major* promastigote culture supernatant as described below in Example 12. These mice were subsequently shown to be protected when challenged with a dose of live *L. major* promastigotes generally found to be lethal. The mouse sera thus obtained were used to screen an *L. major* amastigote cDNA expression library prepared as described in Example 1. Several seroreactive clones were isolated and sequenced using a Perkin Elmer/Applied Biosystems Division automated sequencer Model 373A (Foster City, CA).

One of these clones, referred to herein as MAPS-1A, was found to be full-length. Comparison of the cDNA and deduced amino acid sequences for MAPS-1A (SEQ ID Nos: 23 and 24, respectively) with known sequences in the gene bank using the DNA STAR system revealed no significant homologies to known *Leishmania* sequences, although some sequence similarity was found to a group of proteins, known as thiol-specific antioxidants, found in other organisms.

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Recombinant MAPS-1A protein having an amino-terminal HIS-Tag was prepared using a high level *E. coli* expression system and recombinant protein was purified by affinity chromatography as described in Example 1. Southern blot analysis of genomic DNA from *L. major* digested with a panel of restriction enzymes, seven other *Leishmania* species digested with PstI, and two other infectious-disease pathogens (*T. cruzi* and *T. brucei*), using the full length insert of MAPS-1A, demonstrated that MAPS-1A is present in all eight *Leishmania* species tested (Figure 23). Northern blot analysis of *L. major* promastigote and amastigote RNAs indicated that MAPS-1A is constitutively expressed.

Using oligonucleotide primers (SEQ ID NOs:27 and 28) based on the MAPS-1A cDNA sequence provided in SEQ ID NO: 23, the corresponding gene was isolated from *L. tropica* by means of PCR (using 30 cycles of the following temperature step sequence: 94 °C, 1 minute; 50 °C, 1 minute; 72 °C, 1 minute) The determined cDNA sequence for the *L. tropica* MAPS-1A protein is provided in SEQ ID NO: 25, with the corresponding amino acid sequence being provided in SEQ ID NO: 26.

The ability of recombinant MAPS-1A to stimulate cell proliferation was investigated as follows. PBMC from 3 *L. braziliensis*-infected patients having active mucosal leishmaniasis, from 4 patients post kala-azar infection (previously infected with *L. chagasi* and/or *L. donovani*) and from 3 uninfected-individuals were prepared as described above in Example 7. The ability of MAPS-1A to stimulate proliferation of these PBMC was determined as described in Example 8 above. As shown in Figure 24, significant levels of MAPS-1A specific PBMC proliferation were seen in 2 of the 7 *Leishmania* patients.

The ability of MAPS-1A to stimulate proliferation in mice lymph node cultures was determined as described in Example 8. Figure 25 shows the amount of proliferation stimulated by MAPS-1A (at 25  $\mu$ g/ml, 5  $\mu$ g/ml and 1  $\mu$ g/ml) as compared to that stimulated by the positive control ConA and by crude L. major promastigote supernatant proteins, 20 days post-infection with L. major. Cells isolated 20 days post-

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infection were highly responsive to MAPS-1A, whereas cells isolated 10 days post-infection were unresponsive.

# EXAMPLE 11

# IMMUNOREACTIVITY OF SOLUBLE *LEISHMANIA* ANTIGENS WITH SERA FROM *LEISHMANIA*-INFECTED PATIENTS

The reactivity of MAPS-1A with sera from uninfected individuals, from human leishmaniasis patients with cutaneous infection, from human patients with acute visceral leishmaniasis, and from *L. major*-infected BALB/c mice was determined as follows.

Assays were performed in 96-well plates coated with 200 ng antigen diluted to 50 μL in carbonate coating buffer, pH 9.6. The wells were coated overnight at 4 °C (or 2 hours at 37 °C). The plate contents were then removed and the wells were blocked for 2 hours with 200 μL of PBS/1% BSA. After the blocking step, the wells were washed five times with PBS/0.1% Tween 20<sup>TM</sup>. 50 μL sera, diluted 1:100 in PBS/0.1% Tween 20<sup>TM</sup>/0.1% BSA, was then added to each well and incubated for 30 minutes at room temperature. The plates were then washed again five times with PBS/0.1% Tween 20<sup>TM</sup>.

The enyzme conjugate (horseradish peroxidase - Protein A, Zymed, San Francisco, CA) was then diluted 1:10,000 in PBS/0.1% Tween 20<sup>TM</sup>/0.1% BSA, and 50 μL of the diluted conjugate was added to each well and incubated for 30 minutes at room temperature. Following incubation, the wells were washed five times with PBS/0.1% Tween 20<sup>TM</sup>. 100 μL of tetramethylbenzidine peroxidase (TMB) substrate (Kirkegaard and Perry Laboratories, Gaithersburg, MD) was added, undiluted, and incubated for about 15 minutes. The reaction was stopped with the addition of 100 μL of 1 N H<sub>2</sub>SO<sub>4</sub> to each well, and the plates were read at 450 nm.

As shown in Figure 26, approximately 50% of the samples from human leishmaniasis patients showed reactivities with recombinant MAPS-1A substantially above background. Figure 27 shows the reactivity of MAPS-1A with increasing

dilutions of sera from BALB/c mice previously administered either (i) saline solution; (ii) the adjuvant B. pertussis; (iii) soluble Leishmania antigens plus B. pertussis; (iv) live L. major promastigotes; or (v) soluble Leishmania antigens plus B. pertussis followed by live L. major promastigotes (as described below in Example 12). Considerably higher absorbances were seen with sera from mice infected with live L. major promastigotes and with mice infected with live L. major promastigotes following immunization with soluble Leishmania antigens plus B. pertussis, than with sera from the other three groups of mice, indicating that anti-MAPS-1A antibody titers increase following Leishmania infection.

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#### **EXAMPLE 12**

USE OF LEISHMANIA ANTIGENS FOR VACCINATION AGAINST LEISHMANIA INFECTION

This example illustrates the effectiveness of *Leishmania* antigens in conferring protection against disease in the experimental murine leishmaniasis model system. For a discussion of the murine leishmaniasis model system see, for example, Reiner et al. Annu. Rev. Immunol., 13:151-77, 1995.

The effectiveness of (i) crude soluble Leishmania antigens, (ii) MAPS-1A, and (iii) a mixture of Ldp23, LbeIF4A and M15, as vaccines against Leishmania infection was determined as follows. BALB/c mice (5 per group) were immunized intra-peritoneally three times at biweekly intervals with either (i) 30 µg crude soluble Leishmania antigens, (ii)20 µg MAPS-1A or (iii) a mixture containing 10 µg each of LeIF, Ldp23 and M15, together with 100 µg of the adjuvant C. parvum. Two control groups were immunized with either saline or C. parvum alone. Two weeks after the last immunization, the mice were challenged with 2 x 10<sup>5</sup> late-log phase promastigotes of L. major. Infection was monitored weekly by measurement of footpad swelling. The amount of footpad swelling seen in mice immunized with either crude soluble Leishmania antigens, a mixture of Ldp23, LbeiF4A and M15 (Figure 28), or MAPS-1A (Figure 29) was significantly less than that seen in mice immunized with C. parvum

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alone. These results demonstrate that the *Leishmania* antigens of the present invention are effective in conferring protection against *Leishmania* infection.

#### EXAMPLE 13

# ISOLATION OF DNA ENCODING FOR SOLUBLE ANTIGENS FROM AN L. MAJOR $GENOMIC \ DNA \ LIBRARY$

This example illustrates the isolation of seven soluble *Leishmania* antigen genes from an *L. major* genomic DNA library.

An L. major genomic DNA expression library was prepared from L. major promastigotes using the unidirectional Lambda ZAP (uni-ZAP) kit (Stratagene) according to the manufacturer's protocol. This library was screened with a high titer rabbit sera raised against L. major soluble antigens, as described above in Example 9. Seven positive clones were identified. The phagemid were excised and DNA from each of the seven clones was sequenced using a Perkin Elmer/Applied Biosystems. Division automated sequencer Model 373A. The DNA sequences for these antigens, referred to as LmgSP1, LmgSP3, LmgSP5, LmgSP8, LmgSP9, LmgSP13, LmgSP19, are provided in SEQ ID NO:29-35, respectively, with the corresponding amino acid sequences being provided in SEQ ID NO: 36-42, respectively. LmgSP13 was found to contain a 39 amino acid repeat sequence shown in SEQ ID NO:43.

Subsequent studies resulted in the isolation of a full-length sequence for LmgSP9. The full-length DNA sequence is provided in SEQ ID NO: 54, with the corresponding predicted amino acid sequence being provided in SEQ ID NO: 55. The amino acid sequence was found to contain six 14 amino acid repeat units (SEQ ID NO: 56), with each unit being further divided into two 7 amino acid units, provided in SEQ ID NO: 57 and 58.

Comparison of the DNA and amino acid sequences for the isolated antigens as described above, revealed no significant homologies to LmgSP1, LmgSP3, and LmgSP13. LmgSP5 was found to be related to the known PSA2 family. LmgSP8 was found to bear some homology to a sequence previously identified in *E. coli* (2-

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succinyl-6-hydroxy-2,4-cyclohexadiene-1-carboxylic acid synthase). LmgSP9 and LmgSP19 were found to be homologous to a *L. major* hydrophilic surface protein referred to as Gene B (Flinn, H.M. et al. *Mol. Biochem. Parasit.* 65:259-270, 1994), and to ubiquitin, respectively. To the best of the inventors' knowledge, none of these antigens have been previously shown to elicit T or B cell responses.

The reactivity of recombinant LmgSP9 with sera from patients with visceral leishmaniasis, (from both Sudan and Brazil) and from normal donors was evaluated by ELISA as described above. The absorbance values were compared with those obtained using the known *Leishmania* antigen K39 described above, with *L. chagasi* lysate being employed as a positive control. Representative results of these assays are provided below in Table 2, wherein all the patients from Brazil and those from the Sudan designated as "VL" were inflicted with visceral leishmaniasis. The results demonstrated that LmgSP9 specifically detects antibody in most individuals with visceral leishmaniasis, regardless of geographical location. In several cases, the absorbance values of the antibody reactivity to LmgSP9 were comparable to that observed with K39. In addition, LmgSP9 detected several cases of leishmaniasis that were not detected using K39. These results indicate that LmgSP9 can be used to complement the reactivity of K39.

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Table 2

REACTIVITY OF LMGSP9 WITH SERA FROM LEISHMANIA PATIENTS

Patient No.	L. chagasi lysate	K39	LmgSP9
Sudanese samples:			
B19	1.067	0.306	0.554
B25	1.884	3.435	0.974
B43	1.19	3.225	0.86
B47	2.405	2.892	0.375
B50	0.834	0.748	0.432
B58	0.921	0.235	0.92
B63	1.291	0.303	0.764
B70	0.317	0.089	3.056
· · VL4	1.384	3.035	2.965
VL11	0.382	0.144	0.142
VL12	0.277	0.068	0.098
VL13	0.284	0.12	0.194
Brazilian samples:			· .
105	3.508	3.53	0.374
106	2.979	3.373	2.292
107	2.535	3.444	0.46
109	1.661	3.415	3.319
111	3.595	3.537	0.781
112	2.052	3.469	0.63
113	3.352	3.429	0.963
114	2.316	3.437	1.058
115	2.073	3.502	1.186
116	3.331	3.461	0.96
Normal Donors:		·	
129	0.157	0.104	0.08
130	0.195	0.076	0.095
131	0.254	0.134	0.086
132	0.102	0.035	0.043

In order to obtain a higher specificity for the detection of antibodies in sera from visceral leishmaniasis patients, a homologue of LmgSP9 was isolated from *L. chagasi*, one of the causative agents of visceral leishmaniasis. A total of 80,000 pfu of an amplified *L. chagasi* genomic library were screened with the entire coding region of LmgSP9 (amplified from *L. major* genomic DNA). Seven hybridizing clones were purified to homogeneity. The determined DNA sequences for two of these clones, referred to as Lc Gene A and LcGene B, are provided in SEQ ID NO: 59 and 60, respectively, with the corresponding predicted amino acid sequences being provided in SEQ ID NO: 61 and 62, respectively. The open reading frame for Lc Gene A was found to show some homology to Gene A/C, previously isolated from *L. major* (McKlean et al., *Mol. Bio. Parasitol.*, 85:221-231, 1997). The open reading frame for Lc Gene B showed some homology to Gene B of *L. major*, discussed above, and was found to contain eleven repeats of a 14 amino acid repeat unit (SEQ ID NO: 63), with each repeat being further divided into two 7 amino acid units, provided in SEQ ID NO: 64 and 65.

The diagnostic potentials of Lc Gene A and Lc Gene B were evaluated by ELISA as described above using sera from visceral leishmaniasis patients from Sudan and Brazil, and from uninfected controls. Absorbance values were compared to those obtained using LmgSP9. Much higher absorbance values were obtained with Lc Gene A and Lc Gene B than with LmgSP9, with Lc Gene B appearing to be more effective that Lc Gene A in detecting antibodies in certain cases. These results indicate that Lc Gene B is highly effective in the diagnosis of visceral leishmaniasis.

In order to assess the diagnostic potential of the repeats found within Lc Gene B, a series of 6 peptides were synthesized (SEQ ID NO: 66-71; referred to as Pep 1-6), differing in an R or H residue. An ELISA was carried out using the full-length LcGene B protein and the six peptides. The absorbance values obtained with Pep 3 were higher than those obtained with the other 5 peptides, however they were not as high as those obtained with the full length protein.

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### **EXAMPLE 14**

ISOLATION AND CHARACTERIZATION OF DNA ENCODING FOR SOLUBLE ANTIGENS FROM
AN L. CHAGASI GENOMIC DNA LIBRARY

This example illustrates the preparation of five soluble *Leishmania* antigen genes from an *L. chagasi* genomic DNA library.

An L. chagasi genomic DNA expression library was prepared from L. chagasi promastigotes using the unidirectional Lambda ZAP (uni-ZAP) kit (Stratagene) according to the manufacturer's protocol. This library was screened with a high titer rabbit sera raised against L. major soluble antigens, as described above in Example 9. Five positive clones were identified. The phagemid were excised and DNA from each of the Five clones was sequenced using a Perkin Elmer/Applied Biosystems Division automated sequencer Model 373A. The DNA sequences for these antigens, referred to as LcgSP1, LcgSP3, LcgSP4, LcgSP8, and LcgSP10 are provided in SEQ ID NO:44-48, respectively, with the corresponding amino acid sequences being provided in SEQ ID NO:49-53, respectively.

Comparison of these sequences with known sequences in the gene bank as described above, revealed no known homologies to LcgSP3, LcgSP4, LcgSP8 and LcgSP10. LcgSP1 was found to be homologous to the known antigen HSP70.

Figures 30A and B illustrate the proliferative response of murine lymph nodes to recombinant LcgSP8, LcgSP10 and LcgSP3. Lymph nodes were taken BALB/c mice 17 days after infection with *L. major*. Infection occurred by footpad injection of 2 x 10<sup>6</sup> parasites/footpad. The cells were stimulated with recombinant antigen and proliferation was measured at 72 hours using <sup>3</sup>H-thymidine. Figure 30A shows the CPM, a direct measurement of mitotic activity in response to the antigens, and Figure 30B shows the stimulation index, which measures the proliferative response relative to the negative control.

#### **EXAMPLE 15**

# ISOLATION OF DNA ENCODING FOR *L. MAJOR* ANTIGENS BY CD4+ T CELL EXPRESSION CLONING

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This example illustrates the isolation of T cell antigens of L. major using a direct T cell screening approach.

Leishmania-specific CD4+ T cell lines were derived from the PBMC of an individual who tested positive in a leishmania skin test but had no clinical history of disease. These T cell lines were used to screen a *L. major* amastigote cDNA expression library prepared as described in Example 1. Immunoreactive clones were isolated and sequenced as described above. The determined cDNA sequences for the 8 isolated clones referred to as 1G6-34, 1E6-44, 4A5-63, 1B11-39, 2A10-37, 4G2-83, 4H6-41, 8G3-100 are provided in SEQ ID NO: 72-79, respectively, with the corresponding predicted amino acid sequences being provided in SEQ ID NO: 80-87, respectively. The cDNA sequences provided for 1E6-44, 2A10-37, 4G2-83, 4H6-41 and 8G3-100 are believe to represent partial clones. All of these clones were shown to stimulate T cell proliferation.

20 above revealed no known homologies to the antigen 4A5-63. 1G6-34 was found to have some homology to histone H2B previously identified in *L. enrietti*. Antigens 1E6-44, 1B11-39 and 8G3-100 showed some homology to sequences previously identified in other eukaryotes, in particular Saccharomyces cerevisae. 2A10-37 and 4H6-41 were found to be homologous to the two previously identified proteins alpha tubulin from *L. donovani* and beta tubulin from *L. major*, respectively, and 4G2-83 was found to be homologous to elongation initiation factor 2 previously identified in *T. cruzi*.

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# EXAMPLE 15 SYNTHESIS OF POLYPEPTIDES

Polypeptides may be synthesized on a Perkin Elmer/Applied Biosystems Division 430A peptide synthesizer using FMOC chemistry with HPTU (O-Benzotriazole-N,N,N',N'-tetramethyluronium hexafluorophosphate) activation. A Gly-Cys-Gly sequence may be attached to the amino terminus of the peptide to provide a method of conjugation, binding to an immobilized surface, or labeling of the peptide. Cleavage of the peptides from the solid support may be carried out using the following acid:ethanedithiol:thioanisole:water:phenol cleavage mixture: trifluoroacetic (40:1:2:2:3). After cleaving for 2 hours, the peptides may be precipitated in cold methyl-t-butyl-ether. The peptide pellets may then be dissolved in water containing 0.1% trifluoroacetic acid (TFA) and lyophilized prior to purification by C18 reverse phase HPLC. A gradient of 0%-60% acetonitrile (containing 0.1% TFA) in water (containing 0.1% TFA) may be used to elute the peptides. Following lyophilization of 15 the pure fractions, the peptides may be characterized using electrospray or other types of mass spectrometry and by amino acid analysis.

From the foregoing, it will be appreciated that, although specific embodiments of the invention have been described herein for the purpose of illustration, various modifications may be made without deviating from the spirit and scope of the invention.

#### SEQUENCE LISTING

- (1) GENERAL INFORMATION:
  - (i) APPLICANT: Corixa Corporation
- (ii) TITLE OF INVENTION: LEISHMANIA ANTIGENS FOR USEIN THE THERAPY AND DIAGNOSIS OF LEISHMANIASIS
  - (iii) NUMBER OF SEQUENCES: 87
  - (iv) CORRESPONDENCE ADDRESS:
    - (A) ADDRESSEE: SEED and BERRY LLP
    - (B) STREET: 6300 Columbia Center, 701 Fifth Avenue
    - (C) CITY: Seattle
      - (D) STATE: Washington
      - (E) COUNTRY: USA
      - (F) ZIP: 98104-7092
    - (v) COMPUTER READABLE FORM:
      - (A) MEDIUM TYPE: Floppy disk
      - (B) COMPUTER: IBM PC compatible
      - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
      - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
  - (vi) CURRENT APPLICATION DATA:
    - (A) APPLICATION NUMBER: US
    - (B) FILING DATE: 12-FEB-1998
    - (C) CLASSIFICATION:
  - (viii) ATTORNEY/AGENT INFORMATION:
    - (A) NAME: Maki, David J.
    - (B) REGISTRATION NUMBER: 31,392
    - (C) REFERENCE/DOCKET NUMBER: 210121.42001PC
    - (ix) TELECOMMUNICATION INFORMATION:
      - (A) TELEPHONE: (206) 622-4900
      - (B) TELEFAX: (206) 682-6031
- (2) INFORMATION FOR SEQ ID NO:1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 3134 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: 421..2058

# (xi) SEQUENCE DESCRIPTION: SkEQ ID NO:1:

CAAGTGTCGA AGGACAGTGT TCNCCGTGTG AGATCGCCGG CTGTGCGTGT GAAGGCGGTG	60
CCATCGGANA AACAACACCG GTGGANCCGC AGGAAACCAT CTTTCTCCGC AGGTCTCTTT	120
TTGTTGTCGA TTGAGAGTGC NCCAAACCCT GCTGGTGCCC TTCTCACATA TCATGTTTTT	180
CGTTGTGCGC TCGCTTTGCC TTTCCTCTCC TTTCCCTCTC TTCCGTGGTG CCGTGTATAC	240
TTCTGGCACC CGCTACGTCA CTTCGCTGGT TTGAACAGAA CCACTGTGAA CACCCACGGG	300
CGATCGCACA CATACACATC CCTCACTCAC ACACACAGCT ACATCTATCC TACATAAAGC	360
TGAAAAAAA GTCTACGAAC AATTTTGTTT TTACAGTGCG TTGCCGCACA TTTCTCCGTA	420
ATG GAC GCA ACT GAG CTG AAG AAC AAG GGG AAC GAA GAG TTC TCC GCC Met Asp Ala Thr Glu Leu Lys Asn Lys Gly Asn Glu Glu Phe Ser Ala 1 5 10 15	468
GGC CGC TAT GTG GAG GCG GTG AAC TAC TTC TCA AAG GCG ATC CAG TTG Gly Arg Tyr Val Glu Ala Val Asn Tyr Phe Ser Lys Ala Ile Gln Leu 20 25 30	516
GAT GAG CAG AAC AGT GTC CTC TAC AGC AAC CGC TCC GCC TGT TTT GCA Asp Glu Gln Asn Ser Val Leu Tyr Ser Asn Arg Ser Ala Cys Phe Ala 35 40 45	564
GCC ATG CAG AAA TAC AAG GAC GCG CTG GAC GAC GCC GAC AAG TGC ATC Ala Met Gln Lys Tyr Lys Asp Ala Leu Asp Asp Ala Asp Lys Cys Ile 50 55 60	612
TCG ATC AAG CCG AAT TGG GCC AAG GGC TAC GTG CGC CGA GGA GCA GCT Ser Ile Lys Pro Asn Trp Ala Lys Gly Tyr Val Arg Arg Gly Ala Ala 65 70 75 80	660
CTC CAT GGC ATG CGC CGC TAC GAC GAT GCC ATT GCC GCG TAT GAA AAG Leu His Gly Met Arg Arg Tyr Asp Asp Ala Ile Ala Ala Tyr Glu Lys 85 90 95	708
GGG CTC AAG GTG GAC CCT TCC AAC AGC GGC TGC GCG CAG GGC GTG AAG Gly Leu Lys Val Asp Pro Ser Asn Ser Gly Cys Ala Gln Gly Val Lys 100 105 110	756
GAC GTG CAG GTA GCC AAG GCC CGC GAA GCA CGT GAC CCC ATC GCT CGC Asp Val Gln Val Ala Lys Ala Arg Glu Ala Arg Asp Pro Ile Ala Arg 115 120 125	804
GTC TTC ACC CCG GAG GCG TTC CGC AAG ATC CAA GAG AAT CCC AAG CTG Val Phe Thr Pro Glu Ala Phe Arg Lys Ile Gln Glu Asn Pro Lys Leu 130 135 140	852
TCT CTA CTT ATG CTG CAG CCG GAC TAC GTG AAG ATG GTA GAC ACC GTC Ser Leu Leu Met Leu Gln Pro Asp Tyr Val Lys Met Val Asp Thr Val	900

									0,							
145					150					155					160	
						GGC Gly										948
						CTG Leu										996
						CGT Arg										1044
						CCT Pro 215										1092
						AAC Asn										1140
						CAA Gln										1188
						GTG Val										1236
						GAG Glu										1284
						ACA Thr 295										1332
						CAG Gln										1380
						GAG Glu									Lys	1428
						GAG Glu										1476
						AAG Lys										1524
TTC	AAG	GAG	GAT	AAG	TTC	CCC	GAG	GCC	GTG	GCA	GCG	TAC	ACG	GAG	GCC	1572

Phe	Lys 370	Glu	Asp	Lys	Phe	Pro 375	Glu	Ala	Val	Ala	Ala 380	Tyr	Thr	Glu	Ala	
												AAT Asn				1620
												AAG Lys				1668
AAG Lys	TGC Cys	ATT Ile	GAG Glu 420	CTG Leu	AAG Lys	CCC Pro	GAC Asp	TTT Phe 425	GTT Val	AAG Lys	GGC Gly	TAC Tyr	GCG Ala 430	CGC Arg	AAG Lys	1716
GGT Gly	CAT His	GCT Ala 435	TAC Tyr	TTT Phe	TGG Trp	ACC Thr	AAG Lys 440	CAG Gln	TAC Tyr	AAC Asn	CGC Arg	GCG Ala 445	CTG Leu	CAG Gln	GCG Ala	1764
TAC	GAT Asp 450	GAG Glu	GGC Gly	CTC Leu	AAG Lys	GTG Val 455	GAC Asp	CCG Pro	AGC Ser	AAT Asn	GCG Ala 460	GAC Asp	TGC Cys	AAG Lys	GAT Asp	1812
Gly	CGG Arg	TAT Tyr	CGC Arg	ACA Thr	ATC Ile 470	ATG Met	AAG Lys	ATT Ile	CAG Gln	GAG Glu 475	ATG Met	GCA Ala	TCT Ser	GGC Gly	CAA Gln 480	1860
		Asp		Asp						Ala		GAC Asp				1908
ATC Ile	GCG Ala	GCA Ala	ATC Ile 500	Met	CAA Gln	GAT Asp	AGC Ser	TAC Tyr 505	Met	CAA Gln	CTA Leu	GTG Val	TTG Leu 510	Lys	GAG Glu	1956
ATC Met	G CAG	AAC Asr 515	a Asp	CCC Pro	ACG Thr	Arg	ATT Ile	Gln	GAG Glu	TAC	ATG Met	AAG Lys 525	Asp	TCC Ser	GGG Gly	2004
ATO	C TCA Ser 530	Sei	AAC Lys	ATC	AAC Asn	AAG Lys	Lev	ATI	TCA Ser	GCT Ala	GGC Gly 540	/ Ile	ATT	CGT Arg	TTT	2052
	y Gli	ı			CGCT		CA 1	CTT	TCCC			rgcgi				2108
CG'	AAAT	GCAC	AATA	AAGG	CAG C	GATT	CAC	AT GO	CACG	AGTA	A AG	rgcto	GCGC	CTCI	CAAACA	2168
CG.	ACGT(	CGAG	GCT	GTGG:	rgc 1	AGATO	GCGC	GT C	CTGC	ATGA	A GG	ragt(	BAAG	AGG	AAGTAA	2228
GG	GATG'	TTGT	TTG	rggg(	CCT 7	CGT	GGCT	GC G	CACA	CACC'	r ct	TATC'	TCCT	TCG	CTTGGTA	2288
CC	TTÇT	CCCT	TTT'	rcgT(	CTT (	CACC	CCCC'	TT T	CTCT'	TCTC	A CG	CTCT	CCCT	GGC	CCGCTGG	2348

TGCAACGATT	TCGTTTTATT	TACGTCTGTG	TAGCTCCTCT	ATTCAACGGT	GCGATGACGC	2408
TAACGAAGCT	GGCCTGTATT	CGGCTAAGGC	GAAGGCAAAA	GACTAGGAGG	GGGGGGGAA	2468
GGAGACGGCG	TGACCATCAC	TGCGAAGAAA	CAAGCCGAAG	AAAAGGCCCC	GAACGCCTGC	2528
ATTTCCGCGC	GCCCTCGCCC	GCCTTCCTTC	CTTCCTTCGC	TCTCTCTCTC	TCTCTCTCTC	2588
GCTATCTTCT	CAACGGAGAC	ATGAAAGGCG	TTTGTTAGGA	AAAGAGGGGG	GGGGGAAGAG	2648
TGGGACGACG	CGCTGCGTCT	TTTGGGCACT	GGTCACGTGC	GTCACCCTCT	TTTTTTATCT	2708
CTATTGGCAC	TGTCTTGTTT	CTTTTCCCTT	TCCTATCATA	CGCGTCTCGC	AAACGACTCC	2768
GCGCTGAGCA	GCCATGTGCT	GCGGCGTGGA	GGAAGTACAC	AGACATCACG	GATGCATATG	2828
TGCGCGTCCG	TGTACGCGCT	TGTATGGGGC	TTCTAACAGC	GCCTGTGTGT	GTTTGTGTGT	2888
GTGTGTGT	GTGTGTCTGT	GTATTTCGAG	CGTCTGTATG	CTATTCTATT	AAGCACCGAA	2948
GAAGAGACAC	ACACGACAGC	GAAGGAGATG	GTGTCGGCTT	TTCGGCTAAT	CACTCCCTTC	3008
CATAGCTTCT	CTGAAGGAGG	CTCTCTTCCA	GAGGAATAGA	CTGCAGATGG	GGTCCACGTT	3068
TATCTGAGGA	GTCAACGGAA	АААААААА	АААААААА	AAAAAAAA	AAAAAAAA	3128
CTCGAG					•	3134

### (2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 546 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Asp Ala Thr Glu Leu Lys Asn Lys Gly Asn Glu Glu Phe Ser Ala 1 5 10 15

Gly Arg Tyr Val Glu Ala Val Asn Tyr Phe Ser Lys Ala Ile Gln Leu 20 25 30

Asp Glu Gln Asn Ser Val Leu Tyr Ser Asn Arg Ser Ala Cys Phe Ala 35 40 45

Ala Met Gln Lys Tyr Lys Asp Ala Leu Asp Asp Ala Asp Lys Cys Ile
50 55 60

Ser Ile Lys Pro Asn Trp Ala Lys Gly Tyr Val Arg Arg Gly Ala Ala 65 70 75 80

70 Leu His Gly Met Arg Arg Tyr Asp Asp Ala Ile Ala Ala Tyr Glu Lys 90 Gly Leu Lys Val Asp Pro Ser Asn Ser Gly Cys Ala Gln Gly Val Lys 105 Asp Val Gln Val Ala Lys Ala Arg Glu Ala Arg Asp Pro Ile Ala Arg 125 120 115 Val Phe Thr Pro Glu Ala Phe Arg Lys Ile Gln Glu Asn Pro Lys Leu 135 Ser Leu Leu Met Leu Gln Pro Asp Tyr Val Lys Met Val Asp Thr Val 155 150 Ile Arg Asp Pro Ser Gln Gly Arg Leu Tyr Met Glu Asp Gln Arg Phe 165 Ala Leu Thr Leu Met Tyr Leu Ser Gly Met Lys Ile Pro Asn Asp Gly 180 Asp Gly Glu Glu Glu Arg Pro Ser Ala Lys Ala Ala Glu Thr Ala 200 Lys Pro Lys Glu Glu Lys Pro Leu Thr Asp Asn Glu Lys Glu Ala Leu 220 215 Ala Leu Lys Glu Glu Gly Asn Lys Leu Tyr Leu Ser Lys Lys Phe Glu 235 230 225 Glu Ala Leu Thr Lys Tyr Gln Glu Ala Gln Val Lys Asp Pro Asn Asn 250 245 Thr Leu Tyr Ile Leu Asn Val Ser Ala Val Tyr Phe Glu Gln Gly Asp 265 Tyr Asp Lys Cys Ile Ala Glu Cys Glu His Gly Ile Glu His Gly Arg 280 275 Glu Asn His Cys Asp Tyr Thr Ile Ile Ala Lys Leu Met Thr Arg Asn 295 290 Ala Leu Cys Leu Gln Arg Gln Arg Lys Tyr Glu Ala Ala Ile Asp Leu 315 Tyr Lys Arg Ala Leu Val Glu Trp Arg Asn Pro Asp Thr Leu Lys Lys 330 Leu Thr Glu Cys Glu Lys Glu His Gln Lys Ala Val Glu Glu Ala Tyr 340 345

Ile Asp Pro Glu Ile Ala Lys Gln Lys Lys Asp Glu Gly Asn Gln Tyr 360

Phe Lys Glu Asp Lys Phe Pro Glu Ala Val Ala Ala Tyr Thr Glu Ala

355

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370

375

380

Ile Lys Arg Asn Pro Ala Glu His Thr Ser Tyr Ser Asn Arg Ala Ala 385 390 395 400

Ala Tyr Ile Lys Leu Gly Ala Phe Asn Asp Ala Leu Lys Asp Ala Glu
405 410 415

Lys Cys Ile Glu Leu Lys Pro Asp Phe Val Lys Gly Tyr Ala Arg Lys
420 425 430

Gly His Ala Tyr Phe Trp Thr Lys Gln Tyr Asn Arg Ala Leu Gln Ala 435 440 445

Tyr Asp Glu Gly Leu Lys Val Asp Pro Ser Asn Ala Asp Cys Lys Asp 450 455 460

Gly Arg Tyr Arg Thr Ile Met Lys Ile Gln Glu Met Ala Ser Gly Gln 465 470 475 480

Ser Ala Asp Gly Asp Glu Ala Ala Arg Arg Ala Met Asp Asp Pro Glu 485 490 495

Ile Ala Ala Ile Met Gln Asp Ser Tyr Met Gln Leu Val Leu Lys Glu
500 505 510

Met Gln Asn Asp Pro Thr Arg Ile Gln Glu Tyr Met Lys Asp Ser Gly 515 520 525

Ile Ser Ser Lys Ile Asn Lys Leu Ile Ser Ala Gly Ile Ile Arg Phe
530 535 540

Gly Gln 545

#### (2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 676 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

#### (ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 26..550
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

AATTCGGCAC GAGGCATTGT GCATA ATG GTC AAG TCC CAC TAC ATC TGC GCG Met Val Lys Ser His Tyr Ile Cys Ala

	CGC																100
Gly	Arg	Leu	Val	<b>Arg 560</b>	Ile	Leu	Arg	GIY.	Pro 565	Arg	Gin	Asp	Arg .	570	GIÀ		
GTG	ATC	GTC	GAC	ATT	GTC	GAC	GCG	AAC	CGC	GTG	CTG	GTG	GAG	AAC	CCG		148
Val	Ile	Val	Asp 575	Ile	Val	Asp	Ala	Asn 580	Arg	Val	Leu	Val	Glu 585	Asn	Pro		
	GAC																196
Glu	Asp	Ala 590	Lys	Met	Trp	Arg	His 595	Val	Gln	Asn	Leu	Lys 600	Asn	Val	Glu		
															GCG	(	244
Pro	Leu 605	Lys	Tyr	Суѕ	Val	Ser 610	Val	Ser	Arg	Asn	Cys 615	Ser	Ala	Lys	Ala		e*
CTG	AAG	GAT	GCG	CTG	GCC	TCG	TCG	AAG	GCG	CTG	GAG	AAG	TAC	GCG	AAG		292
	Lys	Asp	Ala	Leu		Ser	Ser	Lys	Ala		Glu	Lys	Tyr	Ala			
620					625					630					635		
ACG	CGC	ACT	GCT	GCG	CGC	GTG	GAG	GCG	AAG	AAG	GCG	TGC	GCC	GCG	TCG		340
Thr	Arg	Thr	Ala		Arg	Val	Glu	Ala		Lys	Ala	Cys	Ala		Ser		
				640					645					650			•
ACG	GAC	TTC	GAG	CGC	TAC	CAG	CTG	CGC	GTT	GCG	CGC	CGT	TCT	CGC	GCG		388
Thr	Asp	Phe		Arg	Tyr	Gln	Leu		Val	Ala	Arg	Arg		Arg	Ala	•	
			655					660					665				
CAC	TGG	GCĢ	CGC	AAG	GTG	TTC	GAC	GAG	AAG	GAC	GCG	AAG	ACG	CCC	GTG		436
His	Trp			Lys	Val	Phe		Glu	Lys	Asp	Ala		Thr	Pro	Val		
		670					675					680					
TCG	TGG	CAC	AAG	GTT	GCG	CTG	AAG	AAG	ATG	CAG	AAG	AAG	GCC	GCA	AAG		484
Ser	Ţrp		Lys	Val	Ala			Lys	Met	Gln			Ala	Ala	Lys		
	685					690		•			695						
ATG	GAC	TCG	ACC	GAG	GGC	GCT	AAG	AGG	CGC	ATG	CAG	AAG	GCG	ATC	GCT		532
	Asp									Met	Gln				Ala		
700	)				705					710					715		
GCC	CGC	. AAG	GCG	AAA :	AAG	TAA	GGCC	ATA	CCCI	'CACT	TC G	CTTG	TTTC	G			580
	Arg				Lys												
TGA	TTTT	TCG	TGGG	SAGTO	GG T	GGCC	CTAC	CC AG	CGGT	CTTT	CAT	TGGC	ATT	TTTC	TATC	CG	640
GTO	TGA	AGA	GGT	CAA	AA A	AAAA	AAA	AA AA	AAAA	1							676

### (2) INFORMATION FOR SEQ ID NO:4:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 175 amino acids
  - (B) TYPE: amino acid

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Val Lys Ser His Tyr Ile Cys Ala Gly Arg Leu Val Arg Ile Leu

1 5 10 15

Arg Gly Pro Arg Gln Asp Arg Val Gly Val Ile Val Asp Ile Val Asp 20 25 30

Ala Asn Arg Val Leu Val Glu Asn Pro Glu Asp Ala Lys Met Trp Arg
35 40 45

His Val Gln Asn Leu Lys Asn Val Glu Pro Leu Lys Tyr Cys Val Ser 50 55 60

Val Ser Arg Asn Cys Ser Ala Lys Ala Leu Lys Asp Ala Leu Ala Ser 65 70 75 80

Ser Lys Ala Leu Glu Lys Tyr Ala Lys Thr Arg Thr Ala Ala Arg Val 85 90 95

Glu Ala Lys Lys Ala Cys Ala Ala Ser Thr Asp Phe Glu Arg Tyr Gln
100 105 110

Leu Arg Val Ala Arg Arg Ser Arg Ala His Trp Ala Arg Lys Val Phe
115 120 125

Asp Glu Lys Asp Ala Lys Thr Pro Val Ser Trp His Lys Val Ala Leu 130 135 140

Lys Lys Met Gln Lys Lys Ala Ala Lys Met Asp Ser Thr Glu Gly Ala 145 150 155 160

Lys Arg Arg Met Gln Lys Ala Ile Ala Ala Arg Lys Ala Lys Lys 165 170 175

- (2) INFORMATION FOR SEQ ID NO:5:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 2040 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: 62..2029
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CGCG	GTGG	CG G	CCGC	CTA	G AA	CTAGʻ	TGGA	TCC	CCCG	GGC	TGCA	GGAA'	TT C	GGCA	CGAGA		60
G AG Se					o Al					u Gl		T CA r Hi			g		106
			Val 1									ACG Thr	Val				154
		Ile					Ala					AAT Asn					202
ATC	GCG	CGC	TCC	GGC	ACG	AAG	GCT	TTC	ATG	GAG	GCA	CTG	GAG	GCC Ala	GĞC G1 v		250
Ile	Ala	225	ser	GIÀ	THE		230	Pne	Mec	GIU	ALG	Leu 235	GIU	A.u.	O±1		
												TTC Phe					298
TAC Tyr 255	CTT Leu	GTG Val	GCG Ala	GAC Asp	CGC Arg 260	GTG Val	ACG Thr	GTG Val	GTG Val	TCG Ser 265	AAG Lys	AAC Asn	AAC Asn	TCG Ser	GAC Asp 270	~	346
GAG Glu	GCG Ala	Tyr	TGG	Glu	TCG Ser	TCT Ser	GCG Ala	GGG Gly	GGC Gly 280	ACG Thr	TTC Phe	ACC Thr	ATC Ile	ACG Thr 285	AGC Ser		394
GTG Val	CAG Gln	GAG Glu	TCG Ser 290	GAC Asp	ATG Met	AAG Lys	CGC Arg	GGC Gly 295	ACG Thr	AGT Ser	ACA Thr	ACG Thr	CTG Leu 300	CAC His	CTA Leu		442
												GTG Val 315					490
		Lys										GAG Glu					538
GAG Glu 335	Lys	ACG Thr	GCG Ala	GAG Glu	AAG Lys 340	Glu	GTG Val	ACG Thr	GAC Asp	GAG Glu 345	Asp	GAG Glu	GAG Glu	GAG Glu	GAC Asp 350		586
GA0 Gli	TCG Ser	AAG Lys	AAG Lys	AAG Lys 355	Ser	TGC Cys	GGG Gly	GAC Asp	GAG Glu 360	Gly	GAG Glu	CCG Pro	AAG Lys	GTG Val 365	GAG Glu		634
				Gly					Lys					Lys	GTG Val		682

				, ,					
		_				CAC His 395			730
						TAC Tyr			778
						GCG Ala			826
						GCG Ala			874
						AAG Lys			922
						AAC Asn 475			970
						GTG Val			1018
						CAG Gln			1066
						CTG Leu			1114
						TTC Phe			1162
						ACG Thr 555			1210
						GAG Glu			1258
						AAG Lys			1306
						AAG Lys			1354

				GCG Ala						1402
				GAG Glu						1450
				TGC Cys 645						1498
				AAG Lys						1546
				ATG Met						1594
	 			CGC Arg						1642
				TCG Ser						1690
	,			AGC Ser 725						1738
	 			GAC Asp						1786
				AAC Asn						1834
				CTG						1882
		Tyr		CGC Arg	Asn				·	1930
	Asp			GAG Glu 805			Pro			1978
				ACC Thr						2026

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2040

815 820 825 830

GAC TGAGCCGGTA A
Asp

#### (2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 656 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Ser Leu Thr Asp Pro Ala Val Leu Gly Glu Glu Thr His Leu Arg Val

1 1 15

Arg Val Val Pro Asp Lys Ala Asn Lys Thr Leu Thr Val Glu Asp Asn 20 25 30

Gly Ile Gly Met Thr Lys Ala Asp Leu Val Asn Asn Leu Gly Thr Ile 35 40 45

Ala Arg Ser Gly Thr Lys Ala Phe Met Glu Ala Leu Glu Ala Gly Gly 50 55 60

Asp Met Ser Met Ile Gly Gln Phe Gly Val Gly Phe Tyr Ser Ala Tyr 65 70 75 80

Leu Val Ala Asp Arg Val Thr Val Val Ser Lys Asn Asn Ser Asp Glu
85 90 95

Ala Tyr Trp Glu Ser Ser Ala Gly Gly Thr Phe Thr Ile Thr Ser Val 100 105 110

Gln Glu Ser Asp Met Lys Arg Gly Thr Ser Thr Thr Leu His Leu Lys 115 120 125

Glu Asp Gln Gln Glu Tyr Leu Glu Glu Arg Arg Val Lys Glu Leu Ile 130 135 140

Lys Lys His Ser Glu Phe Ile Gly Tyr Asp Ile Glu Leu Met Val Glu 145 150 155 160

Lys Thr Ala Glu Lys Glu Val Thr Asp Glu Asp Glu Glu Glu Asp Glu 165 170 175

Ser Lys Lys Ser Cys Gly Asp Glu Gly Glu Pro Lys Val Glu Glu
180 185 190

Val Thr Glu Gly Glu Asp Lys Lys Lys Thr Lys Lys Val Lys

									10						
		195					200					205			
Glu	Val 210	Lys	Lys	Thr		Glu 215	Val	Lys	Asn	Lys	His 220	Lys	Pro :	Leu	Trp
Thr 225	Arg	Asp	Thr	Lys	Asp 230	Val	Thr	Lys	Glu	Glu 235	Tyr	Ala	Ala	Phe	Tyr 240
Lys	Ala	Ile	Ser	Asn 245	Asp	Trp	Glu	Asp	Thr 250	Ala	Ala	Thr	Lys	His 255	Phe
Ser	Val	Glu	Gly 260	Gln	Leu	Glu	Phe	Arg 265	Ala	Ile	Ala	Phe	Val 270	Pro	Lys
Arg	Ala	Pro 275	Phe	Asp	Met	Phe	Glu 280	Pro	Asn	Lys	ГЛЗ	Arg 285	Asn	Asn	Ile
Lys	Leu 290		Val	Arg	Arg	Val 295	Phe	Ile	Met	Asp	Asn 300	Cys	Glu	Asp	Leu
Суs 305		Asp	Trp	Leu	Gly 310	Phe	Val	Lys	Gly	Val 315	Val	Asp	Ser	Glu	Asp 320
Leu	Pro	Leu	Asn	Ile 325	Ser	Arg	Glu	Asn	Leu 330	Gln	Gln	Asn	Lys	Ile 335	Leu
Lys	Val	Ile	Arg 340		Asn	Ile	Val	Lys 345		Cys	Leu	Glu	Leu 350	Phe	Glu
Glu	ılle	ala 355	Glu 5	Asn	Lys	Glu	Asp 360		Lys	Gln	Phe	Tyr 365	Glu	Gln	Phe
Gly	/ Lys		ı Ile	Lys	Leu	Gly 375		His	Glu	Asp	Thr 380	Ala	Asn	Arg	Lys
Lys 385		ı Met	c Glu	. Leu	Leu 390		Phe	Tyr	Ser	Thr 395		Ser	Gly	Glu	Glu 400
Met	: Thi	r Th	r Leu	Lys 405		Туг	. Val	Thr	Arg 410		. Lys	Pro	Glu	Gln 415	Lys
Se	r Il	e Ty	r Ty:		. Thr	Gly	/ Asp	Sei 425		E Lys	Lys	Leu	Glu 430	Ser	Ser
Pr	o Ph	e Il 43		ı Lys	s Ala	Arg	9 Arg		s Gly	y Lei	ı Glu	Val 445	. Leu	Phe	Met
Th	r Gl 45		o Il	e Ası	o Glu	1 Ty:		l Me	t Gli	n Gli	n Val 460	Lys	asp	Phe	Glu
As 46		в Ьу	s Ph	e Ala	a Cys		u Th:	r Ly	s Gl	u Gl;	y Val	l His	s Phe	e Glu	Glu 480
Se	r Gl	u Gl	u Gl	u Ly 48		s Gl	n Ar	g Gl	u Gl 49	и <b>L</b> y 0	s Lys	s Ala	a Ala	495	Glu

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Lys	Leu	Cys	Lys 500	Thr	Met	Lys	Glu	Val 505	Leu	Gly	Asp	Lys	Val 510	Glu	Lys	
Val	Thr	Val 515	Ser	Glu	Arg	Leu	Leu 520	Thr	Ser	Pro	Cys	Ile 525	Leu	Val	Thr	
Ser	Glu 530	Phe	Gly	Trp	Ser	Ala 535	His	Met	Glu	Gln	Ile 540	Met	Arg	Asn	Gln	
Ala 545	Leu	Arg	Asp	Ser	Ser 550	Met	Ala	Gln	Tyr	Met 555	Val	Ser	Lys	Lys	Thr 560	
Met	Glu	Val	Asn	Pro 565	Asp	His	Pro	Ile	Ile 570	Lys	Glu	Leu	Arg	Arg 575	Arg	
Val	Glu	Ala	Asp 580	Glu	Asn	Asp	Lys	Ala 585	Val	Lys	Asp	Leu	Val 590	Phe	Leu	
Leu	Phe	Asp 595	Thr	Ser	Leu	Leu	Thr 600	Ser	Gly	Phe	Gln	Leu 605	Asp	Asp	Pro	
Thr	Gly 610	Tyr	Ala	Glu	Arg	Ile 615	Asn	Arg	Met	Ile	Lys 620	Leu	Gly	Leu	Ser	
Leu 625	Asp	Ģlu	Glu	Glu	Glu 630	Glu	Val	Ala	Glu	Ala 635	Pro	Pro	Ala	Glu	Ala 640	
Ala	Pro	Ala	Glu	Val 645	Thr	Ala	Gly	Thr	Ser 650	Ser	Met	Glu	Gln	Val 655	Asp	
(2)	INFO	ORMA	NOI	FOR	SEQ	ID N	10 : 7 :	ŀ								
	(i)	( <i>I</i> (I	A) LE B) TY C) ST	ength (PE : [rani	HARAC H: 17 nucl DEDNI DGY:	771 l Leic ESS:	ase ació sino	pai:	cs							
	(ix)	(1		ME/I	ŒY: ION:		1698			× .						
-	(xi)	) SE(	QUENC	CE DI	ESCR!	PTIC	ON: 8	SEQ :	ID NO	):7:						
													CGC Arg			48
													CAT His 30			96

			GCC Ala						·	144
			GAG Glu 55							192
			CAG Gln							240
			GAG Glu							288
			TCC Ser					CTC Leu		336
	Leu		GCC Ala							384
			CGC Arg 135							432
	 		GAG Glu							480
			GCC Ala							528
			GAG Glu							576
			CAG Gln							624
			CAC His 215						. ·	672
			CTC Leu							720
			ATG							768

				245	i				250	)		•		255	5	
GCC Ala	CTC Leu	GAG Glu	GAG Glu 260	Ala	GCG	CGT Arg	CTC	CGC Arg 265	Ala	GAG Glu	CTC Leu	GAG Glu	GCG Ala 270	Ala	GAG Glu	816
GAG Glu	GCG Ala	GCC Ala 275	Arg	CTG Leu	GAT Asp	GTC Val	ATG Met 280	His	GAG Glu	GGC	GAG Glu	Gln 285	GCC	CGT	GTC Val	864
CAG Gln	GCC Ala 290	Leu	GAG Glu	GAG Glu	GCG Ala	GCC Ala 295	CGC Arg	CTG Leu	GAG Glu	GCC Ala	ATG Met 300	His	GAG Glu	GCC Ala	GAG Glu	. 912
CAG Gln 305	GCC Ala	CGC Arg	TCC Ser	CAG Gln	GCC Ala 310	CTC Leu	GAG Glu	GAG Glu	GCA Ala	GCG Ala 315	CGT Arg	CTC Leu	TGC Cys	GCG Ala	GAG Glu 320	960
CTG Leu	GAG Glu	GCT Ala	GAG Glu	GAG Glu 325	GAG Glu	GAA Glu	AAA Lys	GAT Asp	GAG Glu 330	CGG Arg	CCG Pro	GCG Ala	ACG Thr	TCG Ser 335	Ser	1008
TAC Tyr	AGC Ser	GAG Glu	GAG Glu 340	TGC Cys	AAA Lys	GGG Gly	CGA Arg	CTG Leu 345	CTA Leu	TCG Ser	AGG Arg	GCG Ala	CGG Arg 350	CCG Pro	GAT Asp	1056
CCG Pro	CGG Arg	AGG Arg 355	CCG Pro	CTG Leu	CCG Pro	CGG Arg	CCG Pro 360	TTC Phe	ATT Ile	GGG Gly	ATG Met	TCA Ser 365	CTG Leu	TTG Leu	GAG Glu	1104
GAT Asp	GTG Val 370	GAG Glu	AAG Lys	AGT Ser	ATT Ile	CTC Leu 375	ATT Ile	GTG Val	GAC Asp	GGG Gly	CTC Leu 380	TAC Tyr	AGG Arg	GAT Asp	GGG Gly	1152
CCG Pro 385	GCG Ala	TAC Tyr	CAG Gln	ACG Thr	GGC Gly 390	ATC Ile	CGC Arg	CTC Leu	GGG Gly	GAT Asp 395	GTC Val	CTC Leu	TTG Leu	CGT Arg	ATC Ile 400	1200
GCG Ala	GGG Gly	GTT Val	TAC Tyr	GTG Val 405	GAT Asp	TCA Ser	ATA Ile	GCG Ala	AAG Lys 410	GCG Ala	AGG Arg	CAG Gln	GTG Val	GTC Val 415	GAT Asp	1248
GCG Ala	CGT Arg	TGC Cys	CGC Arg 420	TGC Cys	GGC Gly	TGC Cys	GTC Val	GTT Val 425	CCC Pro	GTG Val	ACG Thr	CTG Leu	GCG Ala 430	ACG Thr	AAG Lys	1296
ATG Met	AAC Asn	CAG Gln 435	CAG Gln	TAC Tyr	AGC Ser	GTG Val	GCT Ala 440	CTG Leu	TAT Tyr	ATC Ile	ATG Met	ACG Thr 445	GTG Val	GAT Asp	CCG Pro	1344
CAG Gln	CAC His 450	AAC Asn	GAC Asp	AAG Lys	CCC Pro	TTT Phe 455	TTT Phe	TTT Phe	GAT Asp	GTG Val	CAC His 460	ATC Ile	CAC His	CAC His	CGC Arg	1392
ATC	GAG	AGC	TCG	CAC	ATG	GGG	AAG	AAG	GCG	CAG	TGG	ATG	GAA	GTT	CTT	1440

									82							
Ile 465	Glu	Ser	Ser	His	Met 470	Gly	Lys	Lys	Ala	Gln 475	Trp	Met	Glu	Val	Leu 480	
			TCC Ser													1488
			CCG Pro 500													1536
			TTC Phe													1584
			GAA Glu													1632
			GCG Ala													1680
			GGC Gly			TGA	CGTC'	rct (	GTGT(	SAGTO	et G	rgtco	GCTC(	2		1728
GTC	rcct'	rcc :	rttt:	rcgt(	CA TO	GTGT	TTTA'	r TC	ATTT(	CTTT	TTC					1771
(2)	INF		TION	FOR	SEQ	ID I	8 : 07	:								,
		(i) :	(B		NGTH PE:	: 56	6 am o ac	ino a id		S		,		٠.		
	(	<b>ii)</b> 1	MOLE	CULE	TYP	E: p	rote	in			٠					X.
	(	xi)	SEQU	ENCE	DES	CRIP'	TION	: SE	Q ID	NO:	8 :					
Gln 1	Ala	Arg	Val	Gln 5		Leu	Glu	Glu	Ala 10		Arg	Leu	Arg	Ala 15	Glu	
Leu	Glu	Ala	Ala 20		Glu	Ala	Ala	Arg 25		Asp	Val	Met	His 30	Ala	Ala	
Glu	Gln	Ala 35	Arg	Val	Gln	Ala	Leu 40		Glu	Ala	Ala	Arg 45		Arg	Ala	
Glu	Leu 50		Glu	Ala	Glu	Glu 55		Ala	Arg	Leu	Asp 60		Met	His	Ala	

Ala Glu Gln Ala Arg Val Gln Ala Leu Glu Glu Ala Ala Arg Leu Arg

Ala	Glu	Leu	Glu	Ala 85	Ala	Glu	Glu	Ala	Ala 90	Arg	Leu	Glu	Ala	Met 95	His
Glu	Ala	Glu	Gln 100	Ala	Arg	Ser	Gln	Ala 105	Leu	Glu	Glu	Ala	Ala 110	Arg	Leu
Arg	Ala	Glu 115	Leu	Glu	Glu	Ala	Glu 120	Glu	Ala	Ala	Arg	Leu 125	Asp	Val	Met
His	Ala 130	Ala	GÌu	Gln	Ala	Arg 135	Val	Gln	Ala	Leu	Glu 140	Glu	Ala	Ala	Arg
Leu 145	Arg	Ala	Glu	Leu	Glu 150	Glu	Ala	Glu	Glu	Ala 155	Ala	Àrg	Leu	Glu	Ala 160
Met	His	Glu	Ala	Glu 165	Gln	Ala	Arg	Ser	Gln 170	Ala	Leu	Glu	Glu	Ala 175	Ala
Arg	Leu	Arg	Ala 180	Glu	Leu	Glu	Ala	Ala 185	Glu	Glu	Ala	Ala	Arg 190	Leu	Asp
Val	Met	His 195	Glu	Ala	Glu	Gln	Ala 200	Arg	Val	Gln	Ala	Leu 205	Glu	Glu	Ala
Ala	Arg 210	Leu	Asp	Val	Met	His 215	Glu	Ala	Glu	Gln	Ala 220	Arg	Val	Gln	Ala
Leu 225	Glu	Glu	Ala	Ala	Arg 230	Leu	Arg	Ala	Glu	Leu 235	Glu	Ala	Ala	Glu	Glu 240
Ala	Ala	Arg	Leu	Asp 245	Val	Met	His	Glu	Ala 250	Glu	Gln	Ala	Arg	Val 255	Gln
Ala	Leu	Glu	Glu 260	Ala	Ala	Arg	Leu	Arg 265	Ala	Glu	Leu	Glu	Ala 270	Ala	Glu
Glu	Ala	Ala 275	Arg	Leu	Asp	Val	Met 280	His	Glu	Gly	Glu	Gln 285	Ala	Arg	Val
Gln	Ala 290	Leu	Glu	Glu	Ala	Ala 295	Arg	Leu	Glu	Ala	Met 300	His	Glu	Ala	Glu
Gln 305	Ala	Arg	Ser	Gln	Ala 310	Leu	Glu	Glu	Ala.	Ala 315	Arg	Leu	Сув	Ala	Glu 320
Leu	Glu	Ala	Glu	Glu 325	Glu	Glu	Lys	Asp	Glu 330	Arg	Pro	Ala	Thr	Ser 335	Ser
Tyr	Ser	Glu	Glu 340	Сув	Lys	Gly	Arg	Leu 345	Leu	Ser	Arg	Ala	Arg 350	Pro	Asp
Pro	Arg	Arg	Pro	Leu	Pro	Arg	Pro	Phe	Ile	Gly	Met	Ser	Leu	Leu	Glu

360

·365

Asp Val Glu Lys Ser Ile Leu Ile Val Asp Gly Leu Tyr Arg Asp Gly 370 375 380

Pro Ala Tyr Gln Thr Gly Ile Arg Leu Gly Asp Val Leu Leu Arg Ile 385 390 395 400

Ala Gly Val Tyr Val Asp Ser Ile Ala Lys Ala Arg Gln Val Val Asp
405 410 415

Ala Arg Cys Arg Cys Gly Cys Val Val Pro Val Thr Leu Ala Thr Lys
420 425 430

Met Asn Gln Gln Tyr Ser Val Ala Leu Tyr Ile Met Thr Val Asp Pro 435 440 445

Gln His Asn Asp Lys Pro Phe Phe Phe Asp Val His Ile His His Arg 450 455 460

Ile Glu Ser Ser His Met Gly Lys Lys Ala Gln Trp Met Glu Val Leu 465 470 475 480

Glu Ser Pro Ser Val Ser Ser Ala Ala Thr Thr Pro Leu Val Pro Leu 485 490 495

Leu Arg Glu Pro Thr Pro Arg Arg Gly Ser Glu Leu Gln Ser Ser Ala
500 505 510

Arg Ser Ala Phe Val Ala Thr Ser Tyr Phe Ser Ser Ala Arg Arg Ser 515 520 525

Val Ser Ser Glu Ser Glu Arg Pro Arg Gly Ser Ser Ser Val Ala Met 530 535 540

Ala Glu Glu Ala Ile Ala Leu Ala Pro Gln Gly Tyr Thr Pro Pro Asn 545 550 555 560

Gln Val Arg Gly Arg Ser 565

### (2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1618 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 115..1323

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

CCACTCTCTC GGTCGTCTGT	CTCCCACGCG CGCACGCAGT	T TGATTTCCGC CTTCTTAAAC 60
GCTCTCTTTT TTTTTATTTT	TCACCTGACC AACCGCACCA	A CGTCGGCCTC CATC ATG 117  Met 1
TCG CAG CAA GAC CGA GT Ser Gln Gln Asp Arg Va 5		
GAC CAG CCC GGC GTC CG Asp Gln Pro Gly Val Ar		,
CAC CAG AAC CTT\CTG CG His Gln Asn Leu Leu Ar		·
TCC AGC ATC CAG CAG CG Ser Ser Ile Gln Gln Arc 50 5	g Ala Ile Ala Pro Phe	e Thr Arg Gly Gly Asp
ATC ATC GCG CAG GCG CAG Ile Ile Ala Gln Ala Gl:		•
ATC GGC CTG CTG CAG CG Ile Gly Leu Leu Gln Arc 85		
CTC GTG CTC TCC CCG AC Leu Val Leu Ser Pro Th 100		
ATC AGC CGC ATC GGC GA Ile Ser Arg Ile Gly Gl 115		
ACC TTT GTG GGT GGC ACC Thr Phe Val Gly Gly Th	r Arg Val Gln Asp Asp	Leu Arg Lys Leu Gln
GCT GGC GTC GTC GCC Ala Gly Val Val Val Al 150		
ATC AAG CGC GGC GCG CT Ile Lys Arg Gly Ala Le 165		
GAC GAG GCT GAT GAG AT Asp Glu Ala Asp Glu Me 180		

			CTG Leu											741
 			GAG Glu 215											789
			CTC Leu										•	837
			ATC Ile											885
			GAG Glu											933
			AAG Lys											981
			AGC Ser 295											1029
	,		AAC Asn											1077
			GTG Val											1125
	Asn		GAC Asp											1173
			GGC Gly											1221
Val			GAC Asp 375						Glu					1269
			GAT Asp					Asp						1317
GAG Glu	. GCG	GGCC	CCT	GCCC	CCCT	TC C	CTGC	cccc	C TC	TCGC	GACG			1366

AGA	GAAC	GCA	CATC	GTAA	CA C	AGCC	ACGC	G AA	CGAT	AGTA	AGG	GCGT	GCG	GCGG	CGTTCC
CCT	CCTC	CTG	CCAG	CGGC	cc c	CCTC	CGCA	G CG	CTTC	TCTT	TTG	AGAG	GGG	GGCA	.GGGGGA
GGC	GCTG	CGC	CTGG	CTGG	AT G	TGTG	CTTG	A GC	TTGC	ATTC	CGT	CAAG	CAA	GTGC	TTTGTT
TTA	ATTA	TGC	GCGC	CGTT	TT G	TTGC	TCGT	c cc	TTTC	GTTG	GTG	TTTT	TTC	GGCC	GAAACG
GCG	TTTA	AAG	CA `				•								
453											,				
(2)	INF		TION		_										٠
		(i) :	SEQU (A)	ENCE ) LE						s					
				) TY:											
	(	ii) I	MOLE	CULE	TYP:	E: p:	rote	in							
	(:	xi) :	SEQUI	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	10:		•		
Met	Ser	Gln	Gln	Asp	Arg	Val	Ala	Pro	Gln	Asp	Gln	Asp	Ser	Phe	Leu
1				5				·	10					15	
Asp	Asp	Gln	Pro 20	Gly	Val	Arg	Pro	Ile 25	Pro	Ser	Phe	Asp	Asp 30	Met	Pro
Leu	His	Gln 35	Asn	Leu	Leu	Arg	Gly 40	Ile	Tyr	Ser	Tyr	Gly 45	Phe	Glu	Lys
Pro	Ser 50	Ser	I,le,	Gln	Gln	Arg 55	Ala	Ile	Ala	Pro	Phe 60		Arg	Gly	Gly
Asp 65	Ile	Ile	Ala	Gln	Ala 70	Gln	Ser	Gly	Thr	Gly 75	Lys	Thr	Gly	Ala	Phe 80
Ser	Ile	Gly	Leu	Leu 85	Gln	Arg	Leu	Asp	Phe 90	Arg	His	Asn	Leu	Ile 95	Gln
Gly	Leu	Val	Leu 100	Ser	Pro	Thr	Arg	Glu 105	Leu	Ala	Leu	Gln	Thr 110	Ala	Glu
Val	Ile	Ser 115	Arg	Ile	Gly	Glu	Phe 120	Leu	Ser	Asn	Ser	Ala 125	Lys	Phe	Cys
Glu	Thr 130	Phe	Val	Gly		Thr 135	Arg	Val	Gln	Asp	Asp 140	Leu	Arg	Lys	Leu
Gln 145	Ala	Gly	Val	Val	Val 150	Ala	Val	Gly	Thr	Pro 155	Gly	Arg	Val	Ser	Asp 160

Val Ile Lys Arg Gly Ala Leu Arg Thr Glu Ser Leu Arg Val Leu Val

170

165

175

Leu Asp Glu Ala Asp Glu Met Leu Ser Gln Gly Phe Ala Asp Gln Ile 180 185 190

Tyr Glu Ile Phe Arg Phe Leu Pro Lys Asp Ile Gln Val Ala Leu Phe 195 200 205

Ser Ala Thr Met Pro Glu Glu Val Leu Glu Leu Thr Lys Lys Phe Met 210 . 215 220

Arg Asp Pro Val Arg Ile Leu Val Lys Arg Glu Ser Leu Thr Leu Glu 225 230 235 240

Gly Ile Lys Gln Phe Phe Ile Ala Val Glu Glu Glu His Lys Leu Asp 245 250 255

Thr Leu Met Asp Leu Tyr Glu Thr Val Ser Ile Ala Gln Ser Val Ile 260 265 270

Phe Ala Asn Thr Arg Arg Lys Val Asp Trp Ile Ala Glu Lys Leu Asn 275 280 285

Gln Ser Asn His Thr Val Ser Ser Met His Ala Glu Met Pro Lys Ser 290 295 300

Asp Arg Glu Arg Val Met Asn Thr Phe Arg Ser Gly Ser Ser Arg Val 305 310 315 320

Leu Val Thr Thr Asp Leu Val Ala Arg Gly Ile Asp Val His His Val 325 330 335

Asn Ile Val Ile Asn Phe Asp Leu Pro Thr Asn Lys Glu Asn Tyr Leu 340 345 350

His Arg Ile Gly Arg Gly Gly Arg Tyr Gly Val Lys Gly Val Ala Ile 355 360 365

Asn Phe Val Thr Glu Lys Asp Val Glu Leu Leu His Glu Ile Glu Gly 370 375 380

His Tyr His Thr Gln Ile Asp Glu Leu Pro Val Asp Phe Ala Ala Tyr 385 390 395 400

Leu Gly Glu

#### (2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 12 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

- (ix) FEATURE:
  - (A) NAME/KEY: Modified-site
  - (B) LOCATION: 6
- (D) OTHER INFORMATION: /note= "Where Xaa is either a Leu or Lys Residue"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

- (2) INFORMATION FOR SEQ ID NO:12:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 26 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ix) FEATURE:
    - (A) NAME/KEY: -
    - (B) LOCATION: 11
    - (D) OTHER INFORMATION: /note= "Where n is inosine"
  - (ix) FEATURE:
    - (A) NAME/KEY: -
    - (B) LOCATION: 17
    - (D) OTHER INFORMATION: /note= "Where n is inosine"
  - (ix) FEATURE:
    - (A) NAME/KEY: -
    - (B) LOCATION: 20
    - (D) OTHER INFORMATION: /note= "Where n is inosine"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

GGAATTCCCC NCAGCTNGTN TTCGAC

- (2) INFORMATION FOR SEQ ID NO:13:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 5 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Lys Val Phe Asp Glu
1 5

- (2) INFORMATION FOR SEQ ID NO:14:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 30 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

GGATCCATGG TCAAGTCCCA CTACATCTGC

30

- (2) INFORMATION FOR SEQ ID NO:15:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 33 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GAATTCAGAC CGGATAGAAA TAAGCCAATG AAA

33

- (2) INFORMATION FOR SEQ ID NO:16:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 701 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
    - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Met Thr Glu Thr Phe Ala Phe Gln Ala Glu Ile Asn Gln Leu Met Ser 1 5 10 15

Leu Ile Ile Asn Thr Phe Tyr Ser Asn Lys Glu Ile Phe Leu Arg Asp

			20				•	25					30		
Val	Ile	Ser 35	Asn	Ala	Ser	Asp	Ala 40	Cys	Asp	Lys	Ile	Arg 45	Tyr	Gln	Ser
Leu	Thr 50	Asp	Pro	Ala	Val	Leu 55	Gly	Asp	Ala	Thr	Arg 60	Leu	Cys	Val	Arg
Val 65	Val	Pro	Asp	Lys	Glu 70	Asn	Lys	Thr	Leu	Thr 75	Val	Glu	Asp	Asn	Gly 80
Ile	Gly	Met	Thr	Lys 85	Ala	Asp	Leu	Val	Asn 90	Asn	Leu	Gly	Thr	Ile 95	Ala
Arg	Ser	Gly	Thr 100	Lys	Ala	Phe	Met	Glu 105	Ala	Leu	Glu	Ala	Gly 110	Ala	.Asp
Met	Ser	Met 115	Ile	Gly	Gln	Phe	Gly 120	Val	Gly	Phe	Tyr	Ser 125	Ala	Tyr	Leu
Val	Ala 130	Asp	Arg	Val	Thr	Val 135	Thr	Ser	Lys	Asn	Asn 140	Ser	Asp	Glu	Val
Tyr 145	Val	Trp	Glu	Ser	Ser 150	Ala	Gly	Gly	Thr	Phe 155	Thr	Ile	Thr	Ser	Ala 160
Pro	Glu	Ser	Asp	Met 165	Lys	Leu	Pro	Ala	Arg 170	Ile	Thr	Leu	His	Leu 175	Lys
Glu	Asp	Gln	Leu 180	Glu	Tyr	Leu	Glu	Ala 185	Arg	Arg	Leu	Lys	Glu 190	Leu	Ile
Lys	Lys	His 195	Ser	Glu	Phe	Ile	Gly 200	Tyr	Asp	Ile	Glu	Leu 205	Met	Val	Glu
Lys	Thr 210	Thr	Glu	Lys	Glu	Val 215	Thr	Asp	Glu	Asp	Glu 220	Glu	Glu	Ala	Lys
Lys 225	Ala	Asp	Glu	Asp	Gly 230	Glu	Glu	Pro	Lys	Val 235	Glu	Glu	Val	Thr	Glu 240
Gly	Glu	Glu	Asp	Lys 245	Lys	Lys	Lys	Thr	Lys 250	Lys	Val	Lys	Glu	Val 255	Thr
Lys	Glu	Tyr	Glu 260	Val	Gln	Asn	Lys	His 265	Lys	Pro	Leu	Trp	Thr 270	Arg	Asp
Pro	Lys	Asp 275	Val	Thr	Lys	Glu	Glu 280	Tyr	Ala	Ala	Phe	Tyr 285	Lys	Ala	Ile
Ser	Asn 290	Asp	Trp	Glu	Asp	Pro 295	Pro	Ala	Thr	Lys	His 300	Phe	Ser	Val	Glu
Gly 305	Gln	Leu	Glu	Phe	Arg 310	Ala	Ile	Met	Phe	Val 315	Pro	Lys	Arg	Ala	Pro 320

Phe	Asp	Met	Leu	Glu 325	Pro	Asn	Lys	Lys	Arg 330	Asn	Asn	Ile	Lys	Leu 335	Tyr
Val	Arg	Arg	Val 340	Phe	Ile	Met	Asp	Asn 345	Cys	Glu	Asp	Leu	Cys 350	Pro	Asp
Trp	Leu	Gly 355	Phe	Val	Lys	Gly	Val 360	Val	Asp	Ser	Glu	Asp 365	Leu	Pro	Leu
Asn	11e 370	Ser	Arg	Glu	Asn	Leu 375	Gln	Gln	Asn	Lys	Ile 380	Leu	Lys	Val	Ile
Arg 385	Lys	Asn	Ile	Val	Lys 390	Lys	Cys	Leu	Glu	Met 395	Phe	Glu	Glu	Val	Ala 400
Glu	Asn	Lys	Glu	Asp 405	Tyr	Lys	Gln	Phe	Tyr 410	Glu	Gln	Phe	Gly	Lys 415	Asn
	-		420					425		Asn			430		
		435					440			Gly	ř	445	•		
	450					455				Glu	460				
465					470		•			Glu 475					480
				485					490	Leu				495	
_			500					505		Asp			510		
Phe	Ala	Cys 515	Leu	Thr	Lys	Glu	Gly 520		His	Phe	Glu	Glu 525	Ser	Glu	Glu
Glu	Lys 530	_	Gln	Arg	Glu	Glu 535		Lys	Ala	Thr	Cys 540	Glu	Lys	Leu	Cys
Lys 545		Met	Lys	Glu	Val 550	Leu	Gly	Asp	Lys	Val 555	Glu	Lys	Val	Thr	Val 560
Ser	: Glu	Arg	Leu	Ser 565		Ser	Pro	Cys	Ile 570	Leu	Val	Thr	Ser	Glu 575	
Gly	Trp	Ser	Ala 580		Met	Glu	Gln	Met 585		Arg	Asn	Gln	Ala 590		Arg
Asp	Ser	Ser 595		Ala	Gln	Tyr	Met 600		. Ser	Lys	Lys	Thr 605		Glu	Leu

Asn Pro Lys His Pro Ile Ile Lys Glu Leu Arg Arg Arg Val Glu Ala 610 615 620

Asp Glu Asn Asp Lys Ala Val Lys Asp Leu Val Phe Leu Leu Phe Asp 625 630 635 640

Thr Ser Leu Leu Thr Ser Gly Phe Gln Leu Glu Asp Pro Thr Tyr Ala 645 650 655

Glu Arg Ile Asn Arg Met Ile Lys Leu Gly Leu Ser Leu Asp Glu Glu 660 665 670

Glu Glu Glu Glu Ala Val Glu Ala Ala Val Ala Glu Thr Ala Pro Ala 675 680 685

Glu Val Thr Ala Gly Thr Ser Ser Met Glu Leu Val Asp 690 695 700

#### (2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 704 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Met Thr Glu Thr Phe Ala Phe Gln Ala Glu Ile Asn Gln Leu Met Ser

1 5 10 15

Leu Ile Ile Asn Thr Phe Tyr Ser Asn Lys Glu Ile Phe Leu Arg Glu 20 25 30

Leu Ile Ser Asn Ala Ser Asp Ala Cys Asp Lys Ile Arg Tyr Gln Ser 35 40 45

Leu Thr Asn Gln Ala Val Leu Gly Asp Glu Ser His Leu Arg Ile Arg 50 55 60

Val Val Pro Asp Lys Ala Asn Lys Thr Leu Thr Val Glu Asp Thr Gly 65 70 75 80

Ile Gly Met Thr Lys Ala Glu Leu Val Asn Asn Leu Gly Thr Ile Ala 85 90 95

Arg Ser Gly Thr Lys Ala Phe Met Glu Ala Leu Glu Ala Gly Gly Asp 100 105 110

Met Ser Met Ile Gly Gln Phe Gly Val Gly Phe Tyr Ser Ala Tyr Leu 115 120 125

Val	Ala 130	Asp	Arg	Val	Thr	Val 135	Val	Ser	Lys	Asn	Asn 140	Asp	Asp	Glu	Ala
Tyr 145	Thr	Trp	Glu	Ser	Ser 150	Ala	Gly	Gly	Thr	Phe 155	Thr	Val	Thr	Pro	Thr 160
Pro	Asp	Cys	Asp	Leu 165	Lys	Arg	Gly	Thr	Arg 170	Ile	Val	Leu	His	Leu 175	Lys
Glu	Asp	Gln	Gln 180	Glu	Tyr	Leu	Glu	Glu 185	Arg	Arg	Leu	Lys	Asp 190	Leu	Ile
Lys	Lys	His 195	Ser	Glu	Phe	Ile	Gly 200	Tyr	Asp	Ile	Glu	Leu 205	Met	Val	Glu
Lys	Ala 210	Thr	Glu	Lys	Glu	Val 215	Thr	Asp	Glu	Asp	Glu 220	Asp	Gļu	Ala	Ala
Ala 225	Thr	Lys	Asn	Glu	Glu 230	Gly	Glu	Glu	Pro	Lys 235	Val	Glu	Glu	Val	Lys 240
Asp	Asp	Ala	Glu	Glu 245	Gly	Glu	Lys	Lys	Lys 250	Lys	Thr	Lys	Lys	Val 255	Lys
Glu	Val	Thr	Gln 260	Glu	Phe	Val	Val	Gln 265	Asn	Lys	His	Lys	Pro 270	Leu	Trp
Thr		Asp . 275	Pro	Lys	Asp	Val	Thr 280	Lys	Glu	Glu	Tyr	Ala 285	Ala	Phe	Tyr
Lys	Ala 290	Ile	Ser	Asn	Asp	Trp 295	Glu	Glu	Pro	Leu	Ser 300	Thr	Lys	His	Phe
Ser 305	Val	Glu	Gly	Gln	Leu 310	Glu	Phe	Arg	Ala	Ile 315	Leu	Phe	Val	Pro	Lys 320
Arg	Ala	Pro	Phe	Asp 325	Met	Phe	Glu	Pro	Ser 330	Lys	Lys	Arg	Asn	Asn 335	Ile
Lys	Leu	Tyr	Val 340	Arg	Arg	Val	Phe	Ile 345	Met	Asp	Asn	Cys	Glu 350	Asp	Leu
Сув	Pro	Glu 355	Trp	Leu	Ala	Phe	Val 360	Arg	Gly	Val	Val	Asp 365	Ser	Glu	Asp
Leu	Pro 370		Asn	Ile	Ser	Arg 375		Asn	Leu	Gln	Gln 380	Asn	Lys	Ile	Leu
Lys 385		Ile	Arg	Lys	Asn 390		Val	Lys	Lys	Ala 395	Leu	Glu	Leu	Phe	Glu 400
Glu	Ile	Ala	Glu	Asn 405	_	Glu	Asp	Tyr	Lys 410		Phe	Туr	Glu	Gln 415	Phe

Gly	Lys	Asn	Val 420	Lys	Leu	Gly	Ile	His 425	Glu	Asp	Ser	Ala	Asn 430	Arg	Lys
Lys	Leu	Met 435	Glu	Leu	Leu	Arg	Phe 440	His	Ser	Ser	Glu	Ser 445	Gly	Glu	Asp
Met	Thr 450	Thr	Leu	Lys	Asp	Tyr 455		Thr	Arg	Met	Lys 460	Glu	Gly	Gln	Lys
Cys 465	Ile	Tyr	Tyr	Val	Thr 470	.Gly	Asp.	Ser_	Lys	Lys 475	Lys	Leu	Glu	Thr	Ser 480
Pro	Phe	Ile	Glu	Gln 485	Ala	Arg	Arg	Arg	Gly 490	Phe	Glu	Val	Leu	Phe 495	Met
Thr	Glu	Pro	Ile 500	Asp	Glu	Tyr	Val	Met 505	Gln	Gln	Val	Lys	Asp 510	Phe	Glu
Asp	Lys ·	Lys 515	Phe	Ala	Cys	Leu	Thr 520	Lys	Glu	Gly	Val	His 525	Phe	Glu	Glu
Thr	Glu 530	Glu	Glu	Lys	Lys	Gln 535	Arg	Glu	Glu	Glu	Lys 540	Thr	Ala	Tyr	Glu
Arg 545	Leu	Cys	Lys	Ala	Met 550	Lys	Asp	Val	Leu	Gly 555	-	Lys	Val	Glu	Lys 560
Val	Val	Val	Ser	Glu 565	Arg	Leu	Ala	Thr	Ser 570	Pro	Cys	Ile	Leu	Val 575	Thr
Ser	Glu	Phé	Gly 580	Trp	Ser	Ala	His	Met 585	Glu	Gln	Ile	Met	Arg 590	Asn	Gln
Ala	Leu	Arg 595	Asp	Ser	Ser	Met	Ser 600	Ala	Tyr	Met	Met	Ser 605	Lys	Lys	Thr
Met	Glu 610	Ile	Asn	Pro	Ala	His 615	Pro	Ile	Val	Lys	Glu 620	Leu	Lys	Arg	Arg
Val 625	Glu	Ala	Asp	Glu	Asn 630	Asp	Lys	Ala	Val	Lys 635	Asp	Leu	Val	Tyr	Leu 640
Leu	Phe	Asp	Thr	Ala 645	Leu	Leu	Thr	Ser	Gly 650	Phe	Thr	Leu	Asp	Asp 655	Pro
Thr	Ser	Tyr	Ala 660	Glu	Arg	Ile	His	Arg 665	Met	Ile	Lys	Leu	Gly 670	Leu	Ser
Leu	Asp	Asp 675	Glu	Asp	Asn	Gly	Asn 680	Glu	Glu	Ala	Glu	Pro 685	Ala	Ala	Ala
Val	Pro	Ala	Glu	Pro	Val	Ala	Gly	Thr	Ser	Ser	Met	Glu	Gln	Val	Asp

#### (2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 732 amino acids
  - (B) TYPE: amino acid ~
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Met Pro Glu Glu Thr Gln Thr Gln Asp Gln Pro Met Glu Glu Glu Glu 1 5 10 15

Val Glu Thr Phe Ala Phe Gln Ala Glu Ile Ala Gln Leu Met Ser Leu 20 25 30

Ile Ile Asn Thr Phe Tyr Ser Asn Lys Glu Ile Phe Leu Arg Glu Leu 35 40 45

Ile Ser Asn Ser Ser Asp Ala Leu Asp Lys Ile Arg Tyr Glu Ser Leu 50 55 60

Thr Asp Pro Ser Lys Leu Asp Ser Gly Lys Glu Leu His Ile Asn Leu 65 70 75 80

Ile Pro Asn Lys Gln Asp Arg Ala Leu Thr Ile Val Asp Thr Gly Ile 85 90 95

Gly Met Thr Lys Ala Asp Leu Ile Asn Asn Leu Gly Thr Ile Ala Lys
100 105 110

Ser Gly Thr Lys Ala Phe Met Glu Ala Leu Gln Ala Gly Ala Asp Ile 115 120 125

Ser Met Ile Gly Gln Phe Gly Val Gly Phe Tyr Ser Ala Tyr Leu Val 130 135 140

Ala Glu Lys Val Thr Val Ile Thr Lys His Asn Asp Asp Glu Gln Tyr 145 150 155 160

Ala Trp Glu Ser Ser Ala Gly Gly Ser Phe Thr Val Arg Thr Asp Thr 165 170 175

Gly Glu Pro Met Gly Arg Gly Thr Lys Val Ile Leu His Leu Lys Glu 180 185 190

Asp Gln Thr Glu Tyr Leu Glu Glu Arg Arg Ile Lys Glu Ile Val Lys
195 200 205

Lys His Ser Gln Phe Ile Gly Tyr Pro Ile Thr Leu Phe Val Glu Lys 210 215 220

Glu 225	Arg	Asp	Lys	Glu	Val 230	Ser	Asp	Asp	Glu	Ala 235	Glu	Glu	Lys	Glu	Asp 240
Lys	Glu	Glu	Glu	Lys 245	Glu	Lys	Glu	Glu	Lys 250	Glu	Ser	Glu	Asp	Lys 255	
Glu	Ile	Glu	Asp 260	Val	Gly	Ser	Asp	Glu 265	Glu	Asp	Glu	Lys	Lys 270	Asp	Gly
Asp	Lys	Lys 275	Lys	Lys	Lys	Lys	Ile 280	Lys	Glu	Lys	Tyr	Ile 285	Asp	Lys	Glu
Glu	Leu 290	Asn	Lys	Thr	Lys	Pro 295	Ile	Trp	Thr	Arg	Asn 300	Pro	Asp	Asp	Ile
Thr 305	Asn	Glu	Glu	Tyr	Gly 310	Glu	Phe	Tyr	Lys	Ser 315	Leu	Thr	Asn	Asp	Trp 320
Glu	Asp	His	Leu	Ala 325	Val	Lys	His	Phe	Ser 330	Val	Glu	Gly	Gln	Leu 335	Glu
Phe	Arg	Ala	Leu 340	Leu	Phe	Val	Pro	Arg 345	Arg	Ala	Pro	Phe	Asp 350	Leu	Phe
Glu	Asn	Arg 355	Lys	Lys	Lys	Asn	Asn 360	Ile	Lys	Leu	Tyr	Val 365	Arg	Arg	Val
Phe	Ile 370	Met	Asp	Asn	Cys	Glu 375	Glu	Leu	Ile	Pro	Glu 380	Tyr	Leu	Asn	Phe
11e 385	Arg	Gly	Val	Val	Asp 390	Ser	Glu	Asp	Leu	Pro 395	Leu	Asn	Ile	Ser	Arg 400
Glu	Met	Leu	Gln	Gln 405	Ser	Lys	Ile	Leu	Lys 410	Val	Ile	Arg	Lys	Asn 415	Leu
Val	Lys	Lys	Cys 420	Leu	Glu	Leu	Phe	Thr 425	Glu	Leu	Ala	Glu	Asp 430	Lys	Glu
Asn	Tyr	Lys 435	Lys	Phe	Tyr	Glu	Gln 440	Phe	Ser	Lys	Asn	Ile 445	Lys	Leu	Gly
Ile	His 450	Glu	Asp	Ser	Gln	Asn 455	Arg.	Lys	Lys	Leu	Ser 460	Glu	Leu	Leu	Arg
Tyr 465	Tyr	Thr	Ser	Ala	Ser 470	Gly	Asp	Glu	Met	Val 475	Ser	Leu	Lys	Asp	Tyr 480
Cys	Thr	Arg	Met	Lys 485	Glu	Asn	Gln	Lys	His 490	Ile	Tyr	Tyr	Ile	Thr 495	Gly
Glu	Thr	Lys	Asp 500	Gln	Val	Ala	Asn	Ser 505	Ala	Phe	Val	Glu	Arg 510	Leu	Arg

Lys	His	Gly 515	Leu	Glu	Val	Ile	Tyr 520	Met	Ile	Glu	Pro	Ile 525	Asp	Glu	Tyr
Cys	Val 530	Gln	Gln	Leu	Lys	Glu 535	Phe	Glu	Gly	Lys	Thr 540	Leu	Val	Ser	Val
Thr 545	Lys	Glu	Gly	Leu	Glu 550	Leu	Pro	Glu	Asp	Glu 555	Glu	Glu	Lys	Lys	Lys 560
Gln	Glu	Glu	Lys	Lys 565	Thr	Lys	Phe	Glu	Asn 570	Leu	Cys	Lys	Ile	Met 575	Lys
Asp	Ile	Leu	Glu 580	Lys	Lys	Val	Glu	Lys 585	Val	Val	Val	Ser	Asn 590	Arg	Leu
Val	Thr	Ser 595	Pro	Cys	Суѕ	Leu	Val 600	Thr	Ser	Thr		Gly 605	Trp	Thr	Ala
Asn	Met 610	Glu	Arg	Ile	Met	Lys 615	Ala	Gln	Ala	Leu	Arg 620	Asp	Asn	Ser	Thr
Met 625	Gly	Tyr	Met	Ala	Ala 630	Lys	Lys	His	Leu	Glu 635	Ile	Asn	Pro	Asp	His 640
Ser	Ile	Ile	Glu	Thr 645	Leu	Arg	Gln	Lys	Ala 650	Glu	Ala	Asp	Lys	Asn 655	Asp
Lys	Ser	Val	Lys 660	Asp	Leu	Val	Ile	Leu 665	Leu	Tyr	Glu	Thr	Ala 670	Leu	Leu
Ser	Ser	Gly 675	Phe	Ser	Leu	Glu	Asp 680	Pro	Gln	Thr	His	Ala 685	Asn	Arg	Ile
Tyr	Arg 690	Met	Ile	Lys	Leu	Gly 695	Leu	Gly	Ile	Asp	Glu 700		Asp	Pro	Thr
Ala 705	Asp	Asp	Thr	Ser	Ala 710	Ala	Val	Thr	Glu	Glu 715	Met	Pro	Pro	Leu	Glu 720

#### (2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1019 base pairs

725

Gly Asp Asp Asp Thr Ser Arg Met Glu Glu Val Asp

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

### (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 71..523

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

(0.2, 2-202 22 4-2 22 22 10 10 12 1													
GAATTCGGCA CGAGGTTTCT GTACTTTATT GCTTCCAGCC TTTATTCACT CTTCGATTTC 60													
CTCTAACACC ATG TCC TCC GAG CGC ACC TTT ATT GCC GTC AAG CCG GAC  Met Ser Ser Glu Arg Thr Phe Ile Ala Val Lys Pro Asp  1 5 10	109												
GGC GTG CAG CGC GGC CTC GTT GGC GAG ATC ATC GCC CGC TTC GAG CGC Gly Val Gln Arg Gly Leu Val Gly Glu Ile Ile Ala Arg Phe Glu Arg 15 20 25	157												
Lys Gly Tyr Lys Leu Val Ala Leu Lys Ile Leu Gln Pro Thr Thr Glu 30 35 40 45	205												
CAG GCC CAG GGT CAC TAT AAG GAC CTT TGC TCC AAG CCG TTT TTC CCG Gln Ala Gln Gly His Tyr Lys Asp Leu Cys Ser Lys Pro Phe Phe Pro 50 55 60	253												
GCC CTT GTG AAG TAC TTC TCC TCT GGC CCG ATC GTG TGT ATG GTG TGG Ala Leu Val Lys Tyr Phe Ser Ser Gly Pro Ile Val Cys Met Val Trp 65 70 75	301												
GAG GGT AAG AAC GTG GTG AAG AGC GGC CGC GTG CTC CGC GCG ACG Glu Gly Lys Asn Val Val Lys Ser Gly Arg Val Leu Leu Gly Ala Thr 80 85 90	349												
AAC CCG GCC GAC TCA CAG CCC GGC ACG ATC CGT GGC GAC TTT GCC GTG Asn Pro Ala Asp Ser Gln Pro Gly Thr Ile Arg Gly Asp Phe Ala Val 95 100 105	397												
GAT GTG GGC CGC AAC GTG TGC CAC GGG TCC GAC TCT GTG GAG AGC GCG Asp Val Gly Arg Asn Val Cys His Gly Ser Asp Ser Val Glu Ser Ala 110 125	445												
GAG CGC GAG ATC GCC TTT TGG TTC AAG GCG GAT GAG ATC GCG AGC TGG Glu Arg Glu Ile Ala Phe Trp Phe Lys Ala Asp Glu Ile Ala Ser Trp 130 135 140	493												
ACG TCG CAC TCC GTG TCC CAG ATC TAT GAG TAACGGTGAT TGCGGACACG Thr Ser His Ser Val Ser Gln Ile Tyr Glu 145 150	543												
CTTTGAGGAC GTAGCTGTAC CCCCAATGAA TTCTTCTCTG AAAACCACAT CATAAGCCTC	603												
TTAAGAGGTT ATTTTCTTG ATCGATGCCC GGTGGTGACC AGCACCATTC CTTTATCGGA	663												
TTCACTCACA CTCCTAGCGA ATCATGTAGT GCGGTGAGAG TGGGCTCTGG AGGAGACTGT	723												

TGTG	TAGO	CA I	GGC1	TCAG	G AG	AGA#	AAACA	AAA A	TAC	AAGG	AAAC	GCA	ATA :	rgtaz	CTATO	3
GGGT	TCCC	TT T	TTTA	CTAT	G CA	AAGI	TTTT	TATA	ACTO	CCTG	ATC	GCA	AAA A	ACAAC	CAACA	7
CCGC	CATA	.CA C	CAAG	AGCA	LA AJ	GCTI	TCT	CTO	GGG <i>I</i>	ACTG	TGCI	TCTC	TT T	CTTTT	TATT	3
AAGG	AGTG	AC I	CGCG	CGAI	G AA	AAGI	rgtgi	GCG	TGGC	BAGA	TGTA	TTTC	CT 1	TTTT	TGTT	?
ATAG	TGGC	GA C	CAGCI	CACI	G TI	GAC	SATGA	CAA	AAA	AAA	AAA	AAA	AAA (	CTCGF	\G	
(2)	INFC	RMAT	CION	FOR	SEQ	ID N	10:20	): .								
	;	i) S	(B)	TYP	IGTH: PE: a	ACTE 151 minc SY: 1	l ami	ino a id		3						
	(i	.i) M	OLEC	ULE	TYPE	E: pr	otei	in								
	(х	:i) S	EQUE	NCE	DESC	RIPI	NOI?	SEC	) ID	NO:2	:0:					
Met 1	Ser	Ser	Glu	Arg 5	Thr	Phe	Ile	Ala	Val	Lys	Pro	Asp	Gly	Val 15	Gln	
Arg	Gly	Leu	Val 20	Gly	Glu	Ile	Ile	Ala 25	Arg	Phe	Glu	Arg	Lys 30	Gly	Tyr	
Lys	Leu	Val 35	Ala,	Leu	Lys	Ile	Leu 40	Gln	Pro	Thr	Thr	Glu 45	Gln	Ala	Gln	
Gly	His 50	_	Lys	Asp	Leu	Cys 55	Ser	Lys	Pro	Phe	Phe 60	Pro	Ala	Leu	Val	
Lys 65	Tyr	Phe	Ser	Ser	Gly 70	Pro	Ile	Val	Cys	Met 75	Val	Trp	Glu	Gly	Lys 80	
Asn	Val	Val	Lys	Ser 85	Gly	Arg	Val	Leu	Leu 90	Gly	Ala	Thr	Asn	Pro 95	Ala	
Asp	Ser	Gln	Pro 100	Gly	Thr	Ile	Arg	Gly 105	Asp	Phe	Ala	Val	Asp 110	Val	Gly	
Arg	Asn	Val 115	Cys	His	Gly	Ser	Asp 120	Ser	Val	Glu	Ser	Ala 125	Glu	Arg	Glu	
Ile	Ala 130	Phe	Trp	Phe	Lys	Ala 135	Asp	Glu	Ile	Ala	Ser 140	Trp	Thr	Ser	His	
Ser	Val	Ser	Gln	Ile	Tyr	Glu			•				•			

(2) INFORMATION FOR SEQ ID NO:21:

NO 98/35045 PCT/US98/03002

	(i)	· ()	A) L1 B) T' C) S'	ENGT: YPE : TRAN	H: 1 nuc DEDN	523 : leic	ISTI base aci sing ear	pai: d	rs								
	(ii)	MO	LECU	LE T	YPE:	cDN	A							-			
	(vi)	OR:	•				ishma	ania	maj	or							٠
	(ix)		A) N	AME/	KEY:	CDS	. 973										
	(xi)	SE	QUEN	CE DI	ESCR:	IPTI	ON: 8	SEQ :	ID NO	0:21	<b>:</b> .						
GAA:	rtcgo	GCA (					GAC A										49
							AAG Lys 20									- -	97
		Cys					GGT Gly										145
							AAG Lys									* a	193
							AGC Ser									·	241
							AGC Ser										289
			-				GGT Gly 100										337
							AGC Ser										385
							TAC Tyr										433
CCG	CTC	GCC	TCC	<b>አ</b> አር	እጥሮ	GAG	CNC	ጥርር	አጥር	CAC	TOO	CAC	CCA	CAC	N C C		401

Pro	Val	Ala	Cys	Asn 145	Ile	Glu	His	Cys	Met 150	Gln	Cys	Asp	Pro	Gln 155	Thr	
CCG Pro																529
GAC Asp																577
AAG Lys														ACC Thr		625
														ACG Thr		673
														AGC Ser 235		721
														AAG Lys		769
														ATG Met		817
														CTC Leu		865
														GCC Ala		913
										Leu					GCC Ala	961
			GCA Ala 320		TGCG	CAG	CGGC	ATGC	ga a	CAAC	CCCA	с тс	TCAT	TCTC		1013
CAA	CATG	TGC	ATAC	ACAC	AC A	CACA	GACA	G CG	GGGC	AGCA	CCC	CCTC	ccc	ACAC	ACACAC	1073
ACG	CACT	TCC	CCCT	TGTC	TT G	TTCT	TCTT	T CC	TCGT	TCGC	ATT	TCTT	TCT	CTCG	TGCGCT	1133
GGC	GCCG	GCC	TCCT	GCAC	GT C	GCTC	CCCT	'C C	CCTA	ACCT	CTA	TTCT	CTC	TCTC	TCTCTC	1193
TCT	CGCC	:GGC	ATCA	TTGC	TT C	TTAC	CCTI	T TC	TGAT	CCTT	GCI	'CGCG	TGG	GCGG	ACACTG	1253

									10.	,					
CCA	.CAGI	CCC	ACAG	CGCA	GA C	ACAC	GTGI	T TA	AACC	GCGC	AGO	CATO	CCT	CCCI	TATCACT
TCA	TTTC	TCC	TAAA	.GCCA	CTC	ACCA	AGTO	CG CA	CACC	GCCC	TCC	CCCA	ATCG	GCC	SCCCTTC
CGG	GCGC	AGC	TGTG	CGGA	AT C	GGTG	TGTG	C TO	GACC	TCGT	TCC	TGGC	AGC	TCAC	TCGCAT
GTG	TACA	.GCC	ACTC	CAAC	CA C	GAAA	.GCTC	T CI	TCTG	CGCA	CAT	'AAAA	AAA	AAAA	AAAAA
AAA	AACT	CGA	GGGG	GGGC	CC G	GTAC	CCAA	A							
(2)	INF	ORMA	TION	FOR	SEC	ID	NO : 2	2:							
						RACT			:						
	1		(A	) LE	NGTH	: 32 amin	0 am	ino		s					
						GY:									
	(	ii)	MOLE	CULE	TYP	E: p	rote	in							
	(:	xi)	SEQU	ENCE	DES	CRIP	TION	: SE	Q ID	NO:	22:				
Val 1	Leu	Pro	Asp	Met 5	Thr	Cys	Ser	Leu	Thr 10	Gly	Leu	Gln	Cys	Thr 15	Asp
Pro	Asn	Cys	Lys 20	Thr	Cys	Thr	Thr	Tyr 25	Gly	Gln	Cys	Thr	Asp 30	Cys	Asn
Asp	Gly	Tyr 35	Gly	Leu	Thr	Ser	Ser 40	Ser	Val	Cys	Val	Arg 45	Cys	Ser	Val
Ala	Gly 50	Cys	Lys	Ser	Cys	Pro 55	Val	Asp	Ala	Asn	Val 60	Cys	Lys	Val	Cys
Leu 65	Gly	Gly	Ser	Glu	Pro 70	Ile	Asn	Asn	Met	Cys 75	Pro	Cys	Thr	Asp	Pro 80
Asn	Cys	Ala	Ser	Cys 85	Pro	Ser	Asp	Ala	Gly 90	Thr	Cys	Thr	Gln	Суs 95	Ala
Asn	Gly	Tyr	Gly 100	Leu	Val	Asp	Gly	Ala 105	Cys	Val	Arg	Cys	Gln 110	Glu	Pro
Asn	Cys	Phe 115	Ser	Cys	Asp	Ser	Asp 120	Ala	Asn <sup>.</sup>	Lys	Cys	Thr 125	Gln	Cys	Ala
Pro	Asn 130	Tyr	Tyr	Leu	Thr	Pro 135	Leu	Leu	Thr	Cys	Ser 140	Pro	Val	Ala	Cys
Asn 145	Ile	Glu	His	Cys	Met 150	Gln	Cys	Asp	Pro	Gln 155	Thr	Pro	Ser	Arg	Cys 160
Gln	Glu	Cys	Val	Ser	Pro	Tyr	Val	Val	Asp	Ser	Tyr	Asp	Gly	Leu	Cys

Arg Leu Ser Asp Ala Cys Ser Val Pro Asn Cys Lys Lys Cys Glu Thr 185

Gly Thr Ser Arg Leu Cys Ala Glu Cys Asp Thr Gly Tyr Ser Leu Ser 200 195

Ala Asp Ala Thr Ser Cys Ser Ser Pro Thr Thr Gln Pro Cys Glu Val 215

Glu His Cys Asn Thr Cys Val Asn Gly Asp Ser Thr Arg Cys Ala Tyr 235

Cys Asn Thr Gly Tyr Tyr Val Ser Asp Gly Lys Cys Lys Ala Met Gln 250 245

Gly Cys Tyr Val Ser Asn Cys Ala Gln Cys Met Leu Leu Asp Ser Thr 265

Lys Cys Ser Thr Cys Val Lys Gly Tyr Leu Leu Thr Ser Ser Tyr Ser 280

Cys Val Ser Gln Lys Val Ile Asn Ser Ala Ala Ala Pro Tyr Ser Leu 295

Trp Val Ala Ala Ala Val Leu Leu Thr Ser Phe Ala Met His Leu Ala 315 305 310

#### (2) INFORMATION FOR SEQ ID NO:23:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 797 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major
- (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 27..623
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

CTGTACTTTA TTGCCACCAG CCAGCC ATG TCC TGC GGT AAC GCC AAG ATC AAC 53 Met Ser Cys Gly Asn Ala Lys Ile Asn

TCT CCC GCG CCG TCC TTC GAG GAG GTG GCG CTC ATG CCC AAC GGC AGC Ser Pro Ala Pro Ser Phe Glu Glu Val Ala Leu Met Pro Asn Gly Ser 20 10 15

TTC	AAG	AAG	ATC	AGC	CTC	TCC	TCC	TAC	AAG	GGC	AAG	TGG	GTC	GTG	CTC	149
Phe	Lys	Lys	Ile	Ser 30	Leu	Ser	Ser	Tyr	Lys 35	Gly	Lys	Trp	Val	Val 40	Leu	
			CCG Pro 45													197
GCG Ala	TTC Phe	TCC Ser 60	GAC Asp	AGC Ser	GTG Val	AGT Ser	CGC Arg 65	TTC Phe	AAC Asn	GAG Glu	CTC Leu	AAC Asn 70	TGC Cys	GAG Glu	GTC Val	245
CTC Leu	GCG Ala 75	TGC Cys	TCG Ser	ATA Ile	GAC Asp	AGC Ser 80	GAG Glu	TAC Tyr	GCG Ala	CAC His	CTG Leu 85	CAG Gln	TGG Trp	ACG Thr	CTG Leu	293
CAG Gln 90	GAC Asp	CGC Arg	AAG Lys	AAG Lys	GGC Gly 95	GGC Gly	CTC Leu	GGG Gly	ACC Thr	ATG Met 100	GCG Ala	ATC Ile	CCA Pro	ATG Met	CTA Leu 105	341
GCC Ala	GAC Asp	AAG Lys	ACC Thr	AAG Lys 110	AGC Ser	ATC Ile	GCT Ala	CGT Arg	TCC Ser 115	TAC Tyr	GGC Gly	GTG Val	CTG Leu	GAG Glu 120	GAG Glu	389
			GTG Val 125													437
			CAG Gln													485
GAG Glu	GAG Glu 155	GTT Val	CTA Leu	CGC Arg	CTG Leu	CTG Leu 160	GAG Glu	GCT Ala	TTT Phe	CAG Gln	TTC Phe 165	GTG Val	GAG Glu	AAG Lys	CAC His	533
GGC Gly 170	GAĠ Glu	GTG Val	TGC Cys	CCC Pro	GCG Ala 175	AAC Asn	TGG Trp	AAG Lys	AAG Lys	GGC Gly 180	GCC Ala	CCC Pro	ACG Thr	ATG Met	AAG Lys 185	581
CCG Pro	GAA Glu	CCG Pro	AAT Asn	GCG Ala 190	TCT Ser	GTC Val	GAG Glu	GGA Gly	TAC Tyr 195	TTC Phe	AGC Ser	AAG Lys	CAG Gln			623
AAAT	.CCTG	TG A	AGCGI	CGCA	G GA	GTCA	GTGT	GAC	CTCA	.ccc	GCCI	CTGC	CA G	TGGG	TGCGA	683
GAGG	GCGI	GA C	GGAT	TGTG	G GA	AGGC	TGTT	GGA	TATG	ATG	CAGA	CAGC	GA T	GAAT	GCAAC	743
TCCC	ACAC	AC I	rggcc	CTCC	T CA	GCCC	TCTC	CAC	ACAG	ACA	CACG	CACG	CA T	GTG	•	797

## (2) INFORMATION FOR SEQ ID NO:24:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 199 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Met Ser Cys Gly Asn Ala Lys Ile Asn Ser Pro Ala Pro Ser Phe Glu
1 5 10 15

Glu Val Ala Leu Met Pro Asn Gly Ser Phe Lys Lys Ile Ser Leu Ser 20 25 30

Ser Tyr Lys Gly Lys Trp Val Val Leu Phe Phe Tyr Pro Leu Asp Phe 35 40 45

Ser Phe Val Cys Pro Thr Glu Val Ile Ala Phe Ser Asp Ser Val Ser 50 55 60

Arg Phe Asn Glu Leu Asn Cys Glu Val Leu Ala Cys Ser Ile Asp Ser 65 70 75 80

Glu Tyr Ala His Leu Gln Trp Thr Leu Gln Asp Arg Lys Lys Gly Gly
85 90 95

Leu Gly Thr Met Ala Ile Pro Met Leu Ala Asp Lys Thr Lys Ser Ile 100 105 110

Ala Arg Ser Tyr Gly Val Leu Glu Glu Ser Gln Gly Val Ala Tyr Arg 115 120 125

Gly Leu Phe Ile Ile Asp Pro His Gly Met Leu Arg Gln Ile Thr Val

Asn Asp Met Pro Val Gly Arg Ser Val Glu Glu Val Leu Arg Leu Leu 145 150 155 160

Glu Ala Phe Gln Phe Val Glu Lys His Gly Glu Val Cys Pro Ala Asn 165 170 175

Trp Lys Lys Gly Ala Pro Thr Met Lys Pro Glu Pro Asn Ala Ser Val 180 185 190

Glu Gly Tyr Phe Ser Lys Gln 195

- (2) INFORMATION FOR SEQ ID NO:25:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 637 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear

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(ii)	MOLECULE	TYPE:	CDNA
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#### (vi) ORIGINAL SOURCE:

(A) ORGANISM: Leishmania tropica

#### (ix) FEATURE:

(A) NAME/KEY: CDS(B) LOCATION: 7..624

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

	127	., 50	QODI.	CB D	LISCR	1111	ON:	SEQ	ID N	0:25	) :						
TTA		ATG Met 1															48
											Ala				AAC Asn 30	a •	96
		TTC Phe			Ile					Tyr							144
		TTC Phe		Tyr											GAG Glu	. *	192
		GCG Ala 65						Ser					Leu				240
												His			TGG		288
		CAG Gln									Ala				CCA Pro 110	-	336
		GCC Ala								Arg					Leu		384
		AGC Ser															432
CGT Arg	GGC Gly	ATG Met 145	GTG Val	CGT Arg	CAG Gln	ATC Ile	ACC Thr 150	Val	AAC Asn	GAC Asp	ATG Met	CCG Pro 155	GTG Val	GGC Gly	CGC Arg		480
		GAG Glu															528

AAG	CAC	GGC (	GAG	GTG	TGC	CCC	GCG	AAC	TGG	AAG	AAG	GGC	GCC	CCC	ACG		576
					Cys 180												
			Glu		AAG Lys												624
TAAG	AATT	CC A	TG ·														637
(2)	INFO	RMAT	ION	FOR	SEQ	ID N	0:26	5:									
•	(	i) S	(A) (B)	TY!	CHAR IGTH: PE: a	206 minc	ami aci	no a		5	÷						
	(i	.i) M	OLEC	ULE	TYPE	E: pr	otei	in									
	(х	i) S	EQUE	NCE	DESC	CRIPT	ION:	: SEC	QID	NO:2	26 :						
Met 1	His	His	His	His 5	His	His	Met	Ser	Cys 10	Gly	Asn	Ala	Lys	Ile 15	Asn		
Ser	Pro	Ala	Pro 20	Pro	Phe	Glu	Glu	Met 25	Ala	Leu	Met	Pro	Asn 30	Gly	Ser		
Phe	Lys	Lys 35	;Ile,	ser	Leu	Ser	Ala 40	Tyr	Lys	Gly	Lys	Trp 45	Val	Val	Leu	•	
Phe	Phe 50	Tyr	Pro	Leu	Asp	Phe 55	Thr	Phe	Val	Cys	Pro 60	Thr	Glu	Ile	Ile		
Ala 65	Phe	Ser	Asp	Asn	Val 70	Ser	Arg	Phe	Asn	Glu 75	Leu	Asn	Cys	Glu	Val 80		•
Leu	Ala	Сув	Ser	Met 85	Asp	Ser	Glu	Tyr	Ala 90	His	Leu	Gln	Trp	Thr 95	Leu		
Gln	Asp	Arg	Lys 100	Lys	Gly	Gly	Leu	Gly 105	Ala	Met	Ala	Ile	Pro 110	Met	Leu		
Ala	Asp	Lys 115	Thr	Lys	Ser	Ile	Ala 120		Ser	Tyr	Gly	Val 125		Glu	Glu		
Ser	Gln 130	Gly	Val	Ala	Tyr	Arg 135	Gly	Leu	Phe	Ile	Ile 140		Pro	Arg	Gly		
Met 145		Arg	Gln	Ile	Thr 150		Asn	Asp	Met	Pro 155		. Gly	Arg	Asn	160		
Glu	Glu	Ala	Leu	Arg	Leu	Leu	Glu	Ala	Lev	Gln	Phe	val	. Glu	Lys	His		

Gly Glu Val Cys Pro Ala Asn Trp Lys Lys Gly Ala Pro Thr Met Lys 180 185 190

Pro Glu Pro Lys Ala Ser Val Glu Gly Tyr Phe Ser Lys Gln
195 200 205

- (2) INFORMATION FOR SEQ ID NO:27:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 51 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "50 PCR primer (MAPS-1-50His) to simultaneously amplify MAPS-1 cDNA for both L. major and L. tropica while adding 6 His residues to amino terminal end of encoded protein."

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

CAATTACATA TGCATCACCA TCACCATCAC ATGTCCTGCG GTAACGCCAA G

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- (2) INFORMATION FOR SEQ ID NO:28:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 31 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: other nucleic acid
  - (A) DESCRIPTION: /desc = "3' PCR primer (MAPS-1-3ÕR1) to simultaneously amplify MAPS-1 cDNA for both L. major and L. tropica.ó
    - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

CATGGAATTC TTACTGCTTG CTGAAGTATC C

- (2) INFORMATION FOR SEQ ID NO:29:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 520 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
    - (ii) MOLECULE TYPE: DNA (genomic)
    - (vi) ORIGINAL SOURCE:
      - (A) ORGANISM: Leishmania major

(xi) SEQUENCE D	DESCRIPTION:	SEO	ID	NO:29:
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60	CAATTTGCGT	GGGAGTTCTT	GATATTCGCG	TTTGCTCGCC	CTTGCCTACA	GGCACGAGCC
120	CGAAGGTGAT	TCGTGGCGCA	GCGCAGCTCG	CGCGCAACAA	TGCTCAATGT	CGCGTAGAAC
180	ACTGCTCACG	CGGTGAATGT	GCGGCCGATG	TGCGGCCAGT	GTGCGGCCCA	GGCCAGTCCA
240	TGCCACGCGC	TCCACATCAA	CGAGACGCGC	TGAGGACGAG	AAATCGTAGC	CGCGAGCGGC
300	CATAATCTCG	CGGAATGCCG	GTGTCCATTA	NCACGCGGCT	GTTGTAATGA	CCACAGGTCC
360	CTCGCGCGTC	CAGCGAAATT	ATGGCGGACA	GTCGCCGTGC	CCTGGGGCAT	TCGCGCTCCG
420	NGCGTGGGCG	ANAAAACCAC	CGGGTGTTGC	CACCTTTTTG	CAAGCTGCTC	ATCTTCTTGG
480	CTTCGTCNAC	TTGTTTTTCT	GAAATCGAAT	ACTCCATCAA	TGTCGTACAA	ATCGTTAAGC
520			GGTGATCTCA	CGCTNTCCAC	TACTGTTTAA	NGANACAAAN

#### (2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 600 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

GGCACAAGGT	TTTCGGGTTA	TCTTCACGCA	TGGTGGAGCG	CAGATGGGTG	AAGTAAATAC	. 60
GCGGACCGAA	CTGCTTGATC	ATATCAACCA	GATCGTTGTC	AGCACGCACG	CCGTANGAAC	120
CGGTGCACAT	GGTAAAACCG	TNTGCCATGC	TGTTTACGGT	ATCAACCATC	CACTGCATAT	180
CTTCAATGGT	GGAAACAATG	CGCGGCAGGC	CGAGGATCCG	GCGCGGCTCA	TCATNNAGNT	240
NATNAACCAN	TCGCACGTCT	ANTTCTGCAC	TAAACTACAA	NTATCGGTNA	CATATNATAA	300
GGCCNATTTT	CGGTCCAGGA	NTATGTNCTN	TCAAAATGCC	NCGTTANNCA	CTCTTAAATG	360
TCTCANGNGN	AAANTNGTTC	TAAAGGGTGT	CCAAAANNTN	NTTACCNTTC	CCCNCTTACT	420
TCAANANCTC	CTCNAATTCC	CNGGCCCTTN	GACNANNATT	TNCTATTAAA	ANATANAANN	480

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TTCAAATTNA	TTCCCNACCT	NCCNTNNCCA	AANNTANCNA	ATAATCANNC	CCCTNTCANN	540
אאיזירירמאיר	TTACCCTCCN	NTNCNNCCCN	NININICONI A TONI	CCCCAANCCC	NCNCTAAATA	<b>COO</b>
MINICCCMIC	TIACCCICCN	MINGMINGGIN	MMMCCMATIM	CCCCAMINCCC	NCNCIAAAIA	600

#### (2) INFORMATION FOR SEQ ID NO:31:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 600 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE: .
  - (A) ORGANISM: Leishmania major

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

GGCACGAGCC	TCAGTGGAGC	TCAATGAAGA	TATTGCAGTA	TCTTACTCTG	GATGGCACTC	60
AGGTCTCCGG	CACGCTGCCG	CCCCAGTGGA	GCGCGATGGC	ATCGGTGCGA	ATTCTTAACC	120
TGNAGGGTAC	TGAGGTCTCT	GGTACGCTGC	CGCCTGAGTG	GATATCNATG	ANCAGGCTGC	180
AAACTCTGAA	TCTGCGGCGC	ACGAAANTAT	CCGGCACTCT	GCCGCCCGAA	TGGANTTCTA	240
TGAACAGCCT	GGAGTACTTT	CACCTTTATC	TTACTCAGGT	CTCCGGCACG	CTGCCGCCCG	300
AGTGGAGTGG	GATGTCNAAG	GCCGCATACT	TCTGGCTGGA	ATACTGCGAC	CTGTCCGGCA	360
NTCTGCCGCC	CNAGTGGTCG	TCNATGCCAA	AGCTGCGCGG	TATCTCACTG	ANCGGCAACA	420
AATTCTTGCG	NGTGTNTNCC	NGACTCNTGG	GATTCAGAAA	GGTGGTCCTT	GTTGTTGGGC	480
ATCNAAGGAN	CAAACCCCAA	NGGGCCCNCN	AATTGCTTGG	GCNTGCTTAA	GGANTTGCAC	540
NAACCAACNC	CNCCAAAAAC	CCCCCCACC	NCNAAANNAC	NANCECCCAC	TTAANNCCCN	600

#### (2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 600 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

NGCACGAGAA	GCGCAACTGG	CGCATCGCAT	CTGTGACTAT	CTGCCTGAAC	AGGGGCAATN	60
GTTTGTTGGT	AACAGCCTGG	TGGTACGTCT	GATTGATNCG	CTTNCGCAAN	TTCCGGCAGG	120
TTACCCGGTG	TACANCAACC	GTGGGGCCAN	CGGTATCNAC	NGGCTGCTTT	CGACCGCCGC	180
CGGNGTTCAN	CGGGCAANCG	GCAAACCGAC	GCTGGCGATT	GTGGGCGATC	TCTCCGCACT	240
TTACGATCTC	AACGCNCTGG	CGTTATTGCG	TCAGGTTTCT	GCGCCGCTGG	TATTAATTGT	300
GGTGAACAAC	AACGGCNGGG	CAAAATTTTC	TCGCTGTTGC	CAACGCCCCC	AAAGCNAGCG	360
TGAAGCGTTT	CTATCTGATG	CCGCAAAACG	TCCATTTTGA	AACACGCCGC	CNCCCATGTT	420
TCGANCTGAA	AATATCATCG	TCCGCAAAAC	TGGCANGAAA	CTTNGAAAAC	CGCATTTTGC	480
CGACNCCCTG	GCNCACGCCC	AACCCACCCA	CCGGTTGATT	GAAAATGGTG	GGTTAACGAA	540
NCCNNATGGG	TGCCCCAAAN	CNCNNCCANC	CAAATTTCTG	GGCCCAGGTT	AAANCCCTTT	600

#### (2) INFORMATION FOR SEQ ID NO:33:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 600 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

ACGATGACCA	TGCCCCGAAG	GAGGATGGCC	ATGCGCCGAA	GAACGATGAC	CATGCCCCGA	60
AGGAGGATGG	CCATGCGCCG	AAGAACGATG	ACCATGCCCC	GAAGGAGGAT	GGCCATGCGC	120
CGAAGAACGA	CGGGGATGTG	CAGAANAAGA	GCGAAGATGG	AGACAACGTG	GGAGAGGGAG	180
GCAAGGGCAA	TGAGGATGGT	AACGATGATC	AGCCGAAGGA	GCACGCTGCC	GGCAACTAGT	240
GGGCTGCGTC	CGGGCTTGTG	TGCGANCCGT	GCTCTGCACC	CCGCCGCTCG	TGCATCCTCG	300
CATGTGGACT	GCGTGTGTCT	CTCCCGCTTT	GTCTCTCTCC	CCCACACAGT	GGCTGATGCC	360

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TGCACGGGGT	TGCTGTGGCT	GCACCTCCTG	ACCACTGCCA	GCTTTCTTGG	CTTGCCTCCC	420
CTCTGCGCCT	CCGCTCGTGC	CGCTCGTGCC	GAATTCGATA	TCAAGCTTAT	CGATACCGTC	480
NACCTCGAAG	GGGGGCCCGG	TTACCCATTC	GCCCTATANT	GAGTCNTATT	ACAATTCCTG	540
GCGTCGTTTT	ACACGTCGTG	ACTGGGAAAA	ACCCTGGCGT	TCCCCACTTA	TCGCCTTGCA	600

#### (2) INFORMATION FOR SEQ ID NO:34:

WO 98/35045

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 516 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

		•				
AGCTGCAGCA	GCGCCTAGAC	ACCGCCACGC	AGCAGCGCGC	CGAGCTGGAG	GCACGGGTGG	.60
CACGGCTGGC	CGCGGACCGC	GACGAGGCGC	GCCAGCAGCT	GGCCGCGAAC	GCCGAGGAGC	120
TGCAGCAGCG	CCTAGACACC	GCCACGCAGC	AGCGCGCCGA	GCTGGAGGCA	CGGGTGGCAC	180
GGCTGGCCGC	GGACGGCGAC	GAGGCCCGCC	AGCAGCTGGC	CGCGAACGCC	GAGGAGCTGC	240
AGCAGCGCCT	AGACACCGCC	ACGCAGCAGC	GCGCCGAGCT	GGAGGCACAG	GTGGCACGGC	300
TGGCCGCGAA	CGCCGAGGAG	CTGCAGCAGC	GCCTAGACAC	CGCCACGCAG	CAGCGCGCCG	360
AGCTGGAGGC	ACGGGTGGCA	CGGCTGGCCG	CGGACCGCGA	CGAGGCGCGC	CAGCAGCTGG	420
CCGCGAACGC	CGAGGAGCTG	CAGCAGCGCC	TAGACACCGC	CACGCAGCAG	CGCGCCGAGC	480
TGGARGCACA	GGTGGCACGG	CTGGCCGCGA	AMGCCG			516

#### (2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 822 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:

#### (A) ORGANISM: Leishmania major

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

GGCACGANAG	ATCTTCGTGA	AGACGCTGAC	CGGCAANACG	ATCGCGCTGG	AGGTGGAGCC	60
<b>GAGC</b> GACACG	ATCGAGAACG	TGAAGGCCAA	GATCCAGGAC	AAGGAGGGCA	TCCCGCCGGA	120
CCAGCAGCGC	CTGATCTTCG	CCGGCAAGCA	GCTGGAGGAN	GGCCGCACGC	TCTCGGACTA	180
CAACATCCAG	AAGGAGTCCA	CGCTGCACCT	GGTGCTGCGC	CTGCGCGGCG	GCATGCANAT	240
CTTCGTGAAA	ACGCTNACCG	GCAANACAAT	CGCGCTGGAA	GTGGAGCCGA	ACGACCNATC	300
GAAAACGTGA	AGGCCNANAT	CCANGACAAG	GAAGGCNTCC	CGCCGGANCA	GCACGCCTGA	360
TCTTCCNCCG	GCAACCACTT	GANGAAGGGC	NCACGCTCTC	NGACTACNAC	ATCCANAAAG	420
GATTCCNCCC	TGCACCTTGT	TGCTTGCNCC	TTGCTCGGGG	GGCATGCCNA	ATCTTCCTTN	480
AAAACCTCAA	CCGGCAANAA	CAATCCCCCN	CNGAAGTTGG	AACCCAACCA	NCCCATTCNA	540
AAACTTTAAA	GGCCNNNATT	CCNGAACAAN	GAAGGGCTTC	CCCCCGGAC	CNNCAANCNC	600
CCTGATTNTT	CCCCCGGNNN	NCANTTTGGA	ANGAAGGCC	CCNCCCTCCN	CCGAATTNCN	660
ACNTCCCNAA	ANGGATTCCC	CCCCTNCCCT	TGNTTTTTGC	GCCNNNNNC	GGCNNCNTNC	720
CNAAATTCCG	NCCNAAGGNC	CCCANTANAN	CNACTTTCCC	NTTCCCCCCC	NNNTTTTGC	780
TTTTNAAATN	TNCCCCCNNA	AANNTCCCNT	TTNCNANTTN	AN		822

## (2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 146 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:
- Gly Thr Ser Pro Cys Leu His Leu Leu Ala Asp Ile Arg Gly Glu Phe 1 5 10 15

Phe Asn Leu Arg Arg Val Glu Leu Leu Asn Val Ala Gln Gln Ala Gln

20

30

Leu Val Val Ala His Glu Gly Asp Gly Gln Ser Ser Ala Ala His Ala 35 40 45

Ala Ser Ala Ala Asp Ala Val Asn Val Leu Leu Thr Arg Glu Arg Gln 50 55 60

Ile Val Ala Glu Asp Glu Arg Asp Ala Leu His Ile Asn Ala Thr Arg 65 70 75 80

Pro Gln Val Arg Cys Asn Xaa His Ala Ala Val Ser Ile Thr Glu Cys 85 90 95

Arg Ile Ile Ser Ser Arg Ser Ala Trp Gly Met Ser Pro Cys Met Ala 100 105 110

Asp Thr Ala Lys Phe Ser Arg Val Ile Phe Leu Ala Ser Cys Ser Thr 115 120 125

Phe Leu Arg Val Leu Xaa Lys Thr Thr Ala Trp Ala Ile Val Lys Leu 130 135 140

Ser Tyr 145

### (2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 77 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

Ala Gln Gly Phe Arg Val Ile Phe Thr His Gly Gly Ala Gln Met Gly
1 5 10 15

Glu Val Asn Thr Arg Thr Glu Leu Leu Asp His Ile Asn Gln Ile Val 20 25 30

Val Ser Thr His Ala Val Xaa Thr Gly Ala His Gly Lys Thr Val Cys
35 40 45

His Ala Val Tyr Gly Ile Asn His Pro Leu His Ile Phe Asn Gly Gly 50 55 60

Asn Asn Ala Arg Gln Ala Glu Asp Pro Ala Arg Leu Ile 65 70 75

- (2) INFORMATION FOR SEQ ID NO:38:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 68 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania major
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

His Glu Pro Gln Trp Ser Ser Met Lys Ile Leu Gln Tyr Leu Thr Leu

1 10 15

Asp Gly Thr Gln Val Ser Gly Thr Leu Pro Pro Gln Trp Ser Ala Met 20 25 30

Ala Ser Val Arg Ile Leu Asn Leu Xaa Gly Thr Glu Val Ser Gly Thr 35 40 45

Leu Pro Pro Glu Trp Ile Ser Met Xaa Arg Leu Gln Thr Leu Asn Leu 50 60

Arg Arg Thr Lys

- (2) INFORMATION FOR SEQ ID NO:39:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 65 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania major
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

Ala Arg Glu Ala Gln Leu Ala His Arg Ile Cys Asp Tyr Leu Pro Glu 1 5 10 15

Gln Gly Gln Xaa Phe Val Gly Asn Ser Leu Val Val Arg Leu Ile Asp 20 25 30

Xaa Leu Xaa Gln Xaa Pro Ala Gly Tyr Pro Val Tyr Xaa Asn Arg Gly 35 40 45

Ala Xaa Gly Ile Xaa Xaa Leu Leu Ser Thr Ala Ala Gly Val Xaa Arg 50 55 60

Ala 65

- (2) INFORMATION FOR SEQ ID NO:40:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 78 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania major
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

Asp Asp His Ala Pro Lys Glu Asp Gly His Ala Pro Lys Asn Asp Asp 1 5 10 15

His Ala Pro Lys Glu Asp Gly His Ala Pro Lys Asn Asp Asp His Ala 20 25 30

Pro Lys Glu Asp Gly His Ala Pro Lys Asn Asp Gly Asp Val Gln Xaa 35 40 45

Lys Ser Glu Asp Gly Asp Asn Val Gly Glu Gly Gly Lys Gly Asn Glu 50 55 60

Asp Gly Asn Asp Asp Gln Pro Lys Glu His Ala Ala Gly Asn 65 70 75

- (2) INFORMATION FOR SEQ ID NO:41:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 169 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:

#### (A) ORGANISM: Leishmania major

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:
- Leu Gln Gln Arg Leu Asp Thr Ala Thr Gln Gln Arg Ala Glu Leu Glu

  1 5 10 15
- Ala Arg Val Ala Arg Leu Ala Ala Asp Arg Asp Glu Ala Arg Gln Gln 20 25 30
- Leu Ala Ala Asn Ala Glu Glu Leu Gln Gln Arg Leu Asp Thr Ala Thr
  35 40 45
- Gln Gln Arg Ala Glu Leu Glu Ala Arg Val Ala Arg Leu Ala Ala Asp 50 55 60
- Gly Asp Glu Ala Arg Gln Gln Leu Ala Ala Asn Ala Glu Glu Leu Gln 65 70 75 80
- Gln Arg Leu Asp Thr Ala Thr Gln Gln Arg Ala Glu Leu Glu Ala Gln 85 90 95
- Val Ala Arg Leu Ala Ala Asn Ala Glu Glu Leu Gln Gln Arg Leu Asp
  100 105 110
- Thr Ala Thr Gln Gln Arg Ala Glu Leu Glu Ala Arg Val Ala Arg Leu
  115 120 125
- Ala Ala Asp Arg Asp Glu Ala Arg Gln Gln Leu Ala Ala Asn Ala Glu 130 135 140
- Glu Leu Gln Gln Arg Leu Asp Thr Ala Thr Gln Gln Arg Ala Glu Leu 145 150 155 160
- Glu Ala Gln Val Ala Arg Leu Ala Ala 165

### (2) INFORMATION FOR SEQ ID NO:42:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 98 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

Ala Arg Xaa Ile Phe Val Lys Thr Leu Thr Gly Xaa Thr Ile Ala Leu 1 5 10 15

Glu Val Glu Pro Ser Asp Thr Ile Glu Asn Val Lys Ala Lys Ile Gln
20 25 30

Asp. Lys Glu Gly Ile Pro Pro Asp Gln Gln Arg Leu Ile Phe Ala Gly 35 40 45

Lys Gln Leu Glu Xaa Gly Arg Thr Leu Ser Asp Tyr Asn Ile Gln Lys 50 55 60

Glu Ser Thr Leu His Leu Val Leu Arg Leu Arg Gly Gly Met Xaa Ile 65 70 75 80

Phe Val Lys Thr Leu Thr Gly Xaa Thr Ile Ala Leu Glu Val Glu Pro 85 90 95

Asn Asp

## (2) INFORMATION FOR SEQ ID NO:43:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 39 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania major...
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

Leu Gln Gln Arg Leu Asp Thr Ala Thr Gln Gln Arg Ala Glu Leu Glu

1 10 15

Ala Arg Val Ala Arg Leu Ala Ala Asp Arg Asp Glu Ala Arg Gln Gln 20 25 30

Leu Ala Ala Asn Ala Glu Glu 35

#### (2) INFORMATION FOR SEO ID NO:44:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 600 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (	genomic	)
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#### (vi) ORIGINAL SOURCE:

(A) ORGANISM: Leishmania chagasi

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:

				•		
60	AGTTCGAGGA	GAGGCTGCCG	CATGGTGCGC	AGATCGAGCG	AGCGAGGAGG	CGGCCGCCTC
120	GCATCGCGTA	TCGCTAGAGA	AGCGAAGAAC	AACGTGTCGA	AAGGTGCGCG	TGAGGACCGC
180	CCGCGGACGA	GACAAGCTCG	CAAGCTTGGT	ACGACAAGGA	AACCAGATCA	CTCGCTTCGC
240	AGAACCCCAA	TTTGTCCACG	TGCCCTCGAC	CTGTGAAGGA	ATCGAGGAGG	CAAGAAGGCG
300	CGAACCCCAT	CAGAGTGTGA	CACGAAGCTG	AGGCTGCTCG	GAGGAGTTCG	TGCAGACCGT
360	ACGCGATGGA	GAAGAGGCGG	CTCTGGTGCA	GCGCCGCCGG	GTGTACCAGG	CATTCAAAAG
420	NCCACAANAA	GCGGGAACAT	AACAGGGAAA	TGAAAAGAAA	GTCGGCCGCG	TGACTTGTTA
480	NCNTCATGGG	CGGCNCACAT	CGAACACCGA	CGACACCGCT	AAGGGGGTNG	CCNAAGAAGA
540	CGTCTCNGCN	CTCCAAACNC	AGAAGGTTTT	CCAACAAACC	TTTCCTCTCC	CATGCTCAGC
600	TCTTTTGTTT	CAATTGNNGT	CCCCTTCCAC	ANCGAAAAAN	GGAAANGTTA	CCCAAAATAC

## (2) INFORMATION FOR SEQ ID NO:45:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1748 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania chagasi

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:

CI	AGTGGATC	CCCCGGGCTG	CAGGAATTCA	CGGAATACGT	ACCTCCTCCC	CCTTCTTGGT	60
AC	SAAGAACAA	CAACAACGTT	CAAGACGACG	CCGCGCCTTC	TTGTACCGCA	TTTGCTTCTG	120
AC	CACGTTCA	ATCCGTGCCT	TGCAAACATG	GAGGCGTACA	AGAAGCTGGA	AACGATCTTT	180
AC	CGAAGGTCT	ACCGCCTGGA	CCACTTCCTC	GGTCTGGGCA	ACTGGGACAT	GAACACAAAC	240

ATGCCCCCCA AGGGCGAGGA	ATCACGCGGT	' GAGGCGATGG	CGATGCTCTC	GGAGCTCCGC	300
TTTGGCTTCA TCACGGCACC	GGAGGTGAAA	AGCCTGATTG	AGAGTGCCAC	CAAGGGCAGC	360
GAGGAGCTGA ATGCGGTGCA	GCGCGCTAAC	TTGCGGGAGA	TGAGGCGTGC	GTGGAAGAGC	420
GCCACCGCCT TGCCGGCTGA	GTTTGTGGGC	CGCAAGATGC	GCCTCACGAC	ACACGCGCAC	480
AGCGTGTGGC GCGACAGCCG	CAAAGCAAAT	GACTTCGCCA	AGTTCCTACC	GGTGCTCAGG	540
GACCTGGTGG CGCTCGCCCG	TGAGGAGGC	TCATACCTCG	CCGCCGGCAC	CTCCCTCTCC	600
CCGTATGAGG CGCTCATGAA	CGAGTACGAG	CCAGGAATCA	CGACACAAAA	GCTGGATGAG	660
GTGTACGCAA ATGTAAAGTC	GTGGCTGCCG	CAGCTGCTAA	AGGACATTGT	GCAGAAGCAG	720
TCCGGCGAGT CGGTGATTGC	GTTCTCGCAT	AAGTTCCCGC	AGGACAAGCA	GGAAGCACTG	780
TGCAAGGAAT TCATGAAGAT	CTGGCACTTC	GACACCGATG	CCGGTCGCCT	CGACGTCAGC	840
CCCCACCCTT TCACGGGAAT	GACGAAGGAG	GACTGCCGAC	TCACAACAAA	CTACATCGAA	900
GACACGTTTG TTCAGAGCTT	GTATGGCGTC	ATCCACGAGA	GTGGGCATGG	CAAGTACGAG	960
CAGAACTGTG GCCCACGCGA	GCACATCACG	CAGCCGGTGT	GCAACGCCCG	CTCTCTTGGC	1020
CTGCATGAGA GCCAGAGCCT	CTTTGCGGAG	TTTCAGATCG	GCCACGCGAC	GCCCTTCATC	1080
GACTACCTCA CAACTCGCCT	TCCTGAGTTC	TTCGAGGCGC	AGCCAGCGTT	CTCGCAGGAC	1140
AACATGCGCA AGTCGCTGCA	GCAGGTGAAG	CCGGGCTACA	TTCGCGTCGA	TGCCGATGAG	1200
GTGTGCTACC CTCTGCACGT	GATCCTGCGC	TACGAGATCG	AGCGCGACTT	GATGGAGGC	1260
AAAATGGAGG TGGAAGACGT	GCCGCGCGCG	TGGAACGCAA	AGATGCAGGA	GTACTTGGGT	1320
CTCTCAACGG AGGGCCGTGA	CGACGTTGGG	TGCCTGCAGG	ACGTGCATTG	GTCCATGGTG	1380
CGCTCGGCTA CTCTCCGACG	TACTCGCTCG	GCGCCATGTA	TGCGGCGCAG	ATCATGGCGA	1440
GCATCCGAAA GGAGCTGGGA	GACGACAAGG	TGGATGAGTG	CCTGCGCACC	GGTGAGCTCG	1500
GCCCCCTCCT GGAAAAGCAG	CAGGAGAAGA	TCTGGGATCA	TGGGTGCCTG	TACGAGACGG	1560
ACGACCTCAT GACGCGTGCG	ACGGGCGAGA	CGCTGAACCC	CGAGTACCTG	CGCCGCCACC	1620
TGGAGGCGCG CTACATAAAC	GCCTGAGTCG	CGAGCGGTTG	ACACACGCGC	TCGCTAGCAC	1680
ATGACGCGTC TTTATTATTC	TTTGTTGTGC	ATTCGGAATT	CCGCGGAATT	CGATATCAAG	1740
CTTATCGA					1748

<sup>(2)</sup> INFORMATION FOR SEQ ID NO:46:

	122
(i)	SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 560 base pairs
	(B) TYPE: nucleic acid
	(C) STRANDEDNESS: double
	(D) TOPOLOGY: linear
ii)	MOLECULE TYPE: DNA (genomic)
vi)	ORIGINAL SOURCE:
•	(A) ORGANISM: Leishmania chagasi

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:46:

CGGAAGGAGG ATGGCCATAC ACAGAAAAAT GACGGCGATG GCCCTAAGGA GGACGGCCGT 60 ACACAGAAAA ACGACGACGG TGGCCCTAAG GAGGACGGCC ATACACAGAA AAATGACGGC 120 GATGGCCCTA AGGAGGACGG CCGTACACAG AAAAATAACG GCGATGGCCC TNAGGAGGAC 180 GGCCATACAC AGAAAAATGA CGGCGATGCC CCTNAGGAGG ACGGCCGTAC ACANAAAAAT 240 GACGGCNATG GCCCTNAGGA GGACGGCCGT ACACAGAAAA ATGACNGCCA TGGCCCTTAG 300 GANGACGCCG TACACAGAAA AATGACGCNA TGGCCCTNAG GGAGGACGGC CATACCCANA 360 AAAATTGACG GCNATNGCCC TTAGGANGAC GGCCGTNCCC ANAAANANTG ACNGCGGTNG 420 CCCTTAAGGA AGATGAAAAT CTGCCACCAA AACNATTGGG AATGCNCAGG AAAANAACNA 480 ANATHGACCC CACGTGGGGG ATGGANCTTA CNGCNATTAA NATTGTTACC ATTATCNACC 540 NAAGGACNNG TTGCCGNCAA 560

### (2) INFORMATION FOR SEQ ID NO:47:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 600 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania chaqasi
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:47:

60 CGTCCGAGAA ACCCGTACAT GTATGCTGCT GGTAGAAGGC GCAGAGCTGG TCCCTCTGAT GCACAAGCAT GAGGTCGTAC ATTGCCTGGT TCGTCATTTT CCAGAGCACA ACGAGCAGCG 120 WQ 98/35045 PCT/US98/03002

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TCATCATACA	GCATCCAATA	GCCGCCAGAG	TGAATGCGAT	GCGCACACCA	AGTCGAAAGT	180
GGTCGACCAG	TAGGGGAATG	TGACCCTGGC	TGGCGTGCAA	CATGATCGCC	ACGCCAGCGG	240
TGGGCCACAC	CACAACAGAG	GCGACGAAAG	AGAACATGAA	CTTGCTCACG	AAGCTNACAA	300
TAAGGGCGTC	GCTNGTGATG	CTAAGAACCA	CGCCNAGGTA	GACGGCGAAG	ANCAAACTAA	360
ACACAAGCGT	GACGATCCCG	AAAAGAAGGA	TCTCTGCGGA	ATTTTCGTGA	GATAGANAAT	420
GCCCGTACTG	GAAAAANAAG	CCGGCAGGCG	CGCGATAACG	CTGCAACTTG	CCGCTCCTCG	480
CGGGCGCGTT	TTCGCTCCTT	CTCCGACTTG	ATGGCGCNGT	CNGNCTTGAC	AAAACGGTTA	540
AGCTCCTCAT	GCCCCAGCCG	ATTCCCAGCT	CACGGTCCAC	TTCCGGCCAT	GCCCACGGAC	600

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#### (2) INFORMATION FOR SEQ ID NO:48:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1053 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania chagasi
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:48:

GGGAAAAAAG TGGAGCTCCA CCGCGGTGGC GGCCGCTCTA GAACTAGTGG ATCCCCCGGG 60 CTGCAGGAAT TCCGCGGAAT TCCGCGGAAT TCCGTCCGAC GCGGCACCCG 120 CACAGGGGTC GACAGTGACG CAACCTCCTC CACCACTGCG GCCTACGACG GCGCCGGCTC 180 CGCGCCAGTG ATGGTTGACG CCAATGTGAG CCACCCTCCG TACGCGGGGC ATGACCAAGT GTACATGCAC GTCGGCAAGC CCATCGTGGG CAACACCCTC GACGGATACA ACGGGTGCGT 300 GTTCGCCTAC GGGCANACGG GCAGCGGCAA AACCTTCACG ATGCTCGGNT ACGCGCCGAG 360 CACGANCGAC ATCCGCGCTC GCAAAGGGTC CGTCCCCTGC GGGGCCAGCA GCATGGAGAA 420 CAGCACTCCT CTTGACAGCG CTGTGGAGCC GTTTGAGAGC GATGACGGCG ACGACGTGGT 480 GGACAAGACG GGGCTGGATC CGAACGAGCT GCAAGGCATC ATCCCGCGCG CGTGCACGGA 540 CCTGTTCGAT GGTCTCCGTG CGAAGCGCGC CAAGGACTCC GACTTCACGT ACCGCGTGGA 600

GGTGTCTTAC	TACGAGATCT	ACAACGAGAA	GGTGTTCGAT	CTCATCCGGC	CGCAGCGCAA	660
CACGGACCTG	AGGATACGTA	ACTCGCCCAA	CTCCGGTCCA	TTTATCGAAG	GCCTGACGTG	720
GAAGATGGTG	TCCAAGGAGG	AAGACGTCGC	CCGCGTGATT	CGCAAGGGCA	TGCAGGAGCG	780
CCACACGGCT	GCGACCAAGT	TCAACGACCG	CAGCAGCCGC	AGCCACGCCA	TCCTCACCTT	840
CAACATTGTG	CAGCTGTCGA	TGGACGACTC	CGACAACGCG	TTCCAGATGC	GCAGCAAGCT	900
GAACCTGGTG	GACCTTGCTG	GGTCGGAGCG	CACTGGTGCG	GCCGGAGCCG	AGGGCAATGA	960
GTTCCACGAC	GGTGTGAAGA	TCAACCACTC	GCTGACGGTG	CTGGGGCGCG	TGATCGACCG	1020
TCTGGCGGAC	CTCTCGCAGA	ACAAGGGAGG	GGG			1053

#### (2) INFORMATION FOR SEQ ID NO:49:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 136 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania chagasi
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:49:
- Gly Arg Leu Ser Glu Glu Glu Ile Glu Arg Met Val Arg Glu Ala Ala 1 5 10 15
- Glu Phe Glu Asp Glu Asp Arg Lys Val Arg Glu Arg Val Glu Ala Lys
  20 25 30
- Asn Ser Leu Glu Ser Ile Ala Tyr Ser Leu Arg Asn Gln Ile Asn Asp 35 40 45
- Lys Asp Lys Leu Gly Asp Lys Leu Ala Ala Asp Asp Lys Lys Ala Ile 50 55 60
- Glu Glu Ala Val Lys Asp Ala Leu Asp Phe Val His Glu Asn Pro Asn 65 70 75 80
- Ala Asp Arg Glu Glu Phe Glu Ala Ala Arg Thr Lys Leu Gln Ser Val 85 90 95
- Thr Asn Pro Ile Ile Gln Lys Val Tyr Gln Gly Ala Ala Gly Ser Gly
  100 105 110
- Ala Glu Glu Ala Asp Ala Met Asp Asp Leu Leu Val Gly Arg Val Lys

115

120

125

Arg Lys Thr Gly Lys Ala Gly Thr 130 135

- (2) INFORMATION FOR SEQ ID NO:50:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 510 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania chagasi
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:50:

Tyr Leu Leu Pro Leu Leu Gly Arg Arg Thr Thr Thr Thr Phe Lys Thr

1 5 10 15

Thr Pro Arg Leu Leu Val Pro His Leu Leu Leu Ser Thr Phe Asn Pro 20 25 30

Cys Leu Ala Asn Met Glu Ala Tyr Lys Lys Leu Glu Thr Ile Phe Thr 35 40 45

Lys Val Tyr Arg Leu Asp His Phe Leu Gly Leu Gly Asn Trp Asp Met 50 55 60

Asn Thr Asn Met Pro Pro Lys Gly Glu Glu Ser Arg Gly Glu Ala Met 65 70 75 80

Ala Met Leu Ser Glu Leu Arg Phe Gly Phe Ile Thr Ala Pro Glu Val 85 90 95

Lys Ser Leu Ile Glu Ser Ala Thr Lys Gly Ser Glu Glu Leu Asn Ala 100 105 110

Val Gln Arg Ala Asn Leu Arg Glu Met Arg Arg Ala Trp Lys Ser Ala 115 120 125

Thr Ala Leu Pro Ala Glu Phe Val Gly Arg Lys Met Arg Leu Thr Thr 130 135 140

His Ala His Ser Val Trp Arg Asp Ser Arg Lys Ala Asn Asp Phe Ala 145 150 155 160

Lys Phe Leu Pro Val Leu Arg Asp Leu Val Ala Leu Ala Arg Glu Glu 165. 170 175

							-								
Gly	Ser	Tyr	Leu 180	Ala	Ala	Gly	Thr	Ser 185	Leu	Ser	Pro	Tyr	Glu 190	Ala	Leu
Met	Asn	Glu 195	Tyr	Glu	Pro	Gly	Ile 200	Thr	Thr	Gln	Lys	Leu 205	Asp	Glu	Val
Tyr	Ala 210	Asn	Val	Lys	Ser	Trp 215	Leu	Pro	Gln	Leu	Leu 220	Lys	Asp	Ile	Val
Gln 225	Lys	Gln	Ser	Gly	Glu 230	Ser	Val	Ile	Ala	Phe 235	Ser	His	Lys	Phe	Pro 240
Gln	Asp	Lys	Gln	Glu 245	Ala	Leu	Cys	Lys	Glu 250	Phe	Met	Lys	Ile	Trp 255	His
Phe	Asp	Thr	Asp 260	Ala	Gly	Arg	Leu	Asp 265	Val	Ser	Pro	His	Pro 270	Phe	Thr
Gly	Met	Thr 275	Lys	Glu	Asp	Cys	Arg 280	Leu	Thr	Thr	Asn	Tyr 285	Ile	Glu	Asp
Thr	Phe 290	Val	Gln	Ser	Leu	Tyr 295	Gly	Val	Ile	His	Glu 300	Ser	Gly	His	Gly
Lys 305	Tyr	Glu	Gln	Asn	Cys 310	Gly	Pro	Arg	Glu	His 315	Ile	Thr	Gln	Pro	Val 320
Cys	Asn	Ala	Arg	Ser 325	Leu	Gly	Leu	His	Glu 330	Ser	Gln	Ser	Leu	Phe 335	Ala
Glu	Phe	Gln	Ile 340	Gly	His	Ala	Thr	Pro 345	Phe	Ile	Asp	Tyr	Leu 350	Thr	Thr
Arg	Leu	Pro 355	Glu	Phe	Phe	Glu	Ala 360	Gln	Pro	Ala	Phe	Ser 365	Gln	Asp	Asn
Met	Arg 370	Lys	Ser	Leu	Gln	Gln 375	Val	Lys	Pro	Gly	Tyr 380	Ile	Arg	Val	Asp
Ala 385	Asp	Glu	Val	Cys	Tyr 390	Pro	Leu	His	Val	Ile 395	Leu	Arg	Tyr	Glu	Ile 400
Glu	Arg	Asp	Leu	Met 405	Glu	Gly	Lys	Met	Glu 410	Val	Glu	Asp	Val	Pro 415	Arg
Ala	Trp	Asn	Ala 420	Lys	Met	Gln	Glu	Tyr 425		Gly	Leu	Ser	Thr 430	Glu	Gly
Arg	Asp	Asp 435		Gly	Cys	Leu	Gln 440		Val	His	Trp	Ser 445	Met	Val	Arg
Ser	Ala 450		Leu	Arg	Arg	Thr 455	_	Ser	Ala	Pro	Cys 460	Met	Arg	Arg	Arg

Ser Trp Arg Ala Ser Glu Arg Ser Trp Glu Thr Thr Arg Trp Met Ser

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465

470

475

480

Ala Cys Ala Pro Val Ser Ser Ala Pro Ser Trp Lys Ser Ser Arg Arg
485 490 495

Arg Ser Gly Ile Met Gly Ala Cys Thr Arg Arg Thr Thr Ser 500 505 510

- (2) INFORMATION FOR SEQ ID NO:51:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 107 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania chagasi
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:

Gly Arg Arg Met Ala Ile His Arg Lys Met Thr Ala Met Ala Leu Arg 1 5 10 15

Arg Thr Ala Val His Arg Lys Thr Thr Val Ala Leu Arg Arg Thr 20 25 30

Ala Ile His Arg Lys Met Thr Ala Met Ala Leu Arg Arg Thr Ala Val 35 40 45

His Arg Lys Ile Thr Ala Met Ala Leu Arg Arg Thr Ala Ile His Arg 50 55 60

Lys Met Thr Ala Met Pro Leu Arg Arg Thr Ala Val His Xaa Lys Met 65 70 75 80

Thr Ala Met Ala Leu Arg Arg Thr Ala Val His Arg Lys Met Thr Ala 85 90 95

Met Ala Leu Arg Xaa Thr Pro Tyr Thr Glu Lys
100 105

- (2) INFORMATION FOR SEQ ID NO:52:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 63 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide

- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Leishmania chagasi
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:

Val Arg Glu Thr Arg Thr Cys Met Leu Leu Val Glu Gly Ala Glu Leu

1 5 10 15

Val Pro Leu Met His Lys His Glu Val Val His Cys Leu Val Arg His
20 25 30

Phe Pro Glu His Asn Glu Gln Arg His His Thr Ala Ser Asn Ser Arg 35 40 45

Gln Ser Glu Cys Asp Ala His Thr Lys Ser Lys Val Val Asp Gln 50 55 60

- (2) INFORMATION FOR SEQ ID NO:53:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 324 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (vi) ORIGINAL SOURCE:
    - (A) ORGANISM: Leishmania chagasi
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:

Phe Arg Gly Ile Pro Arg Asn Ser Val Arg Arg Gly Thr Arg Thr Gly
1 5 10 15

Val Asp Ser Asp Ala Thr Ser Ser Thr Thr Ala Ala Tyr Asp Gly Ala 20 25 30

Gly Ser Ala Pro Val Met Val Asp Ala Asn Val Ser His Pro Pro Tyr 35 40 45

Ala Gly His Asp Gln Val Tyr Met His Val Gly Lys Pro Ile Val Gly 50 55 60

Asn Thr Leu Asp Gly Tyr Asn Gly Cys Val Phe Ala Tyr Gly Xaa Thr 65 70 75 80

Gly Ser Gly Lys Thr Phe Thr Met Leu Gly Tyr Ala Pro Ser Thr Xaa 85 90 95 WO 98/35045 PCT/US98/03002

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								127							
Asp	Ile	Arg	Ala 100	Arg	Lys	Gly	Ser	Val 105	Pro	Cys	Gly	Ala	Ser 110	Ser	Met
Glu	Asn	Ser 115	Thr	Pro	Leu	Asp	Ser 120	Ala	Val	Glu	Pro	Phe 125	Glu	Ser	Asp
Asp	Gly 130	Asp	Asp	Val	Val	Asp 135	Lys	Thr	Gly	Leu	Asp 140	Pro	Asn	Glu.	Leu
Gln 145	Gly	Ile	Ile	Pro	Arg 150	Ala	Cys	Thr	Asp	Leu 155	Phe	Asp	Gly	Leu	Arg
Ala	Lys	Arg	Ala	Lys 165	Asp	Ser	Asp	Phe	Thr 170	Tyr	Arg	Val	Glu	Val 175	Ser
Tyr	Tyr	Glu	Ile 180	Tyr	Asn	Glu	Lys	Val 185	Phe	Asp	Leu	Ile	Arg 190	Pro	Gln
Arg	Asn	Thr 195	Asp	Leu	Arg	Ile	Arg 200	Asn	Ser	Pro	Asn	Ser 205	Gly	Pro	Phe
Ile	Glu 210	Gly	Leu	Thr	Trp	Lys 215	Met	Val	Ser	Lys	Glu 220	Glu	Asp	Val	Ala
Arg 225	Val	Ile	Arg	Lys	Gly 230	Met	Gln	Glu	Arg	His 235	Thr	Ala	Ala	Thr	Lys 240
Phe	Asn	Asp	Arg	Ser 245	Ser	Arg	Ser	His	Ala 250	Ile	Leu	Thŗ	Phe	Asn 255	
Val	Glń	Ĺeú	Ser 260	Met	Asp	Asp	Ser	Asp 265	Asn	Ala	Phe	Glñ	Met 270	Arg	Ser
Lys	Leu	Asn 275	Leu	Val	Asp	Leu	Ala 280	Gly	Ser	Glu	Arg	Thr 285	Gly	Ala	Ala
Gly	Ala 290	Glu	Gly	Asn	Glu	Phe 295	His	Asp	Gly	Val	Lys 300	Ile	Asn	His	Ser
Leu 305	Thr	Val	Leu	Gly	Arg 310	Val	Ile	Asp	Arg	Leu 315	Ala	Asp	Leu	Ser	Gln 320

Asn Lys Gly Gly

# (2) INFORMATION FOR SEQ ID NO:54:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1585 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:

AAAGCTGGAG	CTCCACCGCG	GTGGCGGCCG	CTCTAGAACT	AGTGGATCCC	CCGGGCTGCA	60
GGAATTCGGC	ACGAGTGCTG	CCCGACATGA	CATGCTCGCT	GACCGGACTT	CAGTGCACAG	120
ACCCGAACTG	CAAGACCTGC	ACAACTTACG	GTCAGTGCAC	AGACTGCAAC	GACGGCTACG	180
GTCTCACCTC	CTCCAGCGTT	TGCGTGCGCT	GCAGTGTAGC	GGGCTGCAAG	AGCTGCCCCG	240
TCGACGCTAA	CGTCTGCAAA	GTGTGTCTCG	GCGGCAGCGA	GCCGATCAAC	AATATGTGCC	300
CCTGCACCGA	CCCCAACTGC	GCCAGCTGCC	CCAGCGACGC	TGGCACGTGC	ACTCAGTGCG	360
CGAACGGCTA	CGGTCTCGTG	GACGGCGCCT	GTGTGAGATG	CCAGGAGCCC	AACTGCTTCA	420
GCTGCGACAG	CGACGCGAAT	AAGTGCACAC	AATGTGCGCC	GAACTACTAC	CTCACCCGC	480
TCTTGACCTG	CTCCCCGGTG	GCCTGCAACA	TCGAGCACTG	CATGCAGTGC	GACCCACAGA	540
CGCCGTCGCG	CTGCCAGGAG	TGCGTGTCCC	CCTACGTGGT	TGACAGCTAC	GACGGCCTCT	600
GCAGGCTCTC	CGATGCCTGC	TCCGTGCCCA	ACTGCAAGAA	GTGCGAGACC	GGTACCTCCA	660
GGCTCTGCGC	CGAGTGCGAC	ACCGGCTACA	GTCTCTCCGC	CGACGCGACG	AGCTGCAGCA	720
GTCCAACCAC	GCAGCCGTGC	GAGGTGGAGC	ACTGCAACAC	ATGTGTGAAC	GGCGATAGCA	780
CCCGCTGTGC	CTACTGCAAC	ACCGGCTACT	ACGTCTCCGA	TGGCAAGTGC	AAGGCCATGC	840
AGGGCTGCTA	CGTGTCGAAC	TGCGCGCAGT	GCATGCTGCT	TGACAGCACC	AAGTGCTCCA	900
CGTGCGTGAA	AGGGTACCTG	CTCACGTCGT	CCTACAGTTG	CGTCTCGCAG	AAAGTCATCA	960
ACAGTGCGGC	CGCGCCCTAC	TCTCTGTGGG	TGGCCGCCGC	CGTGCTCCTC	ACCTCTTTTG	1020
CCATGCACCT	AGCATAGTGC	GCAGCGGCAT	GCGAACAACC	CCACTCTCAT	TCTCCAACAT	1080
GTGCATACAC	ACACACACAG	ACAGCGGGGC	AGCACCCCT	CCCCACACAC	ACACACGCAC	1140
TTCCCCCTTG	TCTTGTTCTT	CTTTCCTCGN	TTCGCATTTC	TTTCTCTCGT	GCGCTGGCGC	1200
CGGCCTCCTG	CACGTCGCTC	CCCTCCCCCT	AACCTCTATT	CTCTCTCTCT	CTCTCTCTCG	1260
CCGGCATCAT	TGCTTCTTAC	CCTTTTCTGA	TCCTTGCTCG	CGTGGGCGGA	CACTGCCACA	1320
GTCCCACAGC	GCAGACACAC	GTGTTTAAAC	GGCGCAGGCA	TCCCTCCCTA	TCACTTCATT	1380
TCTCCTAAAG	CCACTCACCA	AGTCGCACAC	CGCCCTCCCC	CATCGGCCGC	CCTTCCGGGC	1440
GCAGCTGTGC	GGAATGGGTG	TGTGCTCGAC	CTCGTTCCTG	GCAGCTCACT	CGCATGTGTA	1500

CAGCCACTCC AACCACGAAA GCTCTCTTCT GCGCACATAA AAAAAAAAA AAAAAAAAA 1560
CTCGAGGGGG GGCCCGGTAC CCAAA 1585

- (2) INFORMATION FOR SEQ ID NO:55:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 320 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: protein
  - (v) FRAGMENT TYPE: internal
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:
  - Val Leu Pro Asp Met Thr Cys Ser Leu Thr Gly Leu Gln Cys Thr Asp 1 5 10 15
  - Pro Asn Cys Lys Thr Cys Thr Thr Tyr Gly Gln Cys Thr Asp Cys Asn 20 25 30
  - Asp Gly Tyr Gly Leu Thr Ser Ser Ser Val Cys Val Arg Cys Ser Val 35 40 45
  - Ala Gly Cys Lys Ser Cys Pro Val Asp Ala Asn Val Cys Lys Val Cys 50 55 60
  - Leu Gly Gly Ser Glu Pro Ile Asn Asn Met Cys Pro Cys Thr Asp Pro 65 70 75 80
  - Asn Cys Ala Ser Cys Pro Ser Asp Ala Gly Thr Cys Thr Gln Cys Ala 85 90 95
  - Asn Gly Tyr Gly Leu Val Asp Gly Ala Cys Val Arg Cys Gln Glu Pro 100 105 110
  - Asn Cys Phe Ser Cys Asp Ser Asp Ala Asn Lys Cys Thr Gln Cys Ala 115 120 125
  - Pro Asn Tyr Tyr Leu Thr Pro Leu Leu Thr Cys Ser Pro Val Ala Cys 130 135 140
  - Asn Ile Glu His Cys Met Gln Cys Asp Pro Gln Thr Pro Ser Arg Cys 145 . 150 155 160
  - Gln Glu Cys Val Ser Pro Tyr Val Val Asp Ser Tyr Asp Gly Leu Cys 165 170 175

Arg Leu Ser Asp Ala Cys Ser Val Pro Asn Cys Lys Lys Cys Glu Thr 180 185 190

Gly Thr Ser Arg Leu Cys Ala Glu Cys Asp Thr Gly Tyr Ser Leu Ser 195 200 205

Ala Asp Ala Thr Ser Cys Ser Ser Pro Thr Thr Gln Pro Cys Glu Val 210 215 220

Glu His Cys Asn Thr Cys Val Asn Gly Asp Ser Thr Arg Cys Ala Tyr 225 230 235 240

Cys Asn Thr Gly Tyr Tyr Val Ser Asp Gly Lys Cys Lys Ala Met Gln 245 250 255

Gly Cys Tyr Val Ser Asn Cys Ala Gln Cys Met Leu Leu Asp Ser Thr 260 265 270

Lys Cys Ser Thr Cys Val Lys Gly Tyr Leu Leu Thr Ser Ser Tyr Ser 275 280 285

Cys Val Ser Gln Lys Val Ile Asn Ser Ala Ala Ala Pro Tyr Ser Leu 290 295 300

Trp Val Ala Ala Ala Val Leu Leu Thr Ser Phe Ala Met His Leu Ala 305 310 315 320

### (2) INFORMATION FOR SEQ ID NO:56:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 14 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: peptide
  - (v) FRAGMENT TYPE: internal
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:

Pro Lys Glu Asp Gly His Ala Pro Lys Asn Asp Asp His Ala 1 5 10

### (2) INFORMATION FOR SEQ ID NO:57:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 7 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

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(V) FRAGMENT TYPE: Internal	
·	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:57:	
Pro Lys Glu Asp Gly His Ala 1 5	
(2) INFORMATION FOR SEQ ID NO:58:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 7 amino acids</li> <li>(B) TYPE: amino acid</li> <li>(C) STRANDEDNESS:</li> <li>(D) TOPOLOGY: linear</li> </ul>	·
(ii) MOLECULE TYPE: peptide	
(v) FRAGMENT TYPE: internal	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:58:	
Pro Lys Asn Asp Asp His Ala 1 5	
(2) INFORMATION FOR SEQ ID NO:59:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 264 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
•	
	·
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:59:	
ATGCACCATC ATCACCATCA CATGGGAAGC TCCTGCACGA AGGACTCCGC AAAGGAGCCC	60
CAGAAGCGTG CTGATAACAT CGATACGACC ACTCGAAGCG ATGAGAAGGA CGGCATCCAT	120
GTCCAGGAGA GCGCCGGTCC TGTGCAGGAG AACTTCGGGG ATGCGCAGGA GAAGAACGAA	180
GATGGACACA ACGTGGGGGA TGGAGCTAAC GACAATGAGG ATGGTAACGA TGATCAGCCG	240

AAGGAGCAGG	TTGCCGGCAA	CTAG
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264

#### (2) INFORMATION FOR SEQ ID NO:60:

### (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 744 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:

60 ATGGGAGCCT ACTGCACGAA GGACTCCGCA AAGGAGCCCC AGAAGCGTGC TGATAACATC CATAAAACCA CTGAGGCCAA TCACAGAGGC GCCGCCGGTG TGCCCCCGAA GCACGCCGGC 120 GGTGCGATGA ACGACTCTGC CCCGAAGGAG GATGGCCATA CACAGAAAAA TGACGGCGAT 180 GGCCTAAGG AGGACGCCG TACACAGAAA AACGACGACG GTGGCCCTAA GGAGGACGGC 240 CATACACAGA AAAATGACGG CGATGGCCCT AAGGAGGACG GCCGTACACA GAAAAATAAC 300 GGCGATGGCC CTAAGGAGGA CGGCCATACA CAGAAAAATG ACGGCGATGC CCCTAAGGAG 360 GACGGCCGTA CACAGAAAAA TGACGGCGAT GGCCCTAAGG AGGACGGCCG TACACAGAAA 420 AATGACGGCG ATGGCCCTAA GGAGGACGGC CGTACACAGA AAAATGACGG CGATGGCCCT 480 AAGGAGGACG GCCGTACACA GAAAAATGAC GGCGATGGCC CTAAGGAGGA CGGCCATACA 540 CAGAAAAATG ACGGCGATGG CCCTAAGGAG GACGGCCGTA CACAGAAAAA TGACGGCGGT 600 GGCCCTAAGG AGGATGAGAA TCTGCAGCAA AACGATGGGA ATGCGCAGGA GAAGAACGAA 660 GATGGACACA ACGTGGGGGA TGGAGCTAAC GGCAATGAGG ATGGTAACGA TGATCAGCCG 720 AAGGAGCAGG TTGCCGGCAA CTAG 744

### (2) INFORMATION FOR SEQ ID NO:61:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 80 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

47.0 12.

Met Gly Ser Ser Cys Thr Lys Asp Ser Ala Lys Glu Pro Gln Lys Arg

1 10 15

Ala Asp Asn Ile Asp Thr Thr Thr Arg Ser Asp Glu Lys Asp Gly Ile .
20 25 30

His Val Gln Glu Ser Ala Gly Pro Val Gln Glu Asn Phe Gly Asp Ala 35 40 45

Gln Glu Lys Asn Glu Asp Gly His Asn Val Gly Asp Gly Ala Asn Asp 50 55 60

Asn Glu Asp Gly Asn Asp Asp Gln Pro Lys Glu Gln Val Ala Gly Asn 65 70 75 80

# (2) INFORMATION FOR SEQ ID NO:62:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 247 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:

Met Gly Ala Tyr Cys Thr Lys Asp Ser Ala Lys Glu Pro Gln Lys Arg

1 10 15

Ala Asp Asn Ile His Lys Thr Thr Glu Ala Asn His Arg Gly Ala Ala
20 25 30

Gly Val Pro Pro Lys His Ala Gly Gly Ala Met Asn Asp Ser Ala Pro 35 40 45

Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp Gly Pro Lys Glu 50 55 60

Asp Gly Arg Thr Gln Lys Asn Asp Asp Gly Gly Pro Lys Glu Asp Gly 65 70 75 80

His Thr Gln Lys Asn Asp Gly Asp Gly Pro Lys Glu Asp Gly Arg Thr 85 90 95

Gln Lys Asn Asn Gly Asp Gly Pro Lys Glu Asp Gly His Thr Gln Lys
100 105 110

Asn Asp Gly Asp Ala Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp 115 120 125

Gly Asp Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp 130 135 140

Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp Gly Pro
145 150 155 160

Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp Gly Pro Lys Glu 165 170 175

Asp Gly His Thr Gln Lys Asn Asp Gly Asp Gly Pro Lys Glu Asp Gly 180 185 190

Arg Thr Gln Lys Asn Asp Gly Gly Gly Pro Lys Glu Asp Glu Asn Leu 195 200 205

Gln Gln Asn Asp Gly Asn Ala Gln Glu Lys Asn Glu Asp Gly His Asn 210 215 220

Val Gly Asp Gly Ala Asn Gly Asn Glu Asp Gly Asn Asp Asp Gln Pro 225 230 235 240

Lys Glu Gln Val Ala Gly Asn 245

#### (2) INFORMATION FOR SEQ ID NO:63:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 14 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear
- (ix) FEATURE:
  - (A) NAME/KEY: Modified-site
  - (B) LOCATION: 6
- (D) OTHER INFORMATION: /note= "Xaa can be either His or Arg"
  - (ix) FEATURE:
    - (A) NAME/KEY: Modified-site
    - (B) LOCATION: 12
- (D) OTHER INFORMATION: /note= "Xaa can be either Gly or Asp"
- (ix) FEATURE:
  - (A) NAME/KEY: Modified-site
  - (B) LOCATION: 13
- (D) OTHER INFORMATION: /note= "Xaa can be either Asp or Gly"
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:63:

Pro Lys Glu Asp Gly Xaa Thr Gln Lys Asn Asp Xaa Xaa Gly
1 5 10

- (2) INFORMATION FOR SEQ ID NO:64:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 7 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (ix) FEATURE:
    - (A) NAME/KEY: Modified-site
    - (B) LOCATION: 6
    - (D) OTHER INFORMATION: /note= "Xaa can be either His or

Arg"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:64:

Pro Lys Glu Asp Gly Xaa Thr 1 5

- (2) INFORMATION FOR SEQ ID NO:65:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 7 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (ix) FEATURE:
    - (A) NAME/KEY: Modified-site
    - (B) LOCATION: 5
    - (D) OTHER INFORMATION: /note= "Xaa can be either Gly or

"qaA

- (ix) FEATURE:
  - (A) NAME/KEY: Modified-site
  - (B) LOCATION: 6
  - (D) OTHER INFORMATION: /note= "Xaa can be either Asp or

Gly"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:65:

Gln Lys Asn Asp Xaa Xaa Gly

(2) INFORMATION FOR SEQ ID NO:66:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 17 amino acids
  - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:
- Gly Cys Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp 1 5 10 15

Gly

- (2) INFORMATION FOR SEQ ID NO:67:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 31 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:
  - Gly Cys Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp
  - Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp Gly 20 25 30
- (2) INFORMATION FOR SEQ ID NO:68:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 45 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:68:
  - Gly Cys Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp 1 5 10 15

Gly Pro Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp Gly Pro
20 25 30

Lys Glu Asp Gly Arg Thr Gln Lys Asn Asp Gly Asp Gly 35 40 45

- (2) INFORMATION FOR SEQ ID NO:69:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 17 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:69:

Gly Cys Gly Pro Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp 1 5 10 15

Gly

- (2) INFORMATION FOR SEQ ID NO:70:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 31 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:70:

Gly Cys Gly Pro Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp 1 5 10 15

Gly Pro Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp Gly 20 25 30

- (2) INFORMATION FOR SEQ ID NO:71:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 45 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear

(xi)	SEQUENCE	DESCRIPTION:	SEQ	ID	NO:71	:
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Gly Cys Gly Pro Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp 1 5 10 15

Gly Pro Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp Gly Pro
20 25 30

Lys Glu Asp Gly His Thr Gln Lys Asn Asp Gly Asp Gly 35 40 45

### (2) INFORMATION FOR SEQ ID NO:72:

#### (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 664 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:72:

60 GCTGCAGGAA TTCGGCACGA GATTGCTTCC CAGCCCACCT TCGCTATCCA GCCACTCTCG CTCTTCTACA TCTCCCACCC CCTCACACCG CCATGGCTTC TTCCCGCAAG GCTTCCAACC 120 CGCACAGTC GCACCGCAAG CCGAAGCGCT CGTGGAACGT GTACGTGGGC CGCTCGCTGA 180 AGGCGATCAA CGCCCAGATG TCGATGTCGC ACCGCACGAT GAAGATCGTG AACTCGTACG 240 300 TGAACGACGT GATGGAGCGC ATCTGCACTG AGGCCGCGTC GATTGTTCGC GCGAACAAGA 360 AGCGCACGTT GGGTGCGCGC GAGGTGCAGA CGGCGGTGCG CATTGTGCTG CCGGCGGAGC TCGCGAAGCA TGCCATGGCT GAGGGCACGA AGGCCGTGTC GAGCGCGTCC CGCTAAAGCG 420 GCTTGCCGGA TGCCGTGTGA GTAGGAGGGT GGCTTGCCGC AAACGCTGAC CTCGGCGATT 480 GCGGCGTGGC GCTCCCTTC TCCTCCTTGT CCGGCGGTGT GTGTCATGCA TTTGCGTGAC 540 TCCTCCCTCT TATAGATGCA AGCTTTTTT TTCTCTTGAC GTTTTATTTT CTCCTCCCC 600 660 TCCCTTAACG TGAAGTGTAT ATGANAGCGT ACTGGACATG ANANAAAAA AAAANAAACT 664 **CGAG** 

#### (2) INFORMATION FOR SEQ ID NO:73:

## (i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 1432 base pairs(B) TYPE: nucleic acid(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:73:

GATGAAGAAG AGGAGGACAC CACCATCAAC AACTCCGACG TGGTGGTGCG CTACAAGAAG 60 GCCGCAACGT GGTGCAATGA AACGTTGCGC GTGCTTATCG ATGCCACAAA ACCTGGCGCC 120 AAGGTGTGCG ACCTGTGCCG CCTCGGTGAT GACACCATCA CCGCCNAGGT CAAGACAATG 180 TTCAAAGGCA CGGAAAAAGG CATCGCTTTC CCGACCTGCA TCTCGGTCAA CAACTGCGTA 240 TGCCACACA GCCCTGGCGT GTCGGACGAG ACGACGCAGC AAGAGATCGC GATGGGTGAC 300 GTCGTGCACT ACGACCTGGG CATCCACGTG GACGGCTACT GCGCCGTCGT CGCGCACACC 360 ATTCAGGTGA CAGAGGACAA TGAGCTTGGC AAGGACGAGA AGGCGGCGC CGTCATTACA 420 GCGGCGTACA ACATCCTGAA CACGGCGCTG CGCCAGATGC GTCCCGGTAC GACCATCTAC 480 CAGGTGACAG ACGTAGTTGA GAAGGCTGCG GAGCACTACA AGGTGACTCC GGTAGACGGC 540 GTCCTCTCGC ATATGATGAA GCGCTACATC ATAGACNGAT ACCGCTGTAT CCCGCAGCGC 600 AGGGTCGCGG AGCACATGGT GCACGACTAC GATCTCGAGA AAGCGCAGGT GTGGACGCTA 660 GACATTGTCA TGACCTCCGG CAAGGGCAAG CTGAAGGAGC GCGATGCGCG GCCGTGCGTG 720 TTCAAGGTGG CTCTGGACTC CAACTACTCT GTGAAAATGG AAAGCGCGAA GGAGGTTCAG 780 AAGGAAATCG ACTCCNAGTA TGCCACCTTC CCCTTTGCCA TCCGCAACCT GGAGGCCAAG 840 AAGGCCCGCC TCGGTCTCAA CGAGATGGCG AAGCACGGTG CTGTCATCCC GTACCCTATT 900 CTCTTCGAAA AGGAAGGCGA GGTCGTCGCC CATTTCAAGA TTACGGTGCT CATCAGCAAC 960 AAGAAGATTG AGCCGATTAC CGGCCTGAAG CCGCAGAAGG CCCCGGCGCT CGAGCCATAC 1020 ACGGACGAGA TGCTGCTTGC GACGAACAAG CTCTTCGCTG TCGCTAGAGA AGAAGGCGGC 1080 GAAGTAGACG GCCGTGGCAT CCGTGACGCT GTACTGCGAG CTTTCGTAGG CGTACGCCTC 1140 TTGTGAGGCG TACACGTGTG CTGTTTGCGG ACGAGGAGGC ACCCATTCTG TTCCCCTTCT 1200 TCGCTAATCT TCGCGTTTCC TCTGACGCTG GCTTCTYTGC CGGAGTGTGG TGAGGCGCGT 1260 GGGGGAGAAA CGGCCCACTY GCATGCCTGT GCATACGCGA GCACGGTAGG GAGCGCGGTG 1320

TGTGTGTGTG	TGGGGGGGCG	TGTTACGAGT	ACAAAAGAGG	CTCGATCTTT	GCGATCTTTT	1380
CTTTCTGTAA	ACAGGAACAT	AAGTAACCAA	АААААААА	AAAAAACTCG	AG	1432

### (2) INFORMATION FOR SEQ ID NO:74:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 873 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:74:

CTTTATTGTC	ATCACTGTAA	AGCACTGTTT	TTTCTTTCAC	TTTTTCTTGA	GTGTTTTCTT	60
CTATTCACCA	TGAGCATTAT	CAAGGAGGAC	GACGCCGTGG	GCTGCTACAT	GACGGTGACC	120
CTCGTGGACG	ACACCAAGGT	GGAGGGTACC	ATCTTCACCT	ACAATTCCAA	GGAGGGCATC	180
ATAGTACTCC	TGTCCCTCCG	CGACGATCAG	ACGAACATGA	AGCTAATCCG	CACTCCGTAC	240
ATCAAAGACT	TCAGCCTTTC	ACACGCTGAG	GAGGGAGCGC	ACCTGCCCCC	GGCACTGGAC	300
TCCTTCAACG	AGCTTCCGTC	CATGCACGCC	GGCCGCGACA	AGTCCATCTT	CAAGCACGCC	360
AGCACGCAGC	TCAAGAACGC	CGAGGCGAAC	CGCGAAAAGC	ACTTCAACTC	TGTCACGACC	420
GACACACCGA	TTGCCACACT	TGATGCGTAC	CTCAAGCTCC	TGCGGCTATA	CCCCTTAATT	480
GAGTGGAACA	GCGACGAGGG	TGTCATCCAG	GTCTCGGACA	CCGTCATTGT	CGTAGGAGAC	540
CCCGACTGGC	GGACGCCCAA	GGCAATGCTG	GTGGACGCC	CCCCTGAGAA	GGACAGACCG	600
CTTGTAGATC	GCCTGCAGGT	TGCGCTCGGM	AACGGCAAGA	AGTGATTCAG	TGTGTAGCGG	660
ACAGAACATC	GTGTGCTTGT	GTGTCTGTTT	GANGTTTGTT	TGTTTTCTCT	TTGTGGTACT	720
GCGTACGACG	GCGCCTTCTC	CCGGTGGTGG	GTGAGTCCAT	AAGCAGTTGA	GTTCTYGGTT	780
GTAGNAAVGC	CTYACYGÇCG	ACCATATGGG	AGAGGCGAA	CAAATNTTTG	ATAGAAGTTG	840
AAAATCCCAA	AGTYAAAAGA	AAAAAAAA	AAA			873

## (2) INFORMATION FOR SEQ ID NO:75:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1238 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

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# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:75:

TTTCTGTACT	TTATTGAACA	TCAGTAGAAC	ACGTTCTTCC	CGCAAAGATG	GCCAAGAAGC	60
ACCTCAAGCG	CTTĢTATGCG	CCCAAGGACT	GGATGCTGAG	CAAGCTGACC	GGCGTGTTCG	120
CGCCGCGTCC	GCGTCCGGGT	CCGCACAAGC	TGCGCGAGTG	CCTGCCGCTN	CTGGTGATCA	180
ŢCCGCAACCG	GCTGAAGTAC	GCGCTGAACG	CGCGCGAGGG	TGAGATGATC	CTGCGCCAGG	240
GTCTGGTGCA	CGTGGACAAC	CACCCGCGCC	GCGACGGCAA	GTATCCCGCC	GGTTTCATGG	. 300
ACGTGGTCGA	GATCCCGAAG	ACGGGCGACC	GCTTCCGCCT	GATGTACGAC	GTCAAGGGCC	360
GCTTCGCGTT	GGTGAACCTG	TCCGAGGCGG	AGGCGCAGAT	CAAGCTGATG	AAGGTTGTGA	420
ACCTGTACAC	GGCCACCGGC	CGCGTGCCGG	TCGCTGTGAC	GCACGACGGC	CACCGCATCC	480
GCTACCCGGA	CCCGCACACC	TCCATTGGTG	ACACCATCGT	GTACAACGTC	AAGGAGAAGA	540
AGTGCGTGGA	CCTGATCAAG	AACCGCCAGG	GCAAGGCCGT	GATCGTGACC	GGTGGCGCCA	600
ACCGCGGCCG	CATCGGCGAG	ATCGTGAAGG	TGGAGTGCCA	CCCCGGTGCG	TTCAACATTG	660
CGCACCTGAA	GGACGCGTCC	GGCGCCGAGT	TCGCCACCCG	CGCCGCGAAC	ATCTTCGTGA	720
TCGGCAAGGA	CCTGAACAAC	CTGCAGGTAA	CGGTGCCGAA	GCAGCAGGGC	CTGCGCATGA	780
ACGTGATCCA	GGAGCGCGAG	GAGCGCCTGA	TCGCGGCGGA	GGCCCGCAAG	AACGCGCCGG	840
CTCGTGGTGC	CCGCAGGGCC	CGÇAAGTGAG	GAGGCGATTA	CACGCATGCG	TGTTTGTGGC	900
TCTGAAGCGA	CTTGGCGGGT	CGGCTGTGAG	GGTTTGAGAG	GAGGTGTGTG	ATGCGTGTGA	960
AGTCCTTCTC	CGTTCTCAGC	TCTCTCTGTG	CTGTAGCTGT	GCCTTTCCCC	AGATCGCTTT	1020
ACCGCATTTG	CATACATCTG	TGTAGTCGCA	TGTGCGTGTT	TCTGTCTCTC	GGTGGGTCTC	1080
CCTCTCCCTC	CCTTTCTGCC	TCTCTCTTTG	AGTGGGTGTG	CATGCGTCGC	GCGCGACGGG	1140
CTCCGCTTNA	GTGATTCTCT	CGTGTTTTAN	GGCTGTTTTY	TTTCTYAGTT	NAGCGTTTTY	1200
GTTCATGATT	TCCTCAGACC	САААААААА	AAAAAAA			1238

# (2) INFORMATION FOR SEQ ID NO:76:

# (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 712 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single

## (D) TOPOLOGY: linear

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:76:

CTGACGGAGT	TCCAGACGAA	CCTTGTGCCG	TACCCGCGCA	TCCACTTCGT	GCTGACAAGC	60
TACGCTCCGG	TGGTGTCTGC	CGAGAAGGCG	TACCACGAGC	AGCTNTCCGT	CGCGGACATC	120
ACGAACTCGG	TNTTTGAGCC	TGCTGGCATG	CTNACAAAGT	GCGATCCTCG	CCACGGCAAG	180
TACATGTCGT	GCTGCCTCAT	GTACCGCGGT	GATGTCGTGC	CGAAGGATGT	CAACGCCGCG	240
ATTGCGACGA	TCAAGACGAA	GCGCACAATT	CAGTTCGTGG	ACTGGTGCCC	GACCGGCTTC	300
AAGTGCGGCA	TCAACTACCA	GCCGCCGACC	GTTGTGCCCG	GCGGTGACCT	CGCGAAGGTG	360
CAGCGCGCCG	TGTGCATGAT	TGCCAACTCG	ACCGCGATCG	CTGAGGTGTT	TGCCCGCATC	420
GACCACAAGT	TCGACCTGAT	GTACAGCAAG	CGCGCGTTTG	TGCACTGGTA	CGTGGGTGAG	480
GGCATGGAGG	AGGCCGAGTT	CTCCGAGGCG	CGCGAGGATC	TCGCTGCGCT	GGAGAAGGAC	540
TACGAGGAGG	TTGGCGCCGA	GTCCGCCGAC	GACATGGGCG	AGGAGGACGT	CGAGGAGTAC	600
TAAGGTAGAC	TCGTGCCGCG	CGCTGATGAT	GTAGGTGCAC	GCGTGCGTGT	GCTGCAGCGG	660
AGCCGCCGCC	ACCGCGACTG	TGTGTGTGTG	CGCGCGTGAC	GACCGGCTCG	AG	712

## (2) INFORMATION FOR SEQ ID NO:77:

## (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1086 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:77:

CAAGAAGTGG	ATCAAGCAGG	AGACGAACGC	CGATGGCGAG	CGCGTGCGCC	GCGCGTTCTG	60
CCAGTTCTGC	CTAGACCCCA	TCTACCAGAT	CTTCGACGCT	GTGATGAACG	AGAAGAAGGA	120
CAAGGTGGAC	AAGATGCTCA	AGTCGCTGCA	CGTGACGCTN	ACGGCTGAGG	AGCGCGAGCA	180
GGTGCCGAAN	AAGCTTCTGA	AGACGGTGAT	GATGAANTTC	CTGCCGGCTG	CTGAGACGCT	240
GCTACAGATG	ATCGTGGCGC	ACCTGCCGTC	GCCCAAGAAG	GCGCAGGCGT	ACCGTGCGGA	300

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GATGCTGTAC	TCTGGCGAGG	CGTCGCCGGA	GGACAAGTAC	TTCATGGGTA	TCAAGAACTG	360
CGACCCCGCT	GCGCCGCTCA	TGCTGTACAT	CAGCAAGATG	GTGCCGACGG	CCGACCGCGG -	420
CCGCTTCTTC	GCCTTTGGCC	GCATCTTCTC	CGGTAAGGTG	CGCAGCGGCC	AGAAGGTGCG	480
CATCATGGGT	AACAACTACG	TCTACGGCAA	GAAGCAGGAC	CTGTACGAGG	ACAAGCCTGT	540
GCAGCGCTCC	GTGCŢGATGA	TGGGCCGCTA	CCAGGAGGCC	GTGGAGGACA	TGCCGTGCGG	600
TAACGTGGTG	GGCCTTGTGG	GCGTGGACAA	GTACATCGTG	AAGTCCGCGA	CGATCACGGA	660
CGATGGCGAG	AGCCCGCACC	CGCTGCGCGA	CATGAAGTAC	TCTGTGTCGC	CCGTCGTGCG	720
TGTGGCCGTG	GAGGCGAAGA	ACCCGTCCGA	CCTGCCGAAG	CTTGTGGAGG	GCCTGAAGCG	780
CCTTGCCAAG	TCCGACCCGC	TGGTGGTGTG	CAGCATTGAG	GAGTCTGGCG	AGCACATTGT	840
TGCCGGCGCT	GGCGAGCTTC	ACCTTGAGAT	TTGCCTGAAG	GATCTCCAGG	AGGACTTCAT	900
GAACGGCGCG	CCGCTNAAGA	TCTCCGAGCC	GGTGGTGTCG	TTCCGCGAGA	CGGTGACGGA	960
TGTGTCGTCG	CAGCAGTGCC	TGTCGAAGTC	TGCGAACAAG	CACAACCGTC	TCTTCTGCCG	1020
CGGTGCGCCG	CTNACAGAGG	ANCTGGCGCT	GGCGATNGAN	GAAGGCACCG	CTGGTCCCGA	1080
NGCGGA						1086

## (2) INFORMATION FOR SEQ ID NO:78:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 447 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:78:

CGCATCAACG	TCTACTTCGA	TNAGTCGACG	GGAGGCCGCT	ACGTGCCGCG	CGCCGTGCTG	60
ATGGACCTCG	AGCCCGGCAC	TATGGACTCC	GTTCGCGCCG	GCCCGTACGG	CCAGCTGTTC	120
CGCCCGGACA	ACTTCATCTT	TGGTCAGTCC	GGCGCTGGCA	ACAACTGGGC	CAAGGGCCAC	180
TACACTGAGG	GCGCGGAGCT	GATCGACTCC	GTGCTTGATG	TGTGCCGCAA	GGAGGCGGAG	240
AGCTGCGACT	GCCTGCAGGG	CTTCCAGCTG	TCTCACTCCC	TCGGCGGCGG	CACGGGCTCC	300
GGCATGGGCA	CGCTGCTCAT	TTCCAANCTG	CGCGANGAGT	ACCCGGACCG	GATCATGATG	360

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ACCTTCTCCG TCATCCCGTC CCCCCGCGTG TCGGATACCG TTGTGGANCC GTACAACACG	420
ACCCTCTCTG TGCACCAGCT CGTGGAA	447
(2) INFORMATION FOR SEQ ID NO:79:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 375 base pairs</li> <li>(B) TYPE: nucleic acid</li> <li>(C) STRANDEDNESS: single</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:79:	
GTAACCCGCT GGTGTACGCA TATGTAGACA CAGACGGGCA GCACGAGACG ACGTTCCTCG	60 .
CGATCCCTGT GGTGCTTGGC ATGAATGGAA TCGAGAAGCG CCTGCCGATT GGTCCGCTGC	120
ACTCGACGGA GGAAACGCTG CTGAAGGCGG CACTGCCGGT GATCAAGAAG AATATCGTGA	180
AGGGCAGCGA GTTCGCGCGC TCACACCTGT AGCACCTCAG CTTTTTTTT TTGCGTTAAA	240
CGGGCGTGGG AAGCACCTCG ATACTTCGCT TCGCGCTGAC GGACCCGCAC GACATCGTTC	300
GTCATCCCCC TCCCCCTCTT CGGCCCTATA CGCATGAAGG AGTGGAATTA TGCAACAGCA	360
TGTTNATATC AAGTG	375
(2) INFORMATION FOR SEQ ID NO:80:	
<ul> <li>(i) SEQUENCE CHARACTERISTICS:</li> <li>(A) LENGTH: 107 amino acids</li> <li>(B) TYPE: amino acid</li> <li>(C) STRANDEDNESS:</li> <li>(D) TOPOLOGY: linear</li> </ul>	
(D) TOPOLOGI: Tillear	,
	•
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:80:	
Met Ala Ser Ser Arg Lys Ala Ser Asn Pro His Lys Ser His Arg Lys	
1 5 10 15	
Pro Lys Arg Ser Trp Asn Val Tyr Val Gly Arg Ser Leu Lys Ala Ile 20 25 30	
Asn Ala Gln Met Ser Met Ser His Arg Thr Met Lys Ile Val Asn Ser	

35

45

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Tyr Val Asn Asp Val Met Glu Arg Ile Cys Thr Glu Ala Ala Ser Ile 50 55 60

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Val Arg Ala Asn Lys Lys Arg Thr Leu Gly Ala Arg Glu Val Gln Thr 65 70 75 80

Ala Val Arg Ile Val Leu Pro Ala Glu Leu Ala Lys His Ala Met Ala 85 90 95

Glu Gly Thr Lys Ala Val Ser Ser Ala Ser Arg

### (2) INFORMATION FOR SEQ ID NO:81:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 381 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:81:

Asp Glu Glu Glu Glu Asp Thr Thr Ile Asn Asn Ser Asp Val Val 1 10 15

Arg Tyr Lys Lys Ala Ala Thr Trp Cys Asn Glu Thr Leu Arg Val Leu
20 25 30

Ile Asp Ala Thr Lys Pro Gly Ala Lys Val Cys Asp Leu Cys Arg Leu
35 40 45

Gly Asp Asp Thr Ile Thr Ala Xaa Val Lys Thr Met Phe Lys Gly Thr 50 55 60

Glu Lys Gly Ile Ala Phe Pro Thr Cys Ile Ser Val Asn Asn Cys Val 65 70 75 80

Cys His Asn Ser Pro Gly Val Ser Asp Glu Thr Thr Gln Gln Glu Ile 85 90 95

Ala Met Gly Asp Val Val His Tyr Asp Leu Gly Ile His Val Asp Gly
100 105 110

Tyr Cys Ala Val Val Ala His Thr Ile Gln Val Thr Glu Asp Asn Glu 115 120 125

Leu Gly Lys Asp Glu Lys Ala Ala Arg Val Ile Thr Ala Ala Tyr Asn 130 135 140

Ile Leu Asn Thr Ala Leu Arg Gln Met Arg Pro Gly Thr Thr Ile Tyr
145 150 155 160

Gln	Val	Thr	Asp	Val 165	Val	Glu	Lys	Ala	Ala 170	Glu	His	Tyr	Lys	Val 175	Thr
Pro	Val	Asp	Gly 180	Val	Leu	Ser	His	Met 185	Met	Lys	Arg	Tyr	Ile 190	Ile	Asp
Xaa	Tyr	Arg 195	Cys	Ile	Pro	Gln	Arg 200	Arg	Val	Ala	Glu	His 205	Met	Val	His
Asp	Tyr 210	Asp	Leu.	Glu	Lys	Ala 215	Gln	Val	Trp	Thr	Leu 220	Asp	Ile	Val	Met
Thr 225	Ser	Gly	Lys	Gly	Lys 230	Leu	Lys	Glu	Arg	Asp 235	Ala	Arg	Pro	Cys	Val 240
Phe	Lys	Val	Ala	Leu 245	Asp	Ser	Àsn	Tyr	Ser 250	Val	Lys	Met	Glu	Ser 255	Ala
Lys	Glu	Val	Gln 260	Lys	Glu	Ile	Asp	Ser 265	Xaa	Tyr	Ala	Thr	Phe 270	Pro	Phe
Ala	Ile	Arg 275	Asn	Leu	Glu	Ala	Lys 280	Lys	Ala	Arg	Lèu	Gly 285	Leu	Asn	Glu
Met	Ala 290	Lys	His	Gly	Ala	Val 295	Ile	Pro	Tyr	Pro	Ile 300	Leu	Phe	Glu	Lys
Glu 305	-	Glu	Val	Val	Ala 310	His	Phe	Lys	Ile	Thr 315	Val	Leu	Ile	Ser	Asn 320
Lys	Lys	Ile	Glu	Pro 325	Ile	Thr	Gly	Leu	Lys 330		Gln	Lys	Ala	Pro 335	Ala
Leu	Glu	Pro	Tyr 340		Asp	Glu	Met	Leu 345		Ala	Thr	Asn	Lys 350	Leu	Phe
Ala	Val	Ala 355		Glu	Glu	Gly	Gly 360		Val	Asp		Arg 365	Gly	Ile	Arg
Asp	Ala 370		Leu	Arg	Ala	Phe 375	Val	Gly	Val	Arg	Leu 380				

# (2) INFORMATION FOR SEQ ID NO:82:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 191 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

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(xi)	SEQUENCE	DESCRIPTION:	SEO	TD	NO - 82 -

Met Ser Ile Ile Lys Glu Asp Asp Ala Val Gly Cys Tyr Met Thr Val 1 5 10 15

Thr Leu Val Asp Asp Thr Lys Val Glu Gly Thr Ile Phe Thr Tyr Asn 20 25 30

Ser Lys Glu Gly Ile Ile Val Leu Leu Ser Leu Arg Asp Asp Gln Thr 35 40 45

Asn Met Lys Leu Ile Arg Thr Pro Tyr Ile Lys Asp Phe Ser Leu Ser 50 55 60

His Ala Glu Glu Gly Ala His Leu Pro Pro Ala Leu Asp Ser Phe Asn 65 70 75 80

Glu Leu Pro Ser Met His Ala Gly Arg Asp Lys Ser Ile Phe Lys His
85 90 95

Ala Ser Thr Gln Leu Lys Asn Ala Glu Ala Asn Arg Glu Lys His Phe
100 105 110

Asn Ser Val Thr Thr Asp Thr Pro Ile Ala Thr Leu Asp Ala Tyr Leu 115 120 . 125

Lys Leu Leu Arg Leu Tyr Pro Leu Ile Glu Trp Asn Ser Asp Glu Gly
130 135 140

Val Ile Gln Val Ser Asp Thr Val Ile Val Val Gly Asp Pro Asp Trp
145 150 155 160

Arg Thr Pro Lys Ala Met Leu Val Asp Gly Ala Pro Glu Lys Asp Arg 165 170 175

Pro Leu Val Asp Arg Leu Gln Val Ala Leu Gly Asn Gly Lys Lys 180 185 190

## (2) INFORMATION FOR SEQ ID NO:83:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 273 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

## (xi) SEQUENCE DESCRIPTION: SEQ ID NO:83:

Met Ala Lys Lys His Leu Lys Arg Leu Tyr Ala Pro Lys Asp Trp Met

1 5 10 15

Leu Ser Lys Leu Thr Gly Val Phe Ala Pro Arg Pro Arg Pro Gly Pro
20 25 30

His Lys Leu Arg Glu Cys Leu Pro Leu Leu Val Ile Ile Arg Asn Arg
35 40 45

Leu Lys Tyr Ala Leu Asn Ala Arg Glu Gly Glu Met Ile Leu Arg Gln 50 55 60

Gly Leu Val His Val Asp Asn His Pro Arg Arg Asp Gly Lys Tyr Pro 65 70 75 80

Ala Gly Phe Met Asp Val Val Glu Ile Pro Lys Thr Gly Asp Arg Phe 85 90 95

Arg Leu Met Tyr Asp Val Lys Gly Arg Phe Ala Leu Val Asn Leu Ser 100 105 110

Glu Ala Glu Ala Gln Ile Lys Leu Met Lys Val Val Asn Leu Tyr Thr 115 120 125

Ala Thr Gly Arg Val Pro Val Ala Val Thr His Asp Gly His Arg Ile 130 135 140

Arg Tyr Pro Asp Pro His Thr Ser Ile Gly Asp Thr Ile Val Tyr Asn 145 150 155 160

Val Lys Glu Lys Lys Cys Val Asp Leu Ile Lys Asn Arg Gln Gly Lys 165 170 175

Ala Val Ile Val Thr Gly Gly Ala Asn Arg Gly Arg Ile Gly Glu Ile 180 185 190

Val Lys Val Glu Cys His Pro Gly Ala Phe Asn Ile Ala His Leu Lys 195 200 205

Asp Ala Ser Gly Ala Glu Phe Ala Thr Arg Ala Ala Asn Ile Phe Val 210 215 220

Ile Gly Lys Asp Leu Asn Asn Leu Gln Val Thr Val Pro Lys Gln Gln 225 230 235 240

Gly Leu Arg Met Asn Val Ile Gln Glu Arg Glu Glu Arg Leu Ile Ala 245 250 255

Ala Glu Ala Arg Lys Asn Ala Pro Ala Arg Gly Ala Arg Arg Ala Arg 260 265 270

Lys

#### (2) INFORMATION FOR SEQ ID NO:84:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 200 amino acids

- (B) TYPE: amino acid
- (C) STRANDEDNESS:
- (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:84:

Leu Thr Glu Phe Gln Thr Asn Leu Val Pro Tyr Pro Arg Ile His Phe 1 5 10 15

Val Leu Thr Ser Tyr Ala Pro Val Val Ser Ala Glu Lys Ala Tyr His
20 25 30

Glu Gln Leu Ser Val Ala Asp Ile Thr Asn Ser Val Phe Glu Pro Ala 35 40 45

Gly Met Leu Thr Lys Cys Asp Pro Arg His Gly Lys Tyr Met Ser Cys 50 55 60

Cys Leu Met Tyr Arg Gly Asp Val Val Pro Lys Asp Val Asn Ala Ala 65 70 75 80

Ile Ala Thr Ile Lys Thr Lys Arg Thr Ile Gln Phe Val Asp Trp Cys 85 90 95

Pro Thr Gly Phe Lys Cys Gly Ile Asn Tyr Gln Pro Pro Thr Val Val
100 105 110

Pro Gly Gly Asp Leu Ala Lys Val Gln Arg Ala Val Cys Met Ile Ala 115 120 125

Asn Ser Thr Ala Ile Ala Glu Val Phe Ala Arg Ile Asp His Lys Phe 130 135 140

Asp Leu Met Tyr Ser Lys Arg Ala Phe Val His Trp Tyr Val Gly Glu 145 150 155 160

Gly Met Glu Glu Gly Glu Phe Ser Glu Ala Arg Glu Asp Leu Ala Ala 165 170 175

Leu Glu Lys Asp Tyr Glu Glu Val Gly Ala Glu Ser Ala Asp Asp Met 180 185 190

Gly Glu Glu Asp Val Glu Glu Tyr

### (2) INFORMATION FOR SEQ ID NO:85:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 361 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:

# (D) TOPOLOGY: linear

(xi)	SEQU	JENCE	E DES	CRIE	MOIT	: SE	Q II	NO:	85:						
Lys 1	Lys	Trp	Ile	Lys 5	Gln	Glu	Thr	Asn	Ala 10	Asp	Gly	Glu	Arg	Val 15	Arg
.Arg	Ala	Phe	Cys 20	Gln	Phe	Cys	Leu	Asp 25	Pro	Ile	Tyr	Gln	Ile 30	Phe	Asp
Ala	Val	Met <b>3</b> 5	Asn	Glu	Lys	Lys	Asp 40	Lys	Val	Asp	Lys	Met 45	Leu	Lys	Ser
Leu	His 50	Val	Thr	Leu	Thr	Ala 55	Glu	Glu	Arg	Glu	Gln 60	Val	Pro	Xaa	Lys
Leu 65	Leu	Lys	Thr	Val	Met 70	Met	Xaa	Phe	Leu	Pro 75	Ala	Ala	Glu	Thr	Leu 80
Leu	Gln	Met	Ile	Val 85	Ala	His	Leu	Pro	Ser 90	Pro	Lys	Lys	Ala	Gln 95	Ala
Tyr	Arg	Ala	Glu 100	Met	Leu	Tyr	Ser	Gly 105	Glu	Ala	Ser	Pro	Glu 110	Asp	Lys
Tyr	Phe	Меţ 115	Gly	Ile	Lys	Asn	Cys 120	Asp	Pro	Ala	Ala	Pro 125	Leu	Met	Leu
Tyr	Ile 130	Ser	Lys	Met	Val	Pro 135	Thr	Ala	Asp	Arg	Gly 140	Arg	Phe	Phe	Ala
Phe 145	_	Arg	Ile	Phe	Ser 150	Gly	Lys	Val	Arg	Ser 155	Gly	Gln	Lys	Val	Arg 160
Ile	Met	Gly	Asn	Asn 165		Val	Tyr	Gly	Lys 170	Lys	Gln	Asp	Leu	Tyr 175	Glu
Asp	Lys	Pro	Val 180		Arg	Ser	Val	Leu 185		Met	Gly	Arg	Tyr 190	Gln	Glu
Ala	Val	Glu 195		Met	Pro	Cys	Gly 200		Val	Val	Gly	Leu 205		Gly	Val
Asp	Lys 210		lle	· Val	Lys	Ser 215		Thr	Ile	Thr	Asp 220		Gly	Glu	Ser
Pro 225		Pro	Lev	Arg	230		Lys	Tyr	Ser	Val 235		Pro	Val	Val	Arg 240

Val Ala Val Glu Ala Lys Asn Pro Ser Asp Leu Pro Lys Leu Val Glu

250

255

Gly Leu Lys Arg Leu Ala Lys Ser Asp Pro Leu Val Val Cys Ser Ile 260 265 270

Glu Glu Ser Gly Glu His Ile Val Ala Gly Ala Gly Glu Leu His Leu 275 280 285

Glu Ile Cys Leu Lys Asp Leu Gln Glu Asp Phe Met Asn Gly Ala Pro 290 295 300

Leu Lys Ile Ser Glu Pro Val Val Ser Phe Arg Glu Thr Val Thr Asp 305 310 315 320

Val Ser Ser Gln Gln Cys Leu Ser Lys Ser Ala Asn Lys His Asn Arg 325 330 335

Leu Phe Cys Arg Gly Ala Pro Leu Thr Glu Xaa Leu Ala Leu Ala Xaa 340 345 350

Xaa Glu Gly Thr Ala Gly Pro Xaa Ala 355 360

#### (2) INFORMATION FOR SEQ ID NO:86:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 149 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS:
  - (D) TOPOLOGY: linear

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:86:

Arg Ile Asn Val Tyr Phe Asp Xaa Ser Thr Gly Gly Arg Tyr Val Pro 1 5 10 15

Arg Ala Val Leu Met Asp Leu Glu Pro Gly Thr Met Asp Ser Val Arg
20 25 30

Ala Gly Pro Tyr Gly Gln Leu Phe Arg Pro Asp Asn Phe Ile Phe Gly 35 40 45

Gln Ser Gly Ala Gly Asn Asn Trp Ala Lys Gly His Tyr Thr Glu Gly
50 55 60

Ala Glu Leu Ile Asp Ser Val Leu Asp Val Cys Arg Lys Glu Ala Glu 65 70 75 80

Ser Cys Asp Cys Leu Gln Gly Phe Gln Leu Ser His Ser Leu Gly Gly 85 90 95

Gly Thr Gly Ser Gly Met Gly Thr Leu Leu Ile Ser Xaa Leu Arg Xaa

154

100

105.

110

Glu Tyr Pro Asp Arg Ile Met Met Thr Phe Ser Val Ile Pro Ser Pro 115 120 125

Arg Val Ser Asp Thr Val Val Xaa Pro Tyr Asn Thr Thr Leu Ser Val
130 135 140

His Gln Leu Val Glu 145

- (2) INFORMATION FOR SEQ ID NO:87:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 69 amino acids
    - (B) TYPE: amino acid
    - (C) STRANDEDNESS:
    - (D) TOPOLOGY: linear
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:87:

Asn Pro Leu Val Tyr Ala Tyr Val Asp Thr Asp Gly Gln His Glu Thr 1 5 10 15

Thr Phe Leu Ala Ile Pro Val Val Leu Gly Met Asn Gly Ile Glu Lys
20 25 30

Arg Leu Pro Ile Gly Pro Leu His Ser Thr Glu Glu Thr Leu Leu Lys
35 40 45

Ala Ala Leu Pro Val Ile Lys Lys Asn Ile Val Lys Gly Ser Glu Phe 50 55 60

Ala Arg Ser His Leu 65

#### 155

#### **CLAIMS**

- 1. A polypeptide comprising an immunogenic portion of a *Leishmania* antigen having the amino acid sequence recited in SEQ ID NO:2, or a variant of said antigen that differs only in conservative substitutions and/or modifications.
- 2. The polypeptide of claim 1, comprising amino acids 1 564 of SEQ ID NO:2.
- 3. An isolated DNA molecule comprising a nucleotide sequence encoding a polypeptide according to claim 1.
- 4. The DNA molecule of claim 3 wherein the nucleotide sequence is selected from the group consisting of:
  - (a) nucleotides 421 through 2058 of SEQ ID NO:1; and
- (b) DNA sequences that hybridize to a nucleotide sequence complementary to nucleotides 421 through 2058 of SEQ ID NO:1 under moderately stringent conditions.
- 5. A recombinant expression vector comprising the DNA molecule of claim 3.
- 6. A host cell transformed or transfected with the expression vector of claim 5.
- 7. A polypeptide comprising an immunogenic portion of a *Leishmania* antigen having the amino acid sequence recited in SEQ ID NO:4, or a variant of said antigen that differs only in conservative substitutions and/or modifications.

- 8. The polypeptide of claim 7, comprising amino acids 1-175 of SEQ ID NO:4.
- 9. An isolated DNA molecule comprising a nucleotide sequence encoding a polypeptide according to claim 7.
- 10. The DNA molecule of claim 9 wherein the nucleotide sequence is selected from the group consisting of:
  - (a) nucleotides 25 through 549 of SEQ ID NO:3; and
- (b) DNA sequences that hybridize to a nucleotide sequence complementary to nucleotides 25 through 549 of SEQ ID NO:3 under moderately stringent conditions.
- 11. A recombinant expression vector comprising the DNA molecule of claim 9.
- 12. A host cell transformed or transfected with the expression vector of claim 11.
- 13. A polypeptide comprising an immunogenic portion of a *Leishmania* antigen or a variant of said antigen that differs only in conservative substitutions and/or modifications, wherein said antigen comprises an amino acid sequence selected from the group consisting of sequences recited in SEQ ID NO: 22, 24, 26, 36-38, 41, 49-53 and 82.
- 14. An antigenic epitope of a *Leishmania* antigen comprising an amino acid sequence recited in SEQ ID NO:43.
- 15. A polypeptide comprising at least two contiguous antigenic epitopes according to claim 14.

- 16. An isolated DNA molecule comprising a nucleotide sequence encoding a polypeptide according to claims 13 or 15.
- 17. The DNA molecule of claim 16 wherein the nucleotide sequence is selected from the group consisting of:
- (a) sequences recited in SEQ ID NO: 21, 23, 25, 29-31, 34, 45-48 and 74; and
- (b) DNA sequences that hybridize to a nucleotide sequence complementary to a sequence recited in SEQ ID NO: 21, 23, 25, 29-31, 34, 45-48 and 74 under moderately stringent conditions.
  - 18. An expression vector comprising the DNA molecule of claim 16.
- 19. A host cell transformed or transfected with the expression vector of claim 18.
- 20. A polypeptide comprising an immunogenic portion of a *Leishmania* antigen or a variant of said antigen that differs only in conservative substitutions and/or modifications, wherein said antigen comprises an amino acid sequence provided in SEQ ID NO: 20.
- 21. An isolated DNA molecule comprising a nucleotide sequence encoding a polypeptide according to claim 20.
- 22. The DNA molecule of claim 21 wherein the nucleotide sequence is selected from the group consisting of:
  - (a) a sequence recited in SEQ ID NO: 19; and
- (b) DNA sequences that hybridize to a nucleotide sequence complementary to the sequence of SEQ ID NO: 19 under moderately stringent conditions.

- 23. An expression vector comprising the DNA molecule of claim 21.
- 24. A host cell transformed or transfected with the expression vector of claim 23.
- 25. A pharmaceutical composition comprising a polypeptide according to any one of claims 1, 7, 13 and 15, and a physiologically acceptable carrier.
- 26. A pharmaceutical composition comprising a polypeptide according to claim 20 and a physiologically acceptable carrier.
- 27. A pharmaceutical composition comprising at least two different polypeptides selected from the group consisting of:
  - (a) a polypeptide according to claim 1;
  - (b) a polypeptide according to claim 7;
  - (c) a polypeptide according to claim 13
  - (d) a polypeptide according to claim 15;
  - (e) a polypeptide according to claim 20;
- (f) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:6, or a variant of said antigen that differs only in conservative substitutions and/or modifications;
- (g) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:8, or a variant of said antigen that differs only in conservative substitutions and/or modifications; and
- (h) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:10, or a variant of said antigen that differs only in conservative substitutions and/or modifications;

and a physiologically acceptable carrier.

- 28. A pharmaceutical composition according to any one of claims 25, 26 and 27, further comprising soluble *Leishmania* antigens.
- 29. A pharmaceutical composition according to any one of claims 25, 26 and 27, further comprising a K39 Leishmania antigen.
- 30. A pharmaceutical composition comprising soluble *Leishmania* antigens and a physiologically acceptable carrier.
- 31. A vaccine comprising a polypeptide according to any one of claims 1,7. 13 and 15, and a non-specific immune response enhancer.
- 32. A vaccine comprising a polypeptide according to claim 20 and a non-specific immune response enhancer.
- 33. A vaccine comprising at least two different polypeptides selected from the group consisting of:
  - (a) a polypeptide according to claim 1;
  - (b) a polypeptide according to claim 7;
  - (c) a polypeptide according to claim 13;
  - (d) a polypeptide according to claim 15
  - (e) a polypeptide according to claim 21;
- (f) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:6, or a variant of said antigen that differs only in conservative substitutions and/or modifications;
- (g) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:8, or a variant of said antigen that differs only in conservative substitutions and/or modifications; and

(h) a polypeptide comprising an immunogenic portion of a Leishmania antigen having the amino acid sequence recited in SEQ ID NO:10, or a variant of said antigen that differs only in conservative substitutions and/or modifications;

and a non-specific immune response enhancer.

- 34. A vaccine comprising soluble *Leishmania* antigens and a non-specific immune response enhancer.
- 35. A vaccine according to any one of claims 31, 32, 33 and 34 wherein the non-specific immune response enhancer is an immunogenic portion of a *Leishmania* antigen having the amino acid sequence recited in SEQ ID NO:10, or a variant of said antigen that differs only in conservative substitutions and/or modifications.
- 36. A vaccine according to any one of claims 31, 32 and 33, further comprising soluble *Leishmania* antigens.
- 37. A vaccine according to claim 36 wherein the non-specific immune response enhancer is an immunogenic portion of a *Leishmania* antigen having the amino acid sequence recited in SEQ ID NO:10, or a variant of said antigen that differs only in conservative substitutions and/or modifications.
- 38. A vaccine according to any one of claims 31, 32, 33 and 34, further comprising a delivery vehicle.
- 39. The vaccine of claim 38 wherein the delivery vehicle is a biodegradable microsphere.
- 40. A vaccine comprising a DNA molecule according to any one of claims 3, 9 and 16.

- 41. A vaccine comprising a DNA molecule according to claim 21.
- 42. A pharmaceutical composition according to any one of claims 25, 26, 27, 29 and 30, for use in the manufacture of a medicament for inducing protective immunity against leishmaniasis in a patient.
- 43. The composition of claim 42 wherein the leishmaniasis is caused by a Leishmania species selected from the group consisting of L. donovani, L. chagasi, L. infantum, L. major, L. amazonensis, L. braziliensis, L. panamensis, L. tropica and L. guyanensis.
- 44. A vaccine according to any one of claims 31-34, 40 and 41, for use in a method for inducing protective immunity against leishmaniasis in a patient comprising administering.
- 45. The vaccine of claim 44 wherein the leishmaniasis is caused by a Leishmania species selected from the group consisting of L. donovani, L. chagasi, L. infantum, L. major, L. amazonensis, L. braziliensis, L. panamensis, L. tropica and L. guyanensis.
- 46. A pharmaceutical composition according to any one of claims 25, 26, 27, 29 and 30, for use in a method for detecting *Leishmania* infection in a patient.

### 47. A diagnostic kit comprising:

- (a) a pharmaceutical composition according to any one of claims 25, 26, 27, 29 and 30; and
- (b) apparatus sufficient to contact dermal cells of a patient with the pharmaceutical composition.

- 48. A pharmaceutical composition according to any one of claims 25, 26, 27, 29 and 30, for use in the manufacture of a medicament for stimulating a cellular and/or humoral immune response in a patient.
- 49. The composition of claim 48 wherein said response is a Th1' immune response.
- 50. The composition of claim 48 wherein said response is IL-12 production.
- 51. A vaccine according to any of claims 31-34, 40 and 41, for use in the manufacture of a medicament for stimulating a cellular and/or humoral immune response in a patient.
  - 52. The vaccine of claim 51 wherein said response is a Th1 response.
  - 53. The method of claim 51 wherein said response is IL-12 production.
- 54. A pharmaceutical composition according to any one of claims 25, 26, 27, 29 and 30, for use in the manufacture of a medicament for treating a patient afflicted with a disease responsive to IL-12 stimulation.
- 55. A vaccine according to any one of claims 31-34. 40 and 41, for use in the manufacture of a medicament for treating a patient afflicted with a disease responsive to IL-12 stimulation.
- 56. A pharmaceutical composition comprising a polypeptide and a physiologically acceptable carrier, the polypeptide comprising an immunogenic portion of a *Leishmania* antigen or a variant of said antigen that differs only in conservative substitutions

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and/or modifications, wherein said antigen comprises an amino acid sequence selected from the group consisting of SEQ ID NO:39, 42, 55, 61, 62, 80, 81, and 83-87.

- 57. A pharmaceutical composition according to claim 56 further comprising a K39 Leishmania antigen.
- 58. A vaccine comprising a polypeptide and a non-specific immune response enhancer, the polypeptide comprising an immunogenic portion of a *Leishmania* antigen or a variant of said antigen that differs only in conservative substitutions and/or modifications, wherein said antigen comprises an amino acid sequence selected from the group consisting of SEQ ID NO:39, 42, 55, 61, 62, 80, 81 and 83-87.
- 59. A pharmaceutical composition according to any one of claims 56 and 57, for use in the manufacture of a medicament for inducing protective immunity against leishmaniasis in a patient.
- 60. A vaccine according to claim 58, for use in the manufacture of a medicament for inducing protective immunity against leishmaniasis in a patient.
- 61. A pharmaceutical composition according to any one of claims 56 or 57, for use in a method for detecting *Leishmania* infection in a patient.
- 62. A pharmaceutical composition according to claim 56, and a composition comprising a K39 *Leishmania* antigen, for use in a method for detecting *Leishmania* infection in a patient.

- 63. A diagnostic kit comprising:
- (a) a pharmaceutical composition according to any one of claims 56 and 57; and
- (b) apparatus sufficient to contact dermal cells of a patient with the pharmaceutical composition.

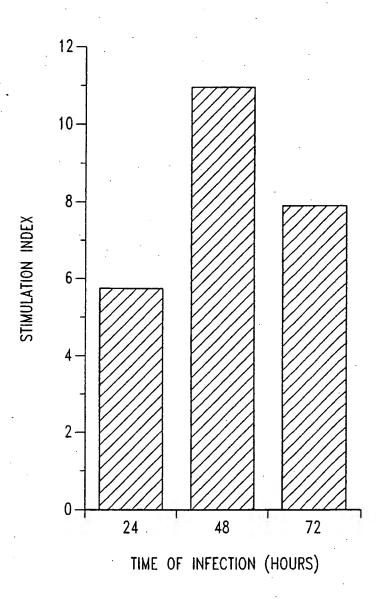
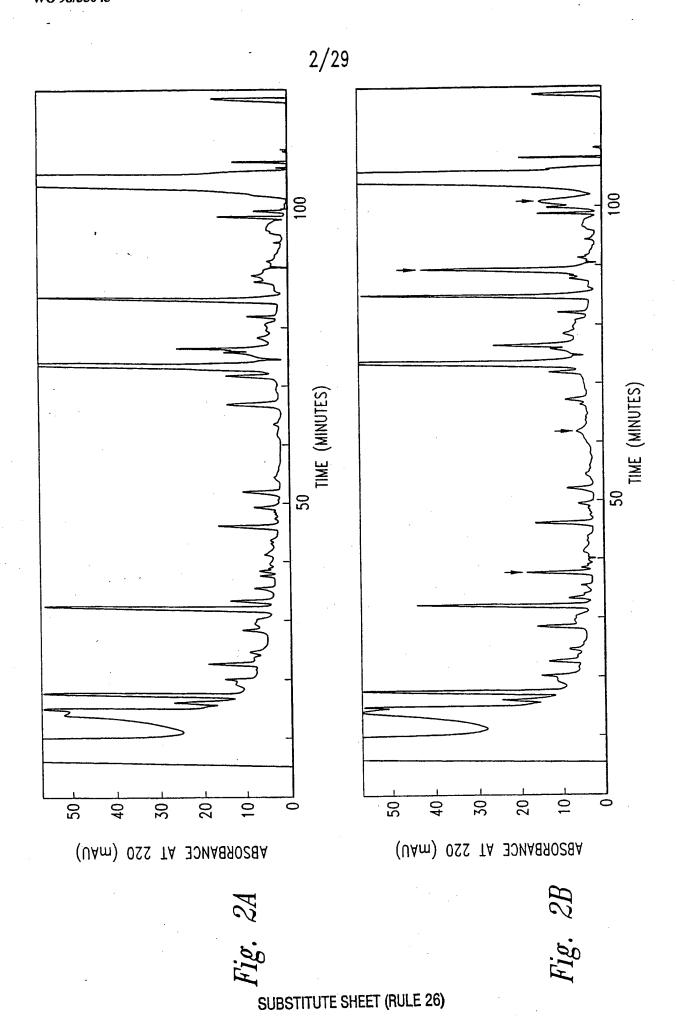
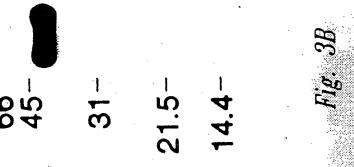
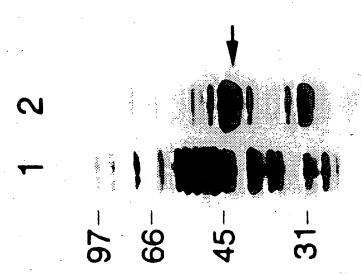


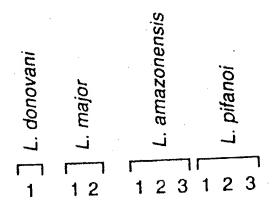
Fig. 1







rig. 34



Wells-

- 9.44 7.46 —
- 4.40 .....
- 2.37
- 1.35



Fig. 4

66 -

45 -

21.5 -14.4 -



Fig. 5

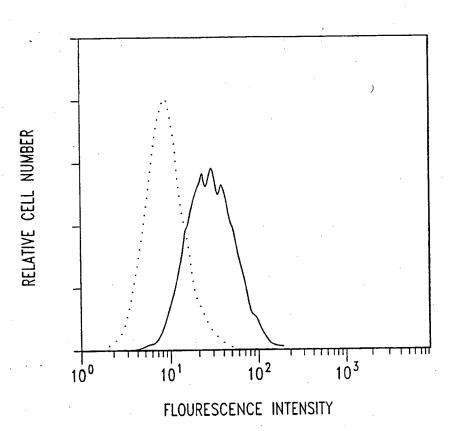


Fig. 6

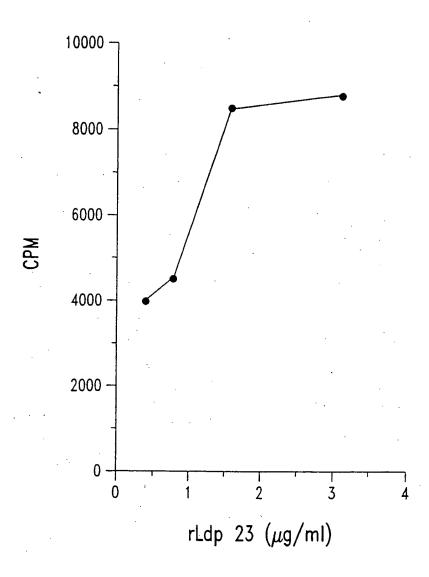
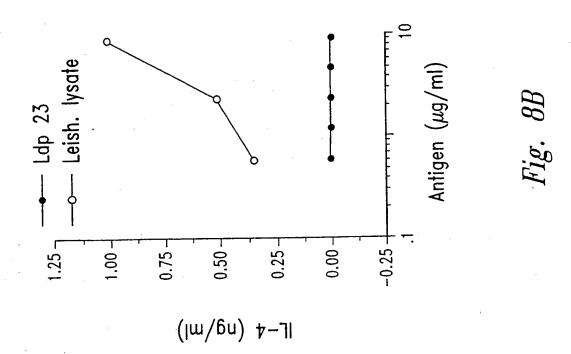
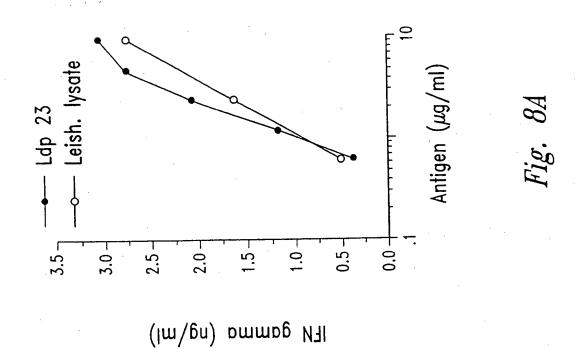
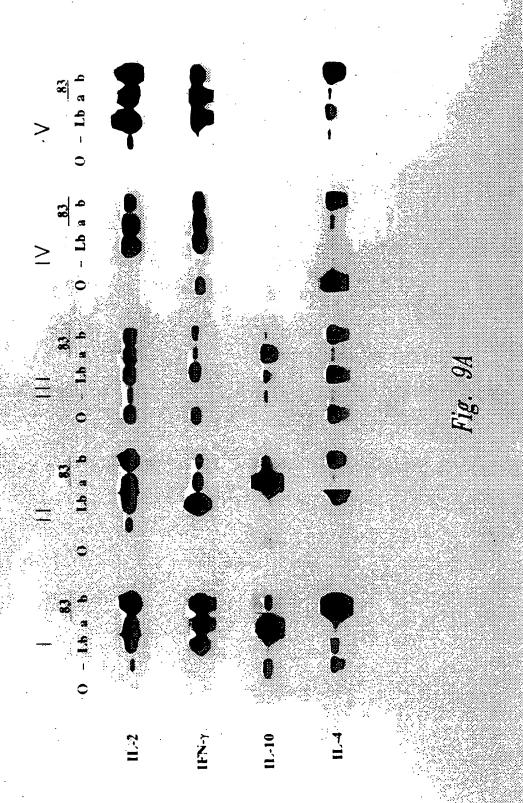
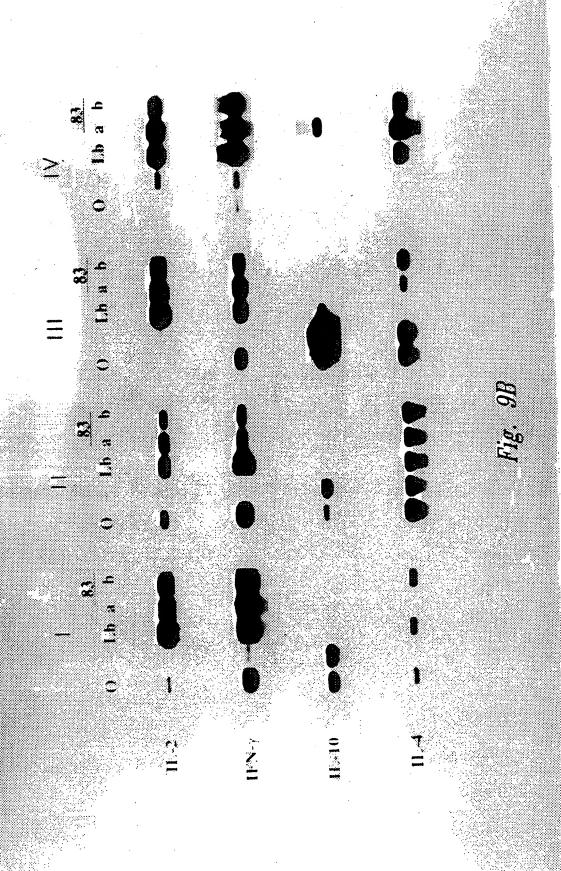


Fig. 7











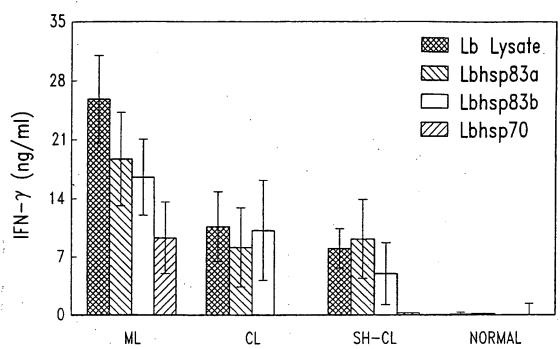


Fig. 10A

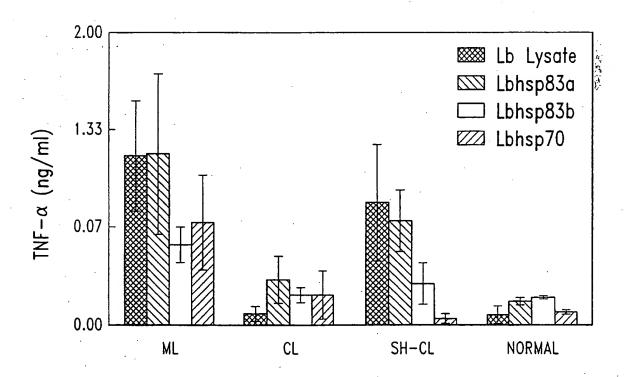


Fig. 10B

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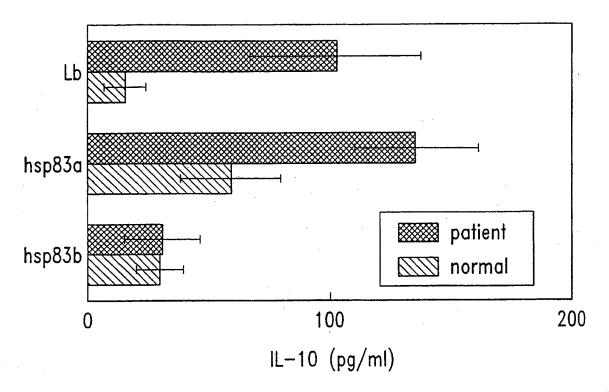


Fig. 11

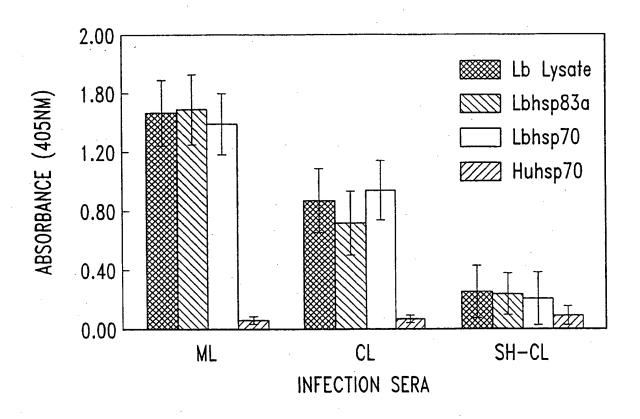
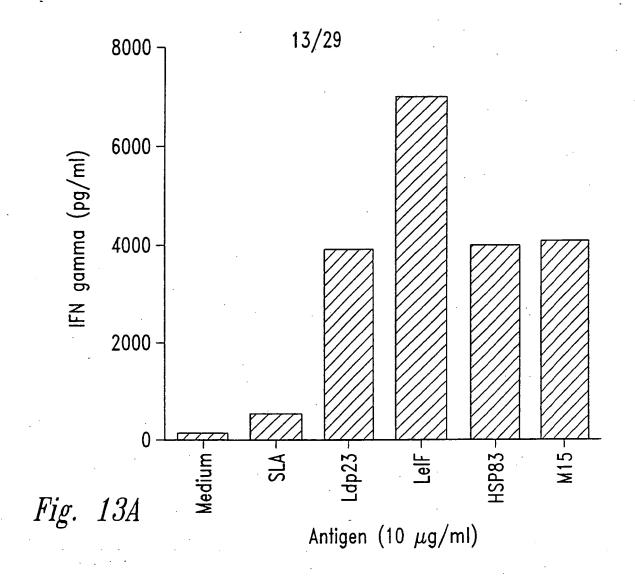


Fig. 12



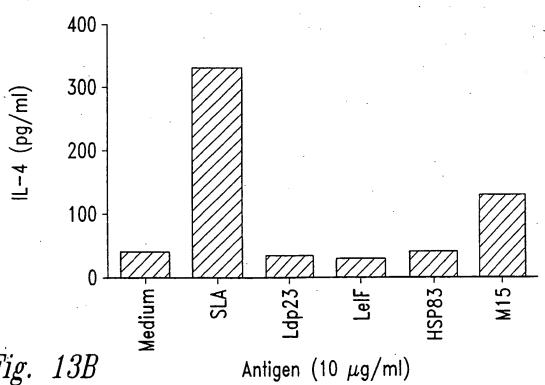


Fig.

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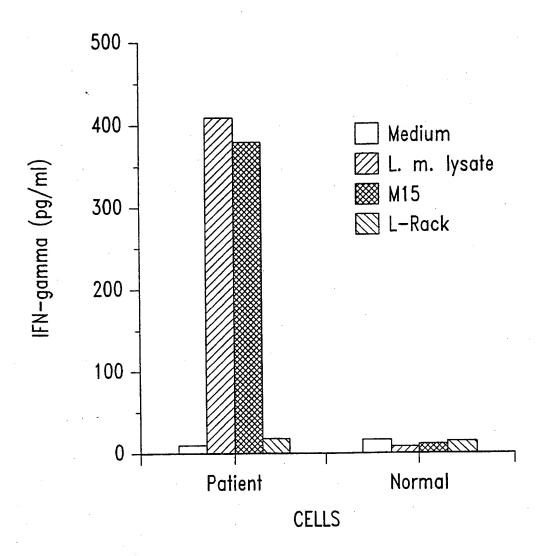


Fig. 14

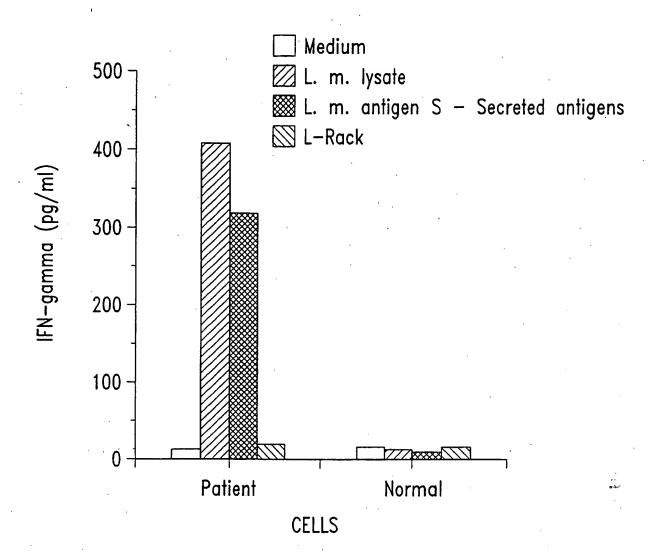


Fig. 15

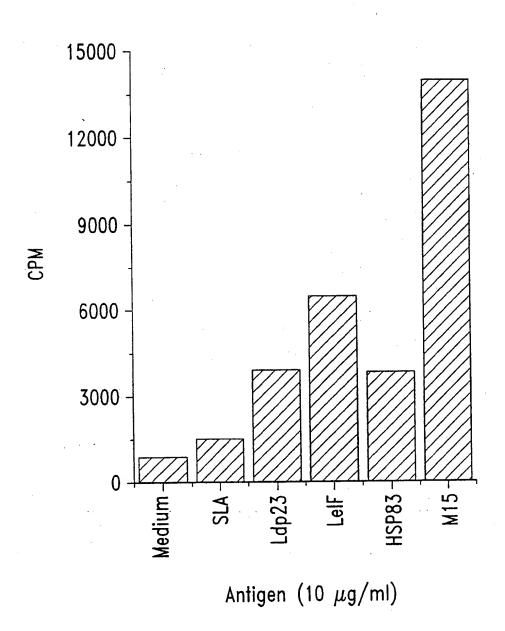


Fig. 16

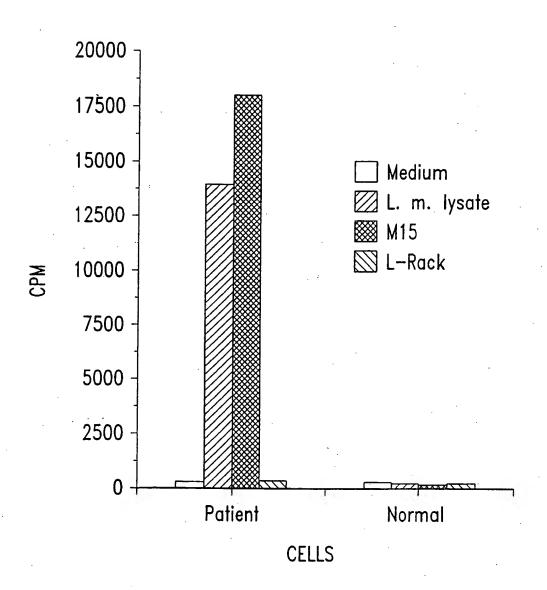


Fig. 17

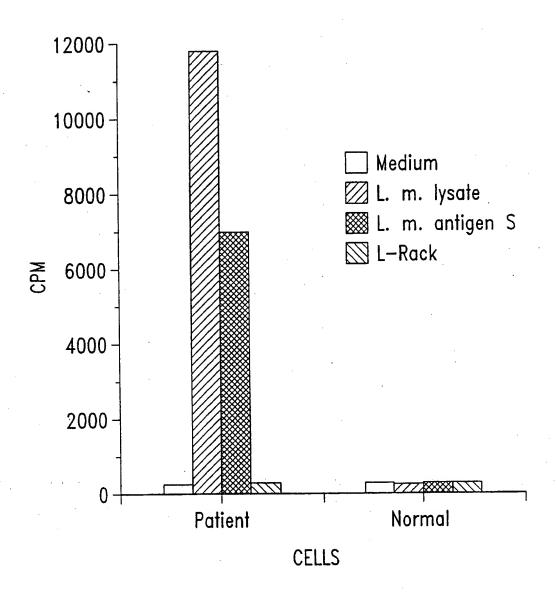


Fig. 18

# 19/29

SLTDPAVLGEETHLRVRVVPDKANKTLTVEDNGIGMTK	85
MTETFAFQAFINQLMSLIINTFYSNKEIFLRDVISNASDACDKIRYQDA.R.CF	85
MPEETQTQDQPMEEEEVAELSLESK. DSGKE. HINLI. N. QDRA IV. T	85
	100
P ADLYNNLGT IARSGTKAFMEALEAGGDMSMIGQFGVGFYSAYLVADRVTVVSKNNSDEAY-VESSAGGTFTITSVQESDMKRGTSTTLHLKEDQQEYLEE	
Δ Τ ΥΥ ΑΡ ΙΡΑΡΙ Ι Α	184 185 185
. E	200
RRVKEL IKKHSEF IGYDIELMVEKTAEKEVTDEDEEEDESKKKSCGDEGEPKVEEVTEGG-ED-KKKKTKKVKEVKKT-YEVKNKHKPLWTRD	274 272
. L	275 300
Lbhsp83b	
TKDVTKFFYAAFYKATSNOVEDTAATKHESVEGOLFERATAEVPKRAPEDMEEPNKKRNNTKLYVRRVETMONCEDLCEDVLGEVKGVVDSEDLPLNTSP	374 372
P	375 400
ENLQQNKILKVIRKNIVKKCLELFEEIAENKEDYKQFYEQFGKNIKLGIHEDTANRKKLMELLRFYSTESGEEMTTLKDYVTRMKPEQKSIYYITGDSKK	474 472
AKVSH.SDEGCV .MSLT.L.D.N.KSSQSY.TSA.D.VSCEN.HET.D	475 500
KLESSPFIEKARRCGLEVLFMTEP JDEYVMQQVKDFEDKKFACLTKEGVHFEESEEEKKQREEKKAACEKLCKTMKEVLGDKVEKVTVSERLLTSPCILV	574 <sup>°</sup>
Q. K. R. FY	572 575 600
Р	
TSEFGWSAHMEQIMRNQALRDSSMAQYMVSKKTMEVNPDHPIIKELRRRVEADENDKAVKDLVFLLFDTSLLTSGFQLDDPTGYAERINRMIKLGLSLDE	674 671
	675 700
EEEEVA-EAPPAEAAPAEVTAGTSSMEQVD 703 Lbhsp83	
EE. V AV TL 701 Lahsp83 .DNGNEEA. VPV 704 Tchsp83	
DDPTADDTSA. VTE. MP. L. GDDD R E 734 Huhsp89	

Fig. 19

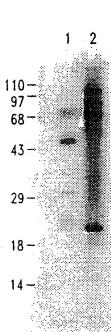


Fig. 20

# 21/29

GAATTCGGCACGAGGTTTCTGTACTTTATTGCTTCCAGCCTTTATTCACTCTTCGATTTCCTCTAACACCATGTCCTCCGAGCGCACCTTTATTGCCGTC
5'-Adaptor Splicad-leader 5'-UT
M S S E R T F I A V  AAGCCGGACGGCGTGCAGGCGCCTCGTTGGCGAGATCATCGCCCCGCTTCGAGGCGCAAGGCTCGTCGCCTTGAAGATACTGCAGCCGACGA 200
K P D G V Q R G L V G E I I A R F E R K G Y K L V A L K I L Q P T  CGCASCAGGGCCAGGGTCACTATAAGGACCTTTGCTCCAAGCCGTTTTTCCCGGCCCTTGTGAAGTACTTCTCCTCTGGCCCGATCGTGTGTATGGTGTG  300
T E O A O G H Y K D L C S K P F F P A L V K Y F S S G P I V C M V W  GCASGGTAAGAACGTGGTGAAGAGCGGCCGCGTGCTGCTGCGCGCGGCGACGACCCGGCCGACTCACAGCCCGGCACGATCCGTGGCGACTTTGCCGTGGAT 400
E G K N V V K S G R V L L G A T N P A D S Q P G T I R G D F A V D  GTGGGCCGCAACGTGTGCCACGGGTCCGACTCTGTGGAGAGCGCGGAGAGCGCCGTTTTGGTTCAAGGCGGATGAGATCGCGAGCTGGACGTCGG-500
V G R N V C H G S D S V E S A E R E I A F W F K A D E I A S W T S  ACTCCGTGTCCCAGATCTATGAGTAACGGTGATTGCGGACACGCTTTGAGGACGTAGCTGTACCCCCAATGAATTCTTCTCTGAAAACCACATCATAAGC  3'-UT
H S V S Q I Y E  CTCTTAAGAGGTTATTTTTCTTGATCGATGCCCGGTGGTGACCAGCACCATTCCTTTATCGGATTCACTCAC
GAGTGGGCTCTGGAGGAGACTGTTGTGTAGCCATGGCTTCAGGAGAGAAAACAAAATACAAGGAAAAGGCAATATGTAACTATGGGGTTCCCTTTTTTACT 3'-UT
ATGCAAAGTTITTATAACTCCTGATCGGCAAAAACAACAACAACCACCACCATACACCAAGAGCAAATGCTTTCTTCTGCGGACTGTGCTTCTTTTTTTT
ATGAAGGAGTGACTCGCGCGATGAAAAGTGTGTGCGTGGGAGATGTATTTCCTTTTTTTGTTCATAGTGGCGACAGCTCACTGTTGACGATGACAAAAAA 1000
AAAAAAAAAAAAACTCGAG 1019
Poly A tail/ Xhoi >

Fig. 21

- L. major — Friedlander

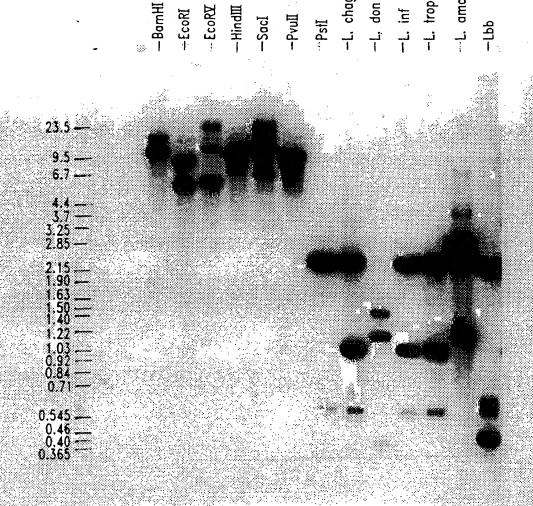


Fig. 22

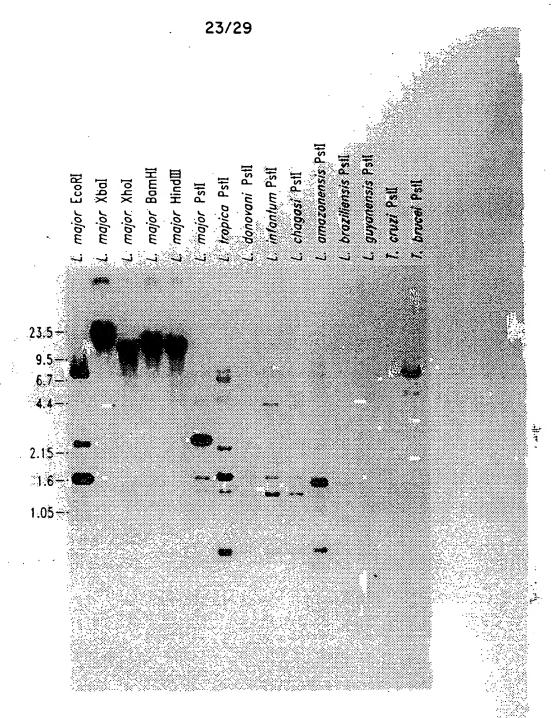


Fig. 23

Proliferative response of human PBMC to MAPS

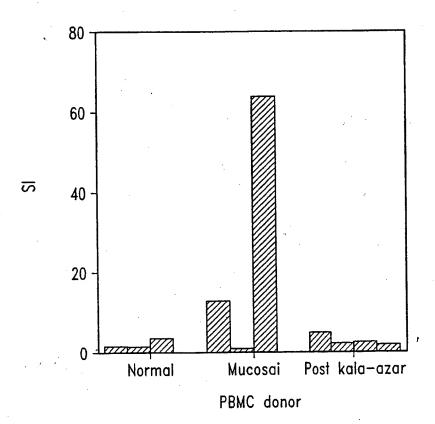


Fig. 24

Proliferative Responses of L. major-infected BALB (20 days post-infection) to recombinant MAPS protein

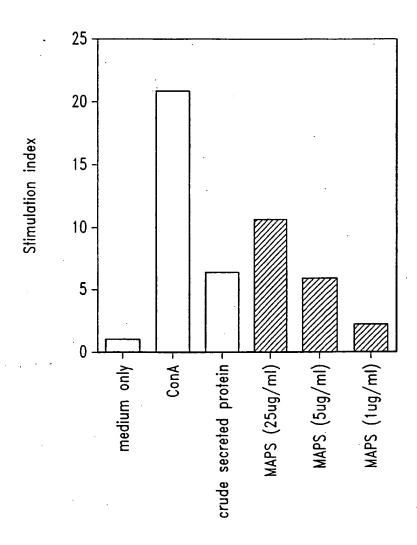


Fig. 25

ELISA analysis of MAPS—specific antibody titre human leishmaniasis patient sera (8/27/96)

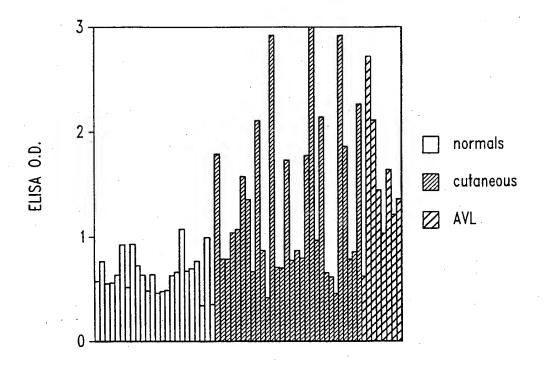


Fig. 26

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# Antibody Response of Immunized/Ir BALB/c Mice to MAPS

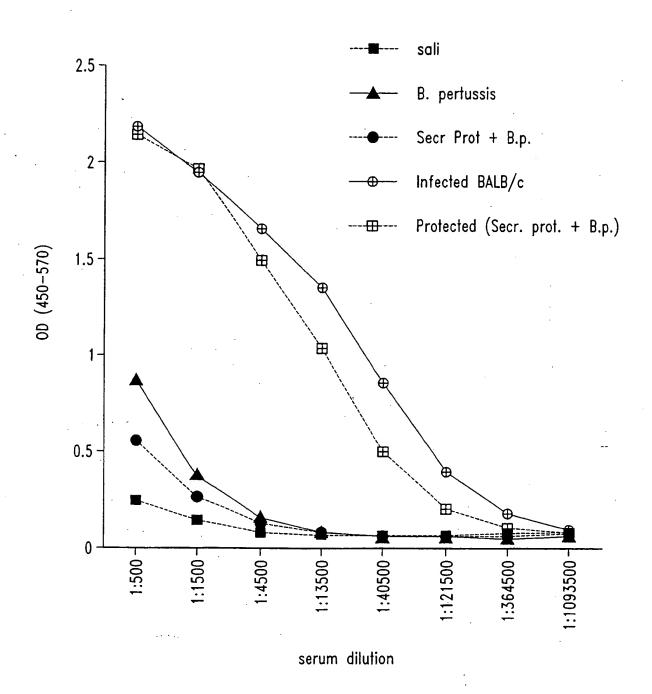
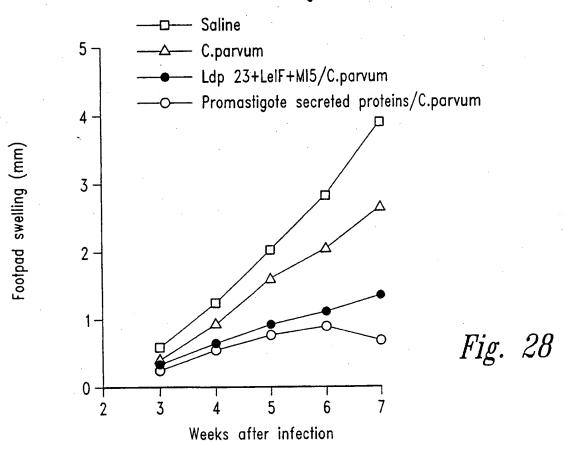


Fig. 27

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Protection against infection with L. major in BALB/c mice immunized leishmanial antigens.



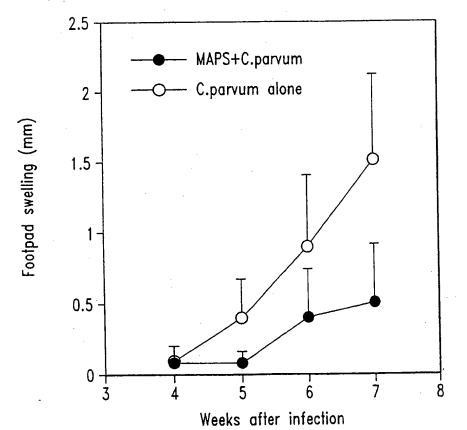
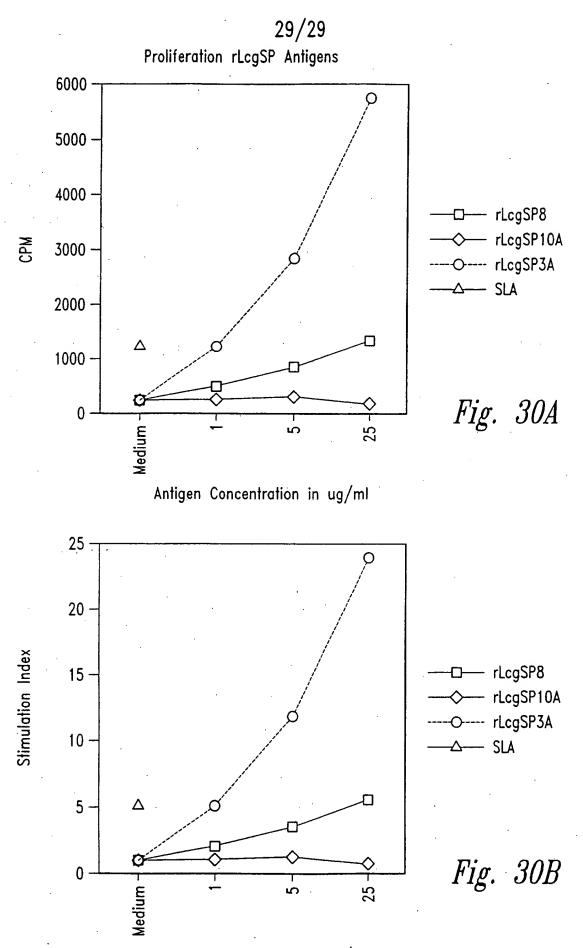


Fig. 29

WO 98/35045



Antigen Concentration in ug/ml
SUBSTITUTE SHEET (RULE 26)

Internati Application No PCT/US 98/03002

A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12N15/30 C07K14/44 C12N15/63 C12N5/10 A61K39/008 A61K48/00

According to International Patent Classification (IPC) or to both national classification and IPC

#### B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N CO7K A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

Category °	Citation of document, with indication, where approp	riate, of the relevant passages	Relevant to claim No.
A	WO 95 29239 A (CORIXA CO November 1995	1-6,25, 27-29, 31, 35-40, 42-55	
	see page 2, line 17 - page see page 5, line 8 - page examples	ge 3, line 37 e 23, line 9; -/	

X Further documents are listed in the continuation of box C.	X Patent family members are listed in annex.		
"A" document defining the general state of the art which is not considered to be of particular relevance  "E" earlier document but published on or after the international filing date  "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)  "O" document referring to an oral disclosure, use, exhibition or other means  "P" document published prior to the international filing date but later than the priority date claimed	<ul> <li>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</li> <li>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</li> <li>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</li> <li>"&amp;" document member of the same patent family</li> </ul>		
Date of the actual completion of the international search	Date of mailing of the international search report		
28 May 1998	2 7 08 1998		
Name and mailing address of the ISA  European Patent Office, P.B. 5818 Patentlaan 2  NL - 2280 HV Rijswijk  Tel. (+31-70) 340-2040, Tx. 31 651 epo ni,  Fax: (+31-70) 340-3016	Authorized officer  MONTERO LOPEZ B.		

Internat I Application No PCT/US 98/03002

C/Continue	ntion) DOCUMENTS CONSIDERED TO BE RELEVANT	PCT/US 98/03002
Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Р,Х	WO 97 11180 A (CORIXA CORPORATION) 27 March 1997  see page 2, line 20 - page 3, line 37 see page 6, line 16 - page 7, line 20 see page 9, line 4 - line 19 see page 12, line 20 - page 14, line 17 see page 15, line 3 - page 16, line 26 see page 17, line 8 - line 30; claims; examples 1,7,8	1-6,25, 27-29, 31, 35-40, 42-55
	•	
	^	

Inter. .onal application No.

PCT/US 98/03002

B x I Obs rvations where certain claims wer if und uns archabl (Continual in the intermediate)
This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
- C
<ol> <li>Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:</li> </ol>
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This International Searching Authority found multiple inventions in this international application, as follows:
see continuation sheet
As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. X No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
see continuation sheet, subject 1
Remark on Protest  The additional search fees were accompanied by the applicant's protest.
No protest accompanied the payment of additional search fees.

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-6, and partially 25, 27-29, 31, 33, 35-40, 42-55

Polypeptide comprising an immunogenic portion of a Leishmania antigen having SEQ ID NO:2, variant thereof, DNA molecule encoding it, expression vector and host cell comprising the same, pharmaceutical compositions and vaccines comprising the polypeptide, vaccine comprising the DNA molecule, diagnostic kit

2. Claims: 7-12, and partially 25, 27-29, 31, 33, 35-40, 42-55

Polypeptide comprising an immunogenic portion of a Leishmania antigen having SEQ ID NO:4, variant thereof, DNA molecule encoding it, expression vector and host cell comprising the same, pharmaceutical compositions and vaccines comprising the polypeptide, vaccine comprising the DNA molecule, diagnostic kit

3. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

Polypeptide comprising an immunogenic portion of a Leishmania antigen comprising SEQ ID NO:22, variant thereof, DNA molecule encoding it, expression vector and host cell comprising the same, pharmaceutical compositions and vaccines comprising the polypeptide, vaccine comprising the DNA molecule, diagnostic kit

- 4. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:24
- 5. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55 Idem as subject 3, for SEQ ID NO:26
- 6. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55 Idem as subject 3, for SEQ ID NO:36
- 7. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:37
- 8. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

Idem as subject 3, for SEQ ID NO:38

- 9. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:41
- 10. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55
  Idem as subject 3, for SEQ ID NO:49
- 11. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:50
- 12. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:51
- 13. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:52
- 14. Claims: Partially 13, 16-19, 25, 27-31, 33-40, 42-55

  Idem as subject 3, for SEQ ID NO:53
- 15. Claims: Partially 13, 16-19, 25, 27-29, 31, 33, 35-40, 42-55

Idem as subject 3, for SEQ ID NO:82

16. Claims: 14, 15, and partially 16, 25, 27-29, 31, 33, 35-40, 42-55

Antigenic epitope of Leishmania antigen comprising SEQ ID NO:43, polypeptide comprising at least two such contiguous epitopes, DNA molecule encoding it, pharmaceutical compositions and vaccines comprising the polypeptide, vaccine comprising the DNA molecule, diagnostic kit

17. Claims: 20-24, 26, 32, 41 and partially 27-30, 33-39, 42-55

Polypeptide comprising an immunogenic portion of a

Leishmania antigen comprising SEQ ID NO:20, variant thereof, DNA molecule encoding it, expression vector and host cell comprising the same, pharmaceutical compositions and vaccines comprising the polypeptide, vaccine comprising the DNA molecule, diagnostic kit

18. Claims: Partially 56-62

Pharmaceutical compositions and vaccines comprising a polypeptide comprising an immunogenic portion of a Leishmania antigen comprising SEQ ID NO:39 or a variant thereof

19. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:42

20. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:55

21. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:61

22. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:62

23. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:80

24. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:81

25. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:83

26. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:84

27. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:85

28. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:86

29. Claims: Partially 56-62

Idem as subject 18, for SEQ ID NO:87

Information on patent family members

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			EP	0854924 A	29-07-1998

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